

5-Hydroxytryptamine Receptors

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I. Introduction

THE first experimental evidence suggesting the existence of an endogenous vasoconstrictor substance was presented by Stevens and Lee (1884) and by Brodie (1900), who showed that the serum obtained after blood clotting increased vascular tonus. Vialli and Erspamer (1933) observed the presence of "enteramine" in the gut, and Rapport et al. (1947) reported the existence of a vasotonic substance in the serum, which was, therefore, called "serotonin." The latter substance was identified as 5-HT* (Rapport et al., 1948; Rapport, 1949) and was

* Abbreviations: 5-HT, 5-hydroxytryptamine; 5-BT, 5-benzyloxytryptamine; 5-CT, 5-carboxamidotryptamine; 5-MeO-T, 5-methoxytryptamine; LSD, lysergic diethylamide acid; TFMPP, 1-(3-trifluoromethylphenyl)piperazine; 8-OH-DPAT, 8-hydroxy-2-(di-*n*-propylamino)tetraline; cAMP, cyclic adenosine monophosphate; GMP, guanosine monophosphate; 5,7-DHT, 5,7-dihydroxytryptamine; IP₃, inositol triphosphate; G_s, stimulatory G protein; G_i, inhibitory G protein; GABA, γ -aminobutyric acid; GTP, guanosine-5-triphosphate; GTP γ -S, guanosine-5-O-thiotriphosphate; DOI, 1-(2,5-dimethoxy-4-iodophenyl)-2-aminopropane; TCA, tricyclic antidepressant; ACh, acetylcholine; mCPP, 1-(3-chlorophenyl)piperazine; pCPA, *p*-chlorophenylalanine; REM, rapid eye movement (sleep); CNS, central nervous system; DOB, 1-(2,5-dimethoxy-4-bromophenyl)-2-aminopropane; cDNA, complementary DNA; mRNA, messenger RNA; DOM, 1-(2,5-dimethoxy-4-methylphenyl)-2-aminopropane; BIMU 1, (endo-N-8-methyl-8-azabicyclo[3.2.1]oct-3-yl)-2,3-dihydro-3-ethyl-2-oxo-¹H-benzimidazol-1-carboxamide hydrochloride; BIMU 8, (endo-N-8-methyl-8-azabicyclo[3.2.1]oct-3-yl)-2,3-dihydro-3-iso-propyl-2-oxo-¹H-benzimidazol-1-carboxamide hydrochloride; BMY 7378, 8-[2-[4-(2-methoxyphenyl)-1-piperazinyl]ethyl-8-azapiprol[4.5]-decane-7,9-dione dihydrochloride; BRL 20627, (2- α ,6 β ,9 α)-(±)-4-amino-5-chloro-2-methoxy-N-(octahydro-6-methyl-2-H-quinolizin)-benzamide; BRL 24924, endo-2-methoxy-4-amino-5-chloro-N-(1-azabicyclo-[3.3.1]-non-4-yl)benzamide mono-hydrochloride; BRL 43694, (granisetron), endo-N-(9-methyl-9-azabicyclo [3,3,1]non-3-yl)-1-methyl-imidazole-3-carboxamide; BRL 47204, 1-methyl-N-(endo-9-methyl-9-azabicyclo-[3,2,1]non-3-yl)indol-3-yl carboxylic acid amide; CGS 12066B, 7-trifluoromethyl-4-(4-methyl-1-piperazinyl)-pyrrolo-(1,2-*a*) quinoxaline 1:2 maleate salt; cisapride, *cis*-4-amino-5-chloro-N-[1-[3-(4-fluoro-phenoxy) propyl]-3-methoxy-4-piperidinyl]-2-methoxybenzamide; compound No. 1, N,N'-bis[3-(4-indolyloxy)-2-hydroxy-propyl]-(Z)-1,8-diamino-*p*-methane; compound No. 2, N⁸-[3-(4-indolyloxy)-2-hydroxypropyl]-N¹-(propidoyl)-(z)-1,8-diamino-*p*-methane-granisetron, endo-N-(methyl-9-azabicyclo-[3.3.1]-non-3-yl)-1-methyl-indazol-3-carboxamide (or BRL43694); CP-93,129, 3-(1,2,5,6-tetrahydro-4-pyridyl)pyrrolo[2,3-*b*]-

rapidly shown to be identical with enteramine. Hamlin and Fisher (1951) were able to synthesize it. Twarog and Page (1953) demonstrated its presence in mammalian brain, and Amin et al. (1954) showed that its distribution

pyril-5-one; CP-96,501, 3-(1,2,5,6-tetrahydro-4-pyridyl)5-*n*-propoxyindol; GR 65630, 3-(5-methyl-¹H-imidazol-4-yl)-1-(1-methyl-¹H-indol-3-yl)-1-propanone; GR 67330, 1,2,3,9-tetrahydro-9-methyl-3-[(5-methyl-¹H-imidazol-4-yl)-methyl]-⁴H-carbazol-4-one; GR 38032, (±)-1,2,3,9-tetrahydro-9-methyl-3[(2-methyl-¹H-imidazol-1-yl)methyl]-⁴H-carbazol-4-one; ICI 169-369, 2-(2-dimethylaminoethylthio)-3-phenylquinoline; ICS 205-930, (3- α -tropanyl)¹H-indole-3-carboxylic acid ester; ICYP, iodocyanopindolol; IMAO, inhibitors of monoaminooxidases; LY 165-163, *p*-aminophenyl ethyl-*m*-trifluoromethyl phenyl piperazine (or PAPP); LY 211-000, (8-methyl-8-azabicyclo[3.2.1]oct-3-yl)-1H-indazole-3-carboxylic acid ester; LY 258-458, N-(8-methyl-8-azabicyclo[3.2.1]oct-3-yl)-¹H-indazole-3-carboxamide; LY 278-584, 1-methyl-N-(8-methyl-8-azabicyclo[3.2.1]oct-3-yl)-¹H-indazole-3-carboxamide; MDL 72222, 1- α H-3 α -5 α H-tropan-3-yl-3,5-dichlorobenzoate; MDL 72832, 8-[4-[(1,4-benzodioxan-2-ylmethyl)amino]butyl azapiro [4.5]decane-7,9-dione; MDL 73005EF, 8-[2-(2,3-dihydro-1,4-benzodioxin-2-yl)Me-thylamino]-8-azaspiro [4.5]decane-7,9-dione methyl sulfonate; MK 212, 6-chloro-2-(1-piperazinyl)pyrazine; NAN-190, (1-(2-methoxyphenyl)-4-(4 (2phtalimido) butyl)piperazine; ondansetron, (1,2,3,9-tetrahydro-9-methyl-3[(2-methyl-¹H-imidazol-1-yl)-methyl]-4-one (or GR 38032); PAPP, 1[2-(4-aminophenyl)ethyl]-4-(3trifluoro methylphenyl) piperazine (or LY 165-163); PKC, protein kinase C; PLC, phospholipase C; QICS 205-930, quaternized-(3- α -tropanyl)-¹H-indole-3-carboxylic acid ester; quipazine, 2-(1-piperazinyl)quinoline; [³H]rauwolscine, (2-methoxy-1,4-[6,7³H]benzodioxan-2-yl)-2-imidazolin (or RX821002); RP 62203, 2-(3-[4-fluorophenyl]piperazinyl)-propyl naphtho-[1,8-*c,d*]-isothiazole-1,1-dioxide; RS 30199-193, 5-chloro-2-methyl-1,2,3,4,8,9,10,10a-octahydronaphth-[1.8-*cd*]-azepine; RU 24969, 5-methoxy-3(1,2,3,6-tetrahydro-4-pyridinyl)¹H indole succinate; SCH 23390, 7-chloro-8-hydroxy-1-phenyl-2,3,4,5-tetrahydro-¹H-3-benzazapine; SCH 23982, (+)-8-iodo-2,3,4,5-tetrahydro-3-methyl-5-phenyl-¹H-3-benzazepine-7-oid; S-CM-GT NH₂, serotonin-O-carboxymethyl-glycyl-tyrosinamide; S-14063, N-[1-(benzothiazol-2-yl)]-N-[4-[(methyl-2-ethoxy)-2-phenyl]methylamino]piperidine; S-14671, 4-[(thenoyl-2)aminoethyl]-1-(7-methoxynophyl piperazine); SDZ 206-557, 2-methoxy-4-amino-5-chloro-benzoic acid 2-(diethylamino)ethyl ester; SDZ 206-792, (N-deasmethyl-3- α -homotropanyl)-¹H-indole-3-carboxylic acid ester; SDZ 206-830, (3 α -homotropanyl)-1-methyl-5-fluoro-indole-3-carboxylic acid ester; SDZ(-)21009, 4[3-*tert*butylamino-2-hydroxypropoxyl-indol-2-carbonic acid-isopropylester SDZ 210-204, (-)-(1*R*,2*R*,4*S*)-¹H-indole-3-carboxylic acid-7-methyl-7-azabicyclo-[2.2.1]hept-2-yl-ester; SDZ 216-525, 4-[4-(1-1,3-trioxo-2H-1,2-benzisothiazol-2-yl)butyl]-1-piperazinyl]1H-indole-2-carboxylate; (S)-UH-301, (S)-5-fluoro-8-hydroxy-2-(di-*n*-propylamino) tetra-

was heterogeneous in dog brain. This led to the suggestion that 5-HT might act as a neurotransmitter in the CNS (Bogdanski et al., 1956; Brodie and Shore, 1957).

Indeed, serotonin is involved in numerous physiological events. At the peripheral level, it affects smooth muscle fibers, causing constriction or relaxation, and thus exerts a major effect on the vascular bed and the digestive tract (Vanhoutte, 1985; Frohlich and Van Zwieten, 1987; Breckenridge et al., 1988; Hollenberg, 1988; Saxena et al., 1989; Saxena and Villalon, 1991; Parsons, 1991). Serotonin to some extent affects various functions of the CNS: sleep (Jouvet, 1962, 1967; Koella, 1988), thermoregulation (Feldberg and Myers, 1964; Jacob and Girault, 1979; Myers, 1981), learning and memory (Altman and Normile, 1986, 1988; McEntee and Crook, 1991), pain (Tenen, 1967; LeBars, 1988; Richardson, 1990), behavior such as aggressive (Sheard, 1969; Di Chiara et al., 1971), sex (Hoyland et al., 1970; Meyerson and Malmnas, 1978; Fernandez-Guasti et al., 1987; Gorzalka et al., 1990), and feeding (Blundell, 1977, 1984; Marazziti et al., 1988; Curzon, 1990), neuroendocrine regulations (Schneider and McCann, 1970; Charli et al., 1978; Tuomisto and Mannisto, 1985; Fuller, 1990; Van de Kar, 1991; Cowen et al., 1990), motor activity (Gershon and Baldessarini, 1980; Sternbach, 1991), and biological rhythms (Wesemann and Weiner, 1990).

Serotonin is thought to play a role in various types of pathological conditions: psychiatric disorders such as anxiety (Iversen, 1984; Gardner, 1985, 1986; Johnsson and File, 1986; Kahn et al., 1988; Nutt and George, 1990), depression (Schildkraut, 1965; Meltzer and Lowy, 1987; Cowen, 1988, 1990; Curzon, 1988; Kalus et al., 1989; Plaznik et al., 1989; Meltzer, 1990; Van Praag et al., 1990), aggressiveness (Pucilowski and Kostowski, 1983; Miczek and Donat, 1989; Olivier et al., 1989; Coccaro, 1989; Wetzler et al., 1991), panic (Eriksson, 1987; Sheeham et al., 1988), obsessive-compulsive disorders (Zohar and Insel, 1987; Zak et al., 1988; Charney et al., 1988; Insel et al., 1990), schizophrenia (Csernansky et al., 1992), suicidal behavior (Roy and Linnoila, 1988; Coccaro and Astill, 1990; Mann et al., 1990), and autism (Cook, 1990; Anderson et al., 1990); neurodegenerative disorders, such as Alzheimer's disease, Parkinsonism, and Huntington's chorea (Morgan et al., 1987; Cross, 1988, 1990; Cross et al., 1988), migraine (Fozard, 1990; Anthony, 1986; Humphrey et al., 1990), emesis (Fozard, 1987; Bunce et al., 1991), and alcoholism (Tollefson, 1989; Sellers et al., 1992).

Specific receptors mediate the physiopathological events triggered.

line; sumatriptan, 3-(2-dimethylamino)ethyl-N-methyl-¹H-indole-5-methane sulfonamide (or GR 43175); tandospirone, (3 α ,4 β ,7 β ,7 α)-hexahydro-2-(4-(4-(2-pyrimidinyl)-1-piperazinyl)-butyl)-4,7-methano-¹H-isoindole-1,3-(²H)-dione dihydrogen citrate (or SM-3997); WB 4101, 2-(2,6-dimethoxyphenoxyethyl)aminomethyl-1,4-benzodioxane; zacopride, 4-amino-N-(1-aza-bicyclo[2.2.2]oct-3-yl)-5-chloro-2-methoxybenzamide.

II. Classification of Serotonin Receptors

Evidence for the existence of serotonergic receptors was first presented by Gaddum and Picarelli (1957), who experimented with the isolated guinea pig ileum. They described two types of receptors controlling muscle contraction: D receptors, for those blocked by dibenzylamine, and M receptors, for those blocked by morphine. Receptor sites were tentatively observed by Marchbanks (1966, 1967), using [³H]5-HT, and by Farrow and van Vunakis (1972, 1973), who showed that [³H]LSD binding could be displaced by 5-HT. The earliest evidence for a selective high-affinity and saturable binding of [³H]5-HT was presented by Bennett and Snyder (1975, 1976) and Fillion et al. (1976). That [³H]spiroperidol could also label a serotonergic receptor (Leysen et al., 1978) led Peroutka and Snyder (1979) to propose the existence of two classes of serotonergic receptors, 5-HT₁ (labeled with high affinity by [³H]5-HT) and 5-HT₂ (labeled by [³H]spiperone). In their report, Peroutka and Snyder provided the initial evidence that the 5-HT₁ receptors might be heterogeneous, because the inhibition of [³H]5-HT binding, caused by spiroperidol, was biphasic. Pedigo et al. (1981) confirmed these results and proposed the existence of two sites labeled by [³H]5-HT: 5-HT_{1A} (exhibiting a high affinity for spiroperidol) and 5-HT_{1B} (a low affinity for spiroperidol). A third [³H]5-HT-binding site, designated 5-HT_{1C}, was proposed on the basis of the high-affinity displacement of [³H]5-HT by mesulergine (Pazos et al., 1985b). Functional studies attempted to attribute a physiological role to these binding sites (Fozard and Kilbinger, 1985; Engel et al., 1985).

It became necessary to reconcile the different nomenclatures and classify the serotonergic receptors by establishing criteria that also respected receptor definition (e.g., binding sites having physiological properties). A group of scientists (Bradley et al., 1986) proposed three major serotonergic receptor classes: "5-HT₁-like," 5-HT₂, and 5-HT₃.

The term "5-HT₁-like" was proposed for the heterogeneous group of receptors having a high affinity for 5-HT and 5-CT, was antagonized by methiothepin and methysergide, and was not fully characterized as functional entities.

The term "5-HT₂" was proposed for the D receptors that had been described much earlier by Gaddum and Picarelli (1957). These receptors mediate a variety of peripheral actions of 5-HT and correspond to the cortical 5-HT₂-binding sites described by Peroutka and Snyder (1979) having a low affinity for 5-HT and a high affinity for particular serotonergic antagonists, such as ketanserin, methysergide, mianserin, and metergoline.

The 5-HT₃ receptors corresponded to the M receptors (Gaddum and Picarelli, 1957) present in peripheral neurons which mediate the depolarizing actions of 5-HT. They were characterized by their high affinity for cocaine derivatives (ICS 205-930 and MDL 72222). However,

the presence of specific binding sites in brain was not demonstrated at that time.

This classification was later slightly modified by Peroutka (1990) who incorporated new data, such as the existence of a novel 5-HT₁ subtype, called 5-HT_{1D} (Heuring and Peroutka, 1987) (fig. 1), the pharmacological and molecular similarities between 5-HT_{1C} and 5-HT₂ receptors (Hoyer, 1988; Hartig, 1989a), and the clear confirmation of the functional role of the 5-HT₁-like receptors. Peroutka proposed three main classes of receptors: 5-HT₁ (including 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1D} subtypes), 5-HT₂ (including 5-HT_{2A}, 5-HT_{2B}, and 5-HT_{1C} subtypes), and 5-HT₃. Peroutka proposed that the latter class was homogenous; the pharmacological differentiation of 5-HT₃ receptor subtypes previously presented by Richardson and Engel (1986) was interpreted as species and tissues differences.

Although useful, this classification, which was based on agonist and antagonist drug specificity, did not account for receptors such as 5-HT_{1nonA,nonB,nonC} which have a high affinity for 5-HT and a low affinity for 5-CT (Fayolle et al., 1988) or for 5-HT_{1P} receptors which have a high affinity for 5-HT and a pharmacological profile different from that of the other 5-HT₁ receptors (Mawe et al., 1986; Branchek et al., 1988). Moreover, this classification cannot distinguish receptors such as 5-HT_{1D α} and 5-HT_{1D β} which have similar pharmacological properties but are encoded by two different genes (Hartig et al., 1992). In contrast, the classification distinguishes between the 5-HT_{1B} and the 5-HT_{1D β} receptors based of their pharmacological properties, whereas the genes encoding these receptors are proposed as species homologues (Hartig et al., 1992).

Moreover, several potential receptors, characterized either by their binding or by their functional properties,

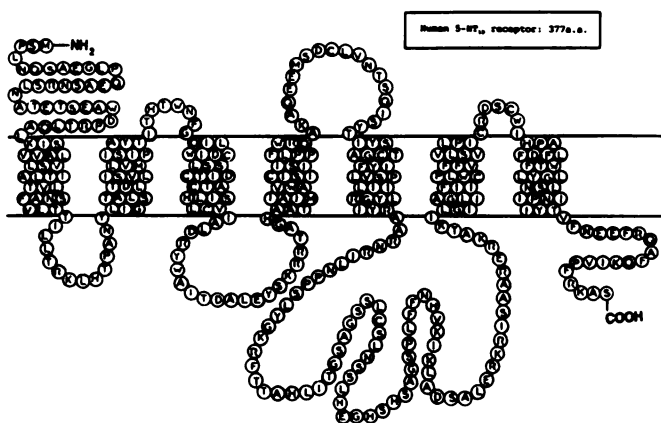


FIG. 1. Topographical model of the 5-HT_{1D} receptor. The amino acid sequence is deduced from the nucleotide sequence of the human gene (Hamblin and Metcalf, 1991). The horizontal lines correspond to the membrane separating extracellular (top) and intracellular (bottom) spaces. The receptor is a monomeric protein possessing seven transmembranal hydrophobic domains linked by three extracellular and three intracellular loops. The NH₂ terminal is extracellular, and the COOH terminal is intracellular.

have a specific pharmacological profile that does not allow one to classify them in any of the proposed categories. This is the case for the following binding sites: 5-HT_{1E} sites (high affinity for 5-HT and low affinity for 5-CT) (Leonhardt et al., 1989), 5-HT_{1S} sites (high affinity for 5-HT and not antagonized by methysergide) (Zelman et al., 1990; Zelman and Schwab, 1991), and 5-HT_{1R} sites (Xiong and Nelson, 1989). On the other hand, potential receptors that mediate 5-HT activities have a pharmacological profile different from that of the following known 5-HT receptors: 5-HT₄ receptors stimulating the adenylyl cyclase activity (Dumuis et al., 1988b; see Bockaert et al., 1992), the novel 5-HT receptors depolarizing neonatal rat motoneurons (Connell and Wallis, 1989), receptors enhancing a hyperpolarization-activated cationic current (I_h) (Pape and McCormick, 1989; Bobker and Williams, 1989), receptors inhibiting the release of endogenous aspartate (Maura et al., 1991), receptors coupled to cyclic GMP production in NG108-15 cells (Tohda et al., 1991a,b), and receptors mediating slow excitatory responses in neurons of the CA1 region of the hippocampus (Chaput et al., 1990).

Thus, since the initial observations of Gaddum and Picarelli (1957), great progress has been made, thanks to the successive discoveries of new ligands which have provided a better understanding of specific 5-HT receptors. However, because of the lack of specific ligands, several potential receptors are still awaiting appropriate classification. Results of molecular biological studies have provided new important perspectives concerning the existence of receptor subtypes. These data will contribute to complete and precise classification of the serotonergic receptors.

Hartig (1989a) proposed simply to classify the 5-HT receptors on the basis of their structural homology, which had been established by molecular biology, and their predominant transduction system. He distinguished serotonergic receptors, the activity of which are coupled to a G protein, from those directly linked to an ionic channel. Of the former, the 5-HT_{1A}, 5-HT_{1B}, 5-HT_{1D}, and 5-HT₄ receptors were associated with adenylyl cyclase, whereas the 5-HT_{1C} and 5-HT₂ receptors were associated with PLC.

Three receptors had been cloned by the time Hartig proposed this classification scheme (5-HT_{1A}, 5-HT_{1C}, and 5-HT₂); these were found to belong to the multigenic family of the G protein-related receptors having a monomeric structure (420 to 460 amino acids) with seven highly conserved transmembrane domains of 20 to 28 amino acids each; the recognition sites for the agonistic ligands are presumably included in these domains (Strader et al., 1989; Curtis et al., 1989). These structures have an extracellular amino-terminal domain containing glycosylation sites, an intracellular carboxy-terminal part which often possesses phosphorylation sites, and three extracellular and three intracellular loops. The

extramembranal domains (intra- and extracellular), are not highly conserved and thus appear to be more specific for each receptor than the intramembranal domain. The third intracellular loop is thought to involve a coupling with the G protein (Strader et al., 1989; Lechleiter et al., 1990) and may have phosphorylation sites. The COOH-terminal domain, particularly the part that is close to the membrane, also might be involved in coupling with the G protein (Strader et al., 1989; O'Dowd et al., 1989).

The recent findings from 5-HT receptor research have progressed so rapidly, especially in molecular biology, that the primary molecular structures of a number of receptors have been described, allowing one to propose a more precise classification (table 1). A definite classification of the receptors based on molecular biology and pharmacology likely will be rapidly established.

Finally, a classification analogous to that used for enzymes was proposed by Frazer et al. (1990). Thus, receptors can be classified by four numbers, the first referring to the activating ligand (e.g., 5-HT), the second to the superfamily (e.g., G protein linked), the third to the predominant second-messenger system (e.g., adenylyl cyclase), and the fourth to the primary amino acid sequence in chronological order of their publication. The merit of this classification is the ability to identify precisely the profile of the receptor following a simple examination of its "enzyme-like code" and then clearly underlining similarities and differences between receptors. However, the pharmacological specificity is not shown, and the fourth code number is not an intrinsic characteristic of the receptor.

The aim of this review is to present the main characteristics of the serotonergic receptors with regard to their molecular biological, pharmacological, and functional properties at the cellular as well as the integrated level in vivo. It should be pointed out that excellent exhaustive reviews of 5-HT receptors were recently published (Sanders-Bush, 1988a; Osborne and Hamon, 1988).

III. 5-HT₁ Receptors

These receptors are characterized by a high affinity (<10 nM) for 5-HT and 5-CT, a medium affinity (10 to 1000 nM) for methiothepin, methysergide, quipazine, and TFMPP, and a low affinity (>1000 nM) for ICS 205-930, MDL 72222, and ketanserin.

The artifactual nature of these receptors has often been proposed, because it was thought that high-affinity binding could not be a characteristic of a functional receptor (Fozard, 1983; Leysen, 1984a,b; Bradley et al., 1986). However, it is now clear that 5-HT₁ receptors belong to a large family of functional receptors and are likely to be involved in slow neuromodulatory phenomena.

In this review, three subclasses of 5-HT₁ receptors are considered: 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1D}; 5-HT_{1C}, according to the suggestion of Hartig (1989a), will be presented as a member of the 5-HT₂ class.

A. 5-HT_{1A} Receptor Subtype

Pedigo et al. (1981) showed that spiperone can be used to discriminate between two different subclasses of 5-HT₁ sites: one having a high affinity for spiperone (5-HT_{1A}) and the other one a low affinity (5-HT_{1B}). The ratio of these two sites in the hippocampus differs from that in the cortex.

1. *Selective radioligands and pharmacological properties.* The investigation of 5-HT_{1A} receptors became possible when Hjorth et al. (1982) discovered a tetraline derivative, 8-OH-DPAT, and Gozlan et al. (1983) demonstrated that this drug, labeled with tritium (³H]8-OH-DPAT), bound selectively to the 5-HT_{1A} site. The following ligands have since been used for studies of receptor binding: [³H]ipsapirone (³H]TVXQ 7821) (Dompert et al., 1985; Glaser and Traber, 1985), [³H]WB 4101 (Norman et al., 1985), [³H]PAPP (Ramson et al., 1986), [³H]5-MeO-DPAT (Cossery et al., 1987), [³H]spiroxatrine (Herrick-Davis and Titeler, 1988a), ¹²⁵I-BH-8-MeON-

TABLE 1
Serotonergic receptor classification

Receptor type	Receptor family	Transduction system (predominant)	Characterized receptors	Potential receptors
G protein-related receptors	5-HT ₁	Adenylyl cyclase (cAMP production)	5-HT _{1A}	5-HT _{1A} subtypes ?
			5-HT _{1D}	5-HT _{1R}
7 transmembrane domains	5-HT ₂	Phospholipase C (IP ₃ /DAG production)	5-HT _{1B}	5-HT _{1E}
			5-HT ₂	5-HT _{1nonA, nonB, nonC}
			5-HT _{1C}	5-HT ₄
Ionic channel receptors	5-HT ₃	Monovalent cations flux		5-HT _{3-LIKE}
				5-HT _{3A}
				5-HT _{3B}
				5-HT _{3C}

PAT (Gozlan et al., 1988), [³H]buspirone (Brüning et al., 1989), [³H]NAN-190 (Rydelek-Fitzgerald et al., 1990), and [³H]tandospirone (Tanaka et al., 1991). Recently, [³H]rauwolscine ([³H]RX821002), a marker for α_2 -adrenoceptors, was presented as a potential 5-HT_{1A} marker in the human frontal cortex (De Vos et al., 1991).

The highly selective prototypical 5-HT_{1A} agonist, 8-OH-DPAT (an aminotetralin), remains unique in its combination of both high efficacy and striking potency in vivo. Numerous 5-HT_{1A} agonists are presently available and exhibit potential therapeutic value, notably for the treatment of anxiety and depression; however, most of these drugs are partial agonists which also have antagonistic properties: pyrimidinylpiperazines (buspirone, ipsapirone, gepirone, tandospirone), naphthylpiperazines (S-14671), benzodioxanes (MDL 72832, spiroxatrine, MDL 73005EF, WB 4101), heterobicyclic arylpiperazines (flesinoxan), and phenylpiperazines (BMY 7378, NAN-190). Other agonistic drugs that interact potently with the 5-HT_{1A} receptor, but are nonselective for 5-HT receptor subtypes, are ergot derivatives (*d*-LSD, metergoline) and various indoles (5-HT, 5-CT, N,N-dipropyl-5-CT, and RU 24969).

The 5-HT_{1A} receptor antagonists currently used are not selective; spiperone and spiroxatrine also bind with a high affinity to 5-HT₂ and dopamine sites, propranolol and pindolol bind to 5-HT_{1B} and β -adrenoceptor sites, and methiothepin is nonselective with respect to different 5-HT receptors and binds with a high affinity to histaminergic and dopaminergic sites. In addition, MDL 73005EF (Hibert et al., 1988) and the phenylpiperazine derivatives BMY 7378 and NAN-190, which first appeared to be 5-HT_{1A} antagonists (Chaput and de Montigny, 1988a; Glennon et al., 1988, 1989), have been shown to be mixed partial agonists/antagonists (Yocca et al., 1987). Presently, four drugs are proposed to be selective and potent 5-HT_{1A} antagonists: N¹-(bromoacetyl)-[N⁶]-3-(4-indolyloxy)-2-hydroxypropyl]-*(Z)*-1,8-diamino-*p*-methane, a pindolol derivative quite selective of 5-HT_{1A} receptors (an order of magnitude less potent at the β -adrenoceptors) (Liau et al., 1991); S-14063, devoid of β -adrenoceptor-blocking properties (Dabiré et al., 1991), SDZ 216-525, proposed as a potent, selective, and silent 5-HT_{1A} receptor antagonist, both in vitro and in vivo (Hoyer et al., 1992); and a novel 8-OH-DPAT analog, (*S*)-UH-301, able to antagonize completely several (*R*)-8-OH-DPAT-induced effects (Johansson et al., 1991; Björk et al., 1991). The confirmation of the antagonistic nature of these drugs and the development of new 5-HT_{1A}-specific antagonists are urgently needed to really understand the physiological and functional role(s) of 5-HT_{1A} receptors.

The differences in intrinsic activity and potency of the compounds acting at 5-HT_{1A} receptors depend on the brain region and on the functional model used. 8-OH-DPAT, ipsapirone, gepirone, buspirone, MDL 73005EF,

BMY 7378, and NAN-190, in electrophysiological and behavioral studies, act as full agonists at raphe 5-HT_{1A} receptors and as antagonists at hippocampal receptors (Smith and Peroutka, 1986; Martin and Mason, 1987; Sprouse and Aghajanian, 1988; Moser et al., 1990; Gartside et al., 1990; Yocca, 1990; Van den Hooff and Galvan, 1991) (table 2). 8-OH-DPAT, buspirone, and ipsapirone act as full agonists in inhibiting the cAMP production in hippocampus, whereas they are either partial agonists (8-OH-DPAT) or competitive antagonists (buspirone, ipsapirone) in cortex (Dumuis et al., 1988c). Moreover, in the hippocampus, 8-OH-DPAT produces a greater reduction in amplitude of population spikes in the CA1 area than in the dentate gyrus (Klancnik et al., 1991). Intrinsic activity values of 5-HT_{1A} agonists also depend on the test model; in the hippocampus, 8-OH-DPAT, ipsapirone, and gepirone are full agonists in inhibiting adenylyl cyclase activity (De Vivo and Maayani, 1986), whereas they are partial agonists or even antagonists on the hyperpolarizing response to 5-HT (Segal et al., 1989); NAN-190 antagonizes the 8-OH-DPAT-induced syndrome, whereas it was shown to be an agonist inhibiting the release of 5-HT in microdialysis studies (Hjorth and Sharp, 1990).

Surprisingly, the pharmacological properties of the 5-HT_{1A} recognition sites in raphe, hippocampus, and cortex are similar. Several mechanisms were proposed to explain the variations of the intrinsic activities of 5-HT_{1A} ligands. One hypothesis is that there are regional differences in 5-HT_{1A} spare receptor number. Indeed, a large receptor reserve exists in the raphe nucleus (Meller et al., 1990) but not in the hippocampus (Yocca et al., 1990a). Then, it would be expected that partial agonists elicit a full agonistic response in the raphe, whereas in the hippocampus, the partial agonist activity of the compound is insufficient to evoke an agonistic response and will result in an antagonist effect (Van den Hooff and Galvan, 1991). However, the fact that partial agonists such as buspirone or ipsapirone are full agonists in the hippocampus, which is devoid of spare receptors (De Vivo and Maayani, 1986), suggests that the mechanism might be more complex. Indeed, the intrinsic activity of agonists is dependent not only on ligand binding to 5-HT_{1A} receptor but also depends on the formation of the ligand-receptor-G protein ternary complex. The fact that compounds acting at receptors with similar recognition domains may have different efficacies suggests the existence of differences in postreceptor signal transduction processes (different G_i proteins, different coupling, or stoichiometry between receptor and G_i proteins) (Yocca and Maayani, 1990). Finally, another hypothesis proposes the existence of several types of 5-HT_{1A} receptors (Dumuis et al., 1988c; Sprouse et al., 1990).

Several recent findings have supplied additional information concerning the mechanism of action of agonist/antagonist 5-HT_{1A} drugs. Boddeke et al. (1992), compar-

TABLE 2
Affinity values of drugs for 5-HT_{1A} recognition sites in mammalian brain membranes

Drug	Affinity	Reference	Drug	Affinity	Reference
Very high affinity ($K_i < 10$ nM)			High affinity ($K_i = 10-100$ nM)		
5-CT	0.21	Hoyer et al., 1985b	Bufotenine	5.0	Peroutka, 1986
	0.31	Harrington et al., 1991		7.7	Harrington et al., 1991
	0.316	Hoyer and Schoeffter, 1991		21.4	Gozlan et al., 1983
	2.13	Engel et al., 1986	PAPP	6.16	Schoeffter and Hoyer, 1989a
			SDZ(-)21009	5.1	Harrington et al., 1991
DP-5-CT	0.28	Van Wijngaarden et al., 1990		7.4	Hoyer et al., 1985b
	0.316	Hoyer and Schoeffter, 1991		10	Hoyer and Schoeffter, 1991
MDL 72832	0.79	Van Wijngaarden et al., 1990; Hoyer and Schoeffter, 1991	Spiroxitrine	7.94	Hoyer and Schoeffter, 1991
8-OH-DPAT	0.4	Dompert et al., 1985	Metergoline	6	Hoyer et al., 1985b
	1	Peroutka, 1986; Harrington et al., 1991		7.9	Hoyer and Schoeffter, 1991
	1.86	Engel et al., 1986; Hoyer and Schoeffter, 1991		13	Peroutka, 1986
	2.45	Van Wijngaarden et al., 1990		34	Gozlan et al., 1983
	3	Gozlan et al., 1983			
DHE Compound No. 1	1.2	Hoyer and Schoeffter, 1991	Buspirone	14	Dompert et al., 1985
No. 2	1	Liau et al., 1991; Harrington et al., 1991		14.8	Van Wijngaarden et al., 1990
	2.2	Liau et al., 1991; Harrington et al., 1991	(-)Pindolol	15	Peroutka, 1986
Flesinoxan	1.5	Van Wijngaarden et al., 1990		25	Hoyer and Schoeffter, 1991
	0.31	Dompert et al., 1985		29.5	Gozlan et al., 1983
	2.2	Harrington et al., 1991		19	Hoyer and Schoeffter, 1991
	2.57	Hoyer and Schoeffter, 1991		23.4	Hoyer et al., 1985b
	3.0	Hall et al., 1985	Methysergide	25	Peroutka, 1986; Hoyer and Schoeffter, 1991
d-LSD	3.9	Peroutka, 1986		40	Dompert et al., 1985
	0.2	Norman et al., 1985		70.8	Gozlan et al., 1983
	0.79	Hoyer and Schoeffter, 1991		23	Dompert et al., 1985
	1.8	Harrington et al., 1991		41.7	Hoyer et al., 1985b
	3.8	Hall et al., 1985		48	Hall et al., 1985
Lisuride	3.9	Peroutka, 1986		63	Hoyer and Schoeffter, 1991
	0.2	Norman et al., 1985		74	Norman et al., 1985
	0.79	Hoyer and Schoeffter, 1991		320	Wander et al., 1987
	1.8	Harrington et al., 1991		25.1	Hoyer and Schoeffter, 1991
	3.8	Hall et al., 1985	Bromocriptine	38	Peroutka, 1986
NAN-190	1.3	Van Wijngaarden et al., 1990	<i>l</i> -LSD	42-50	Sijbema et al., 1990
	1.9	Harrington et al., 1991	Eltoprazine	53	Engel et al., 1986
	2.2	Peroutka, 1986	(+)CYP	64.5	Schoeffter and Hoyer, 1989a
	3.1	Engel et al., 1986; Hoyer and Schoeffter, 1991	CGS 12066B	63.0	Hoyer and Schoeffter, 1991
	3.3	Schlicker et al., 1989	Urapidil	117	Schlicker et al., 1989
5-HT	4.3	Dompert et al., 1985		68.5	Doble, 1990
	6.7	Norman et al., 1985	RP 62203	79.4	Hoyer and Schoeffter, 1991
	6.8	Gozlan et al., 1983	Gepirone	72	Hoyer et al., 1985b
	2.2	Dompert et al., 1985	Methiothepin	79	Hoyer and Schoeffter, 1991
	2.9	Peroutka, 1986		89.1	Gozlan et al., 1983
	5.9	Van Wijngaarden et al., 1990			
	7.9	Harrington et al., 1991			
Ipsapirone (TVXQ 7821)	19	Hoyer and Schoeffter, 1991			
	2.0	Peroutka, 1986			
	3.8	Norman et al., 1985			
	5.6	Harrington et al., 1991			
	12.5	Hoyer and Schoeffter, 1991			
WB 4101	2.5	Peroutka, 1986			
	4.3	Harrington et al., 1991			
	5.1	Hoyer et al., 1985b			
	7.9	Hoyer and Schoeffter, 1991			
	9.8	Gozlan et al., 1983			
(±)CYP	2.29	Engel et al., 1986			
	5.37	Hoyer and Schoeffter, 1991			
	4.8	Harrington et al., 1991			
	2.39	Van Wijngaarden et al., 1990			
	2.4	Yocca et al., 1987			
BMJ 7378	2.5	Peroutka, 1986			
	9.12	Schlicker et al., 1989			
	3.4	Harrington et al., 1991			
5-MeO-DMT	4.1	Hoyer et al., 1985b			
	3.9	Hoyer and Schoeffter, 1991			
	3.98	Hoyer and Schoeffter, 1991			
(±)ICYP					
Ergotamine					
α -Dihydroergocryptine					

TABLE 2—Continued

Drug	Affinity	Reference	Drug	Affinity	Reference
GR 43175	794	Hoyer and Schoeffter, 1991	Quipazine	780	Peroutka, 1986
<i>d</i> -Butaclamol	800	Wander et al., 1987		3,231	Schoeffter and Hoyer, 1989a
Mianserin	800	Peroutka, 1986		3,310	Hoyer et al., 1985b
	933	Schlicker et al., 1989		3,390	Gozlan et al., 1983
	1,000	Hoyer and Schoeffter, 1991	Ketanserin	1,170	Dompert et al., 1985
	1,150	Hoyer et al., 1985b		1,258	Hoyer and Schoeffter, 1991
Amitriptyline	960	Peroutka, 1986		1,290	Norman et al., 1985
	Very low affinity ($K_i > 1,000$ nM)			1,910	Hoyer et al., 1985b
Piremperone	1,258	Hoyer and Schoeffter, 1991		4,170	Gozlan et al., 1983
	1,700	Hoyer et al., 1985b	Haloperidol	2,800	Wander et al., 1987
	2,500	Peroutka, 1986	Chlorpromazine	3,200	Wander et al., 1987
2-Me-5-HT	1,698	Van Wijngaarden et al., 1990	Ritanserin	831	Van Wijngaarden et al., 1990
Clozapine	1,800	Wander et al., 1987	DOI	6,309	Hoyer and Schoeffter, 1991
Tryptamine	3,720	Gozlan et al., 1983	Imipramine	6,938	Van Wijngaarden et al., 1990
				21,000	Gozlan et al., 1983

ing the effects of a variety of 5-HT_{1A}-selective drugs on the PLC/calcium cascade in H7 and H6 cells (H6 cells express five to six times more 5-HT_{1A} receptors than H7 cells), demonstrated that the intrinsic activity of a ligand can vary widely, depending on tissue variables such as receptor density or presence of second-messenger systems. They concluded that the differences are probably related to both the degree of receptor reserve and the receptor-effector coupling. Moreover, Radja et al. (1992) provided experimental evidence suggesting the existence of different 5-HT_{1A} receptor subtypes in the CA1 area and in the dentate gyrus of rat brain.

Another parameter that affects the intrinsic activity of the 5-HT_{1A} drugs involves their stereoisomeric properties. The partial agonist properties of several racemic mixtures of 5-HT_{1A} drugs presumably reside in the fact that the two enantiomers have different intrinsic activities (i.e., the *R*-enantiomer of 8-OH-DPAT acts as a full agonist in inhibiting the cAMP production, whereas the *S*-enantiomer acts as a partial agonist) (Cornfield et al., 1991). Screening the 5-HT_{1A} activity of the enantiomers of 8-OH-DPAT analogs can ultimately be used to differentiate full agonists from partial agonists or antagonists, thereby establishing the importance of the orientation of the aminotetralin nucleus relative to the substitution in determining intrinsic activity. The three-dimensional model described recently by Mellin et al. (1991) proposed a flexible pharmacophore based on the relative position of the aromatic nucleus and a nitrogen-dummy atom vector of various DPAT derivatives; this was independent of the orientation of the *N*-alkyl group. These studies will favor the design of new 5-HT_{1A} agonists.

2. *Regional distribution within the brain.* The distribution of 5-HT_{1A} receptors within the brain has been studied in many animal species, including rat (Marcinkiewicz et al., 1984; Pazos and Palacios, 1985), mouse, guinea pig, calf, cat (Waeber et al., 1989a), pig (Hoyer et al., 1985b), monkey (Köhler et al., 1986), and humans (Hoyer et al., 1986a; Pazos et al., 1987a). A similar receptor distribution was reported in all of these species.

Thus, the highest density was observed in the limbic system: hippocampus (dentate gyrus and CA1), septum, amygdala, and the cortical limbic area (entorhinal cortex). The preferential distribution of 5-HT_{1A} receptors in limbic areas is consistent with their involvement in the control of mood and anxiety. These receptors are generally sparse in brain areas rich in other 5-HT₁ subtypes (5-HT_{1B/1D}).

Interestingly, 5-HT_{1A} receptors are highly localized in raphe dorsal and median nuclei that contain serotonergic cell bodies. In this area, the 5-HT_{1A} receptors act as somatodendritic autoreceptors regulating the firing of serotonergic neurons. These autoreceptors are involved in neural mechanisms of spontaneous locomotor activity (median raphe) and in the mediation of thermoregulation (dorsal raphe), hyperphagia (dorsal raphe), and male rat sexual behavior (dorsal and median raphe) (see Hillegaart, 1991).

3. *Cellular localization.* The 5-HT_{1A} receptors located in raphe nuclei are presynaptic and correspond to somatodendritic sites present on serotonergic neurons (autoreceptors). This pattern was shown by Weissman-Nanopoulos et al. (1985) and Vergé et al. (1985, 1986) who observed a marked decrease of the receptor density after provoking selective serotonergic lesions by using 5,7-DHT. However, the decrease was only 50% which strongly suggests that some of the receptors are not located on serotonergic neurons.

5-HT_{1A} receptors also are localized postsynaptically to the 5-HT nerve terminals (e.g., hippocampal pyramidal cells). This was shown by Hall et al. (1985) and Vergé et al. (1986) who observed no decrease in these sites after treatment with 5,7-DHT but reported a 50% decrease after kainic acid lesions were produced in this area. The latter observation suggests that, in the hippocampus, the 5-HT_{1A} receptors are somatodendritic and also are located at terminals of nonserotonergic neurons or on glial cells.

4. *Molecular structure.* The cloning of the 5-HT_{1A} receptor made it possible to determine precisely the

molecular structure of the receptor protein. Kobilka et al. (1987), studying human β -adrenoceptors, isolated a cloned gene (G21) located on chromosome 5 which was later identified by Fargin et al. (1988) as that encoding the 5-HT_{1A} receptor. The efficiency of β -adrenergic antagonists at 5-HT_{1A} receptors (i.e., propranolol, pindolol) can be explained by the close homology of the two receptors. Albert et al. (1990) and Fujiwara et al. (1990) cloned the 5-HT_{1A} receptor in the rat; it was found to have an 88% homology with the human receptor. Surprisingly, there is greater similarity between the 5-HT_{1A} and β -adrenoceptor than between 5-HT_{1A} and the serotonergic receptors 5-HT_{1C} (18 and 40% identity for total structure and transmembranal domains, respectively) and 5-HT₂ (19 and 41% identity, respectively) (Hartig et al., 1990a; Harrisson et al., 1991). Genes coding for proteins resembling 5-HT_{1A} receptors have been cloned from *Drosophila* (Witz et al., 1990) and dog (RDC4 gene) (Libert et al., 1989, 1991). However, a partial sequence of RDC4 was recently used to isolate the human gene coding for a different receptor subtype (5-HT_{1D}) (Hamblin and Metcalf, 1991).

The 5-HT_{1A} gene has been shown to be located on chromosome 5 in humans, at bands 5q11.2 to q13, on the proximal side of D5S76 (Kobilka et al., 1987; Melmer et al., 1991), and on chromosome 13 in mouse (Oakey et al., 1991); a conserved linkage has been described for these chromosomes in the two species (Maurer et al., 1985).

The 5-HT_{1A} protein receptor consists of a monomeric chain (422 amino acids in rat, 421 amino acids in human) characterized by a short carboxy-terminal domain (16 amino acids) and a long third intracellular loop (128 amino acids); in contrast, the β_2 -adrenoceptor contains a long carboxy-terminal chain and a short third intracellular loop (Strader et al., 1989). The third intracellular loop is one of the most specific domains of G protein-coupled receptors; thus, El Mestikawy et al. (1990) raised antibodies against a 26-amino acid portion of this loop that were specific for 5-HT_{1A} receptors. The third 5-HT_{1A} intracellular loop has no target either for protein kinase A (cAMP-dependent kinase), as does the β -adrenoceptor (Benovic et al., 1988), or for protein kinase II (calcium/calmodulin II-dependent kinase), which is abundant in the hippocampus. However, this loop possesses two threonine residues, targets for PKC phosphorylation.

The close link between phosphorylation and desensitization of the human 5-HT_{1A} receptor was demonstrated by Raymond (1991a), who found that the efficacy of the coupling to adenylyl cyclase decreased upon PKC activation in Chinese hamster ovary cells transfected with the 5-HT_{1A} receptor gene. Moreover, acute (2 min) pretreatment with 12-O-tetradecanoyl phorbol-13-acetate, a phorbol ester activator of protein kinase C, preferentially blocks some functions related to 5-HT_{1A} activation (IP₃ production and increase in [Ca²⁺]_i in LZD-7 cells and

inhibition of cAMP basal production in CH₄ZD₁₀ cells) but not others (inhibition of forskolin-stimulated cAMP production in both LZD-7 and CH₄ZD₁₀ cells) (Liu and Albert, 1991). The action of protein kinase C was potentiated by activation of protein kinase A, indicating that second messengers may modulate the expression of the pathways related to the 5-HT_{1A} receptor (Liu and Albert, 1991). The reduction of 5-HT_{1A} receptor function controlled by 5-HT₂ receptors, known to be coupled to phosphatidylinositides turnover and PKC activation (Kidd et al., 1990, 1991; Backus et al., 1990), may be of functional significance in this mechanism shown only in transfected cells.

Another particular feature of the 5-HT_{1A} structure is the presence of a "leucine zipper" in transmembrane domain III (Hartig et al., 1990a). This zipper exists in a series of DNA-binding proteins and oncogenes, where it is thought to facilitate dimer formation and, thus, interaction with other proteins (Landschulz et al., 1988; Struhl, 1989). This property is shared by 5-HT₂ receptors but not by other G protein-coupled receptors (Hartig et al., 1990a). Thus, although not demonstrated, it is not excluded that the functional interactions observed between 5-HT₂ and 5-HT_{1A} receptors may involve such protein-protein interactions.

As shown by mutagenesis experiments (Strader et al., 1989), the recognition site for agonistic ligands in the β -adrenoceptor included the Asp-113 residue. This residue is also present in 5-HT_{1A} transmembrane domain III, where it is thought to play a similar role (Hartig et al., 1990b). Hibert et al. (1991), using computerized modeling of receptors related to the G protein, suggested that this aspartate residue forms an ion pair with the amino group of 5-HT; this binding is stabilized by surrounding aromatic groups (one tryptophan and two phenylalanines). The indole moiety is also stabilized by a corresponding aromatic group, and, finally, the hydroxyl group of 5-HT is located in front of a serine residue. Weinstein and Osman (1990) proposed that the positively charged side chain (imidazolium) of the His-162 residue is involved in the binding of agonistic ligands and that such binding induces a proton transfer which, in turn, activates the receptor.

5. Transduction systems. The predominant transducing system linked to 5-HT_{1A} receptors is that of adenylyl cyclase. 5-HT_{1A} receptor was demonstrated to exert a negative control of adenylyl cyclase activity in numerous experiments, in which the activity of the enzyme was first strongly stimulated by forskolin in rat (De Vivo and Maayani, 1986; Oksenberg and Peroutka, 1988), guinea pig (De Vivo and Maayani, 1986, 1990), and calf hippocampus (Schoeffter and Hoyer, 1988), by vasoactive intestinal polypeptide in mice neuronal cultures from striatum, hippocampus, and cortex (Weiss et al., 1986; Bockaert et al., 1987; Dumuis et al., 1988c) or by calcium in rat hippocampus (Mörk and Geisler, 1990). The phar-

macological correlation of this activity with the 5-HT_{1A}-binding site is well established (Bockaert et al., 1987; Schoeffter and Hoyer, 1988).

The negative coupling of 5-HT_{1A} receptor with adenylyl cyclase was confirmed by Harrington et al. (1988) and Okada et al. (1989), who showed that pertussis toxin, a specific inhibitor of G_i protein, attenuated the observed effect. Moreover, Fargin et al. (1989) and Albert et al. (1990), using cultured transfected cells (COS-7, HeLa, and GH₄ZD₁₀) expressing 5-HT_{1A} receptors, demonstrated that 5-HT (in the nanomolar range) inhibited forskolin-stimulated adenylyl cyclase activity. Electrophysiological experiments (Andrade et al., 1986; Clarke et al., 1987) with pertussis toxin also indicated that 5-HT_{1A} receptor is coupled to a G_i protein.

Although, the negative coupling of 5-HT_{1A} receptors with adenylyl cyclase activity is well established and involves G_i protein, several experimental observations indicate that there may also be a positive coupling. Initially, serotonergic agonists were found to activate this enzyme *in vitro* in the rat hippocampus (Barbaccia et al., 1983a; Markstein et al., 1986), guinea pig hippocampus (Shenker et al., 1983, 1985, 1987), and rat brain cortex (Fayolle et al., 1988, Mörk and Geisler, 1990). Recently, an increase in cAMP production induced by 8-OH-DPAT and antagonized by *d*-propranolol was observed *in vivo* by using a microdialysis technique (Sijbesma et al., 1991). The activation of adenylyl cyclase correlates pharmacologically to 5-HT_{1A}-binding sites, suggesting that the receptors are coupled to a G_s protein.

However, other authors did not observe any 5-HT_{1A}-related activation in the cortex (Weiss et al., 1986; De Vivo and Maayani, 1990) or in the hippocampus (Oksenberg and Peroutka, 1988). Maayani's group, who initially reported the activation of adenylyl cyclase by 5-HT_{1A} in guinea pig hippocampus (Shenker et al., 1985, 1987), did not find the same pharmacological profile of the 5-HT-induced activation after modifying the experimental procedure (De Vivo and Maayani, 1990).

Thus, the adenylyl cyclase activation related to 5-HT_{1A} receptors might be attributed to experimental artifacts, because preparation of the tissue may result in the coupling of receptors to G_s-proteins, which are not normally present in the same cellular compartment. However, it is difficult to conceive that membrane proteins (G_s) may be released into the medium and relocated in other membranes close to 5-HT_{1A} receptors as functional entities. Moreover, these activations have been demonstrated *in vivo* by a microdialysis technique (Sijbesma et al., 1991).

The stimulation of the enzyme by 5-HT_{1A} agonists raises the question of whether the 5-HT_{1A} receptors related to the activation and/or the inhibition of the adenylyl cyclase activity correspond to two different receptors (not distinguished by the existing ligands) linked to G_s and G_i, respectively, or to a single type

coupled either to G_i or to G_s protein. Experimental evidence showed, on the one hand, that reconstituted receptors coupled to a G_s protein (D₁ or β_2 -adrenoceptors) are also able to couple to inhibitory G proteins (i.e., G_i) in the presence of G_s (Sidhu et al., 1991; Rubenstein et al., 1991). The alternative coupling of the receptor may depend on the phosphorylation of the third loop by a protein kinase A. The β_2 -adrenoceptor-G_s protein coupling was drastically reduced (resulting in a total loss of adenylyl cyclase stimulatory activity) after phosphorylation of a serine residue located in the third intracellular loop of the receptor; at the same time, the coupling with G_i protein was potentiated (Okamoto et al., 1991). On the other hand, G_i-coupled inhibitory receptors (α_2 , D₂, A₁) expressed in cultured cells could also stimulate cAMP synthesis through the activation of a particular type of adenylyl cyclase (type II) by β subunits of a G_i protein (Federman et al., 1992).

In conclusion, the fact that 5-HT_{1A} receptors have been found to inhibit or to stimulate the adenylyl cyclase activity may be due either to the existence of different 5-HT_{1A} receptors specifically linked to G_i or to G_s, or to the same receptor linked alternatively to G_i or G_s, depending on its phosphorylation state, or even an inhibitory G protein can mediate the stimulation of certain types of adenylyl cyclase through $\beta \gamma$ subunits.

The 5-HT_{1A} receptors are also directly linked to the PLC activity. This signal transduction pathway of the 5-HT_{1A} receptor depends on the cell type in which it is expressed. 5-HT_{1A} receptors stimulate PLC in HeLa and LZD-7 cells but not in GH₄ZD₁₀ pituitary and COS-7 kidney cells (Fargin et al., 1989; Raymond et al., 1989b; Liu and Albert, 1991). In HeLa cells, the EC₅₀ of 5-HT for phosphatidylinositides turnover was 10-fold higher than it was for inhibition of adenylyl cyclase; however, the EC₅₀ values for these two actions were similar in LZD-7 cells.

The increase of the IP₃ production is sensitive to pertussis toxin and independent of cAMP (Fargin et al., 1989). It is accompanied by the mobilization of calcium (Middleton et al., 1990), by an increase of the sodium-dependent phosphate uptake via protein kinase C activation (Raymond et al., 1989b, 1991), and by an activation of Na⁺/K⁺-ATPase (Middleton et al., 1990).

That PLC activation is observed in transfected cells expressing an unusually high density of 5-HT_{1A} receptors may raise the question of the relevance of the mechanism under physiological conditions. However, the level of expression of 5-HT_{1A} receptors in LZD-7 cells (1.9 pmol/mg protein) is comparable to that of their expression in CH₄ZD₁₀ cells (1.1 pmol/mg protein), in which no changes in phosphatidylinositides hydrolysis is observed. This indicates that the high density of expressed receptors in transfected cells is not the sole factor responsible for the activation of the PLC. Moreover, the IP₃ production was shown to be induced by 5-HT₁ receptors in the

hippocampus (Janowsky et al., 1984); a more complete understanding of the pharmacology is needed to attribute this effect to 5-HT_{1A} receptor function.

On the other hand, Claustre et al. (1988a) showed that 8-OH-DPAT (10 nM) reduces the IP₃ production induced by muscarinic receptor activation; this effect is independent of cAMP, K⁺ channels, and pertussis toxin and is probably related to phospholipase A₂ interacting with carbachol-stimulated PLC (Claustre et al., 1991).

In addition to the coupling of 5-HT_{1A} receptors with membrane-bound enzymes (adenylyl cyclase, phospholipase), these receptors also control G protein-coupled K⁺ channels, notably in the hippocampus (Andrade et al., 1986), in the dorsal raphe nucleus (Innis et al., 1988), and in the ventromedial hypothalamus (Newberry, 1992). This system is sensitive to pertussis toxin (Andrade et al., 1986; Clarke et al., 1987) and to 4-aminopyridine (Haj-Dahmane et al., 1991) and clearly does not require second messengers such as cAMP, diacylglycerol, and IP₃. Activation of the 5-HT_{1A} receptor leads to the hyperpolarization of the cell membrane through increased K⁺ conductance, resulting in decreased neuronal firing. In the hippocampus, the opening of the potassium channels by 5-HT_{1A}, GABA_B (Andrade et al., 1986; Colino and Hallivell, 1987; Innis and Aghajanian, 1987), and A₁ receptors (Zgombick et al., 1989) involves a pertussis toxin-sensitive G protein, which may directly couple these receptors to the same K⁺ channel, because their responses are nonadditive.

It is not surprising that 5-HT_{1A} receptors take part in different transduction systems, because a similar situation prevails for other neurotransmitter receptors, namely, m₁ (Stein et al., 1988), m₂ (Ashkenazi et al., 1987; Lai et al., 1991), m₄ muscarinic receptors (Peralta et al., 1988), and D₁ dopaminergic (Sidhu et al., 1991). The existence of these multifunctional receptors raises certain questions. Data indicate that various receptors may interact with more than one of the G proteins present in the membrane (see review by Birnbaumer et al., 1990, and above, this section). Although the molecular mechanism by which a single receptor couples to multiple G proteins is poorly understood, the phosphorylation of the receptor seems to be an important mechanism in defining the specificity of the coupling with a given G protein (Okamoto et al., 1991; Liu and Albert, 1991). Another question is whether the different electrophysiological and biochemical responses occur in the same cell or in different neuronal populations. This question is still unresolved, although it was addressed by Andrade (1990). In addition, further investigations are needed to explain the relative importance of the receptors coupled to several transduction systems, in the function of a single cell or in that of different cell populations.

6. *Possible heterogeneity of the 5-HT_{1A} receptor subtype.* Several experimental observations suggest the existence of different 5-HT_{1A} receptor subtypes. In the raphe, the

5-HT_{1A} receptors are not linked to the inhibition of adenylyl cyclase activity as they are in the hippocampus (Yocca and Maayani, 1990). Moreover, as demonstrated by electrophysiological, biochemical and behavioral studies, several 5-HT_{1A} agonists have different efficacies, depending on the brain region (raphe, hippocampus, cortex, or even different areas of hippocampus such as CA1 or dentate gyrus). Brown et al. (1990), who compared the efficacy of RS 30199-193 in rat brain and ileum, also suggested the existence of several 5-HT_{1A} receptor subtypes.

Studies of the adaptive regulatory mechanisms of 5-HT_{1A} receptors in response to chronic treatment with various drugs also revealed differences between the pre- and postsynaptic receptors. For example, chronic treatment with IMAO (Blier and de Montigny, 1985), SRI (Blier and de Montigny, 1983; Chaput et al., 1986), or the 5-HT_{1A} agonists, gepirone (Blier and de Montigny, 1987b, 1990) and ipsapirone (Schechter et al., 1990), provokes a desensitization of the somatodendritic 5-HT_{1A} autoreceptors (raphe), without any change of hippocampal postsynaptic 5-HT_{1A} receptors.

The possibility that regional differences exist for 5-HT_{1A} receptors in the CNS was recently investigated by Radja et al. (1992) using quantitative autoradiography. Although pronounced regional differences in the pharmacological properties were not observed, physicochemical properties of the receptors revealed a heterogeneity more related to the CA1 area versus dentate gyrus than to the pre- or postsynaptic location of these receptors. Therefore, the functional and regulatory differences that depend on the stimulation of pre- or postsynaptic receptors could not be due to the intrinsic properties of binding sites but, rather, might be related to the intracellular pathways of transduction.

These data suggest that all 5-HT_{1A} receptors are not identical throughout the brain. In agreement with these findings, a northern blot analysis by Albert et al. (1990) of 5-HT_{1A} mRNAs in rat brain showed that the detected mRNA sequences had sizes of 3.9, 3.6, 3.3, and 2.2 kb in some brain areas. This suggests that there are either different transcriptional start sites or different polyadenylation sites. Similar results were obtained in our laboratory with a more specific probe (unpublished results). Further experiments are needed to explain the functional relevance of these bands.

7. *Regulation.* In vitro, 5-HT_{1A} receptors, like G protein-related receptors, interact with the guanine nucleotides. GTP modulates receptor affinity allosterically in a reversible manner; it decreases the receptor affinity without any change in the apparent B_{max} of the [³H]8-OH-DPAT-binding sites. However, its nonhydrolysable analogs (5-guanylyl imidodiphosphate, GTP γ -S) induce a persistent decrease in both the affinity and the apparent density of 5-HT_{1A} sites (Hall et al., 1985; Schlegel and Peroutka, 1986; Harrington and Peroutka, 1990a).

According to the model proposed by Harrington and Peroutka (1990a), the receptor-G protein complex can exist in different interconverting states and only few of them would bind the agonist. The K_d and the B_{max} values depend on the equilibrium that exists among the different states. The addition of GTP moves the equilibrium toward "nonbinding" states, resulting in a lower affinity of the agonist. The significant decrease of the B_{max} by the nonhydrolysable nucleotides could be explained by the persistent nature of their binding to G proteins, resulting in a disruption of the interconvertibility of the receptor states.

A GTP-insensitive component of the 5-HT_{1A}-solubilized binding sites was described (Emerit et al., 1991; Elliott and Phipps, 1992). It was proposed that the different states of oxidation of SH-groups of the receptor may influence the coupling with the G protein and induce the occurrence of the GTP-insensitive component.

During brain development, 5-HT_{1A} receptors are highly regulated. Thus, Daval et al. (1987) showed that the number of 5-HT_{1A} receptors in the rat cortex, hippocampus, and septum increases markedly during the first 3 weeks after birth and is stable up to an advanced age (Gozlan et al., 1990). In contrast, in the human brain, an age-related decrease of 5-HT_{1A} binding is observed that is specific to cortical, hippocampal, and raphe nuclei (Dillon et al., 1991). A different regulation mechanism exists in the cerebellum where a high number of functional receptors are already present at birth but disappear by adulthood (Daval et al., 1987; Hamon et al., 1990b). This presence of 5-HT receptors at an early stage of the cerebellum development might reflect a trophic role of the serotonergic system in this area.

In general, long-term variations of receptor stimulation regulate the receptor number. However, lesions of the serotonergic neurons that project to the hippocampus, when provoked by repetitive administration of 5,7-DHT, modify neither the number of 5-HT_{1A} receptors (Vergé et al., 1986) nor their activity (Hamon et al., 1990b). Several studies have shown that a single administration of a 5-HT_{1A} agonist causes a rapid, marked, and prolonged decrease of the 5-HT_{1A}-mediated functions (hypothermia, hyperalgesia) (Larsson et al., 1990; Kennett et al., 1987c; Beer et al., 1990); these decreases are accompanied by a 25% rapid downregulation of the number of 5-HT_{1A}-binding sites in the raphe but not in the hippocampus (Beer et al., 1990). Hjorth (1991), however, did not observe any alteration of the inhibitory effect of 8-OH-DPAT on 5-HT release under similar conditions; these discrepancies could be resolved if it turns out that a large pool of autoreceptors is held in reserve (Meller et al., 1990). That hypothermia and hyperphagia are desensitized, whereas 5-HT release is not affected under the same conditions, suggests that different 5-HT_{1A} receptor populations are involved in these phenomena. The validity of this hypothesis remains to be established.

Functional interactions occur between 5-HT₂ and 5-HT_{1A} receptors; in particular, the former seem to control the activity of the latter. Weiss et al. (1986) showed that, in vitro, in cortical neurons, ketanserin, at low concentration, significantly increases the negative effect of 5-HT on the adenylyl cyclase activity. This interaction also takes place in vivo because long-term treatment with a 5-HT₂ antagonist (ketanserin) or agonist (DOI), which both desensitize the 5-HT₂ receptors, decreased the 5-HT_{1A} autoreceptor effect on 5-HT release (Kidd et al., 1990, 1991). Moreover, acute administration of ritanserin or ketanserin, which desensitizes 5-HT₂ receptors, increased the 5-HT_{1A} syndrome (Backus et al., 1990). In addition, activation of 5-HT₂ receptors on pyramidal cells of layer V of the medial prefrontal cortex reduces the ability of 5-HT_{1A} receptors to hyperpolarize these cells (Araneda and Andrade, 1991). Thus, it is likely that 5-HT₂ activity inhibits the function of 5-HT_{1A} receptors.

However, Arnt and Hyttel (1989), measuring forepaw treading in the rat, reported that DOI facilitated 5-HT_{1A} function. Moreover, Garratt et al. (1991) recently showed that the inhibitory effects of DOI on the firing of 5-HT neurons and the decrease of the extracellular 5-HT concentration in the frontal cortex were not mediated by 5-HT₂ because they were not antagonized by ketanserin, ritanserin, or pindolol. Recent findings demonstrating the membrane-stabilizing properties of DOI (Martin et al., 1991b) may explain these contradictory results. In addition, the 5-HT_{1A}-5-HT₂ interactions, as shown by Pranzatelli and Pluchino (1991), are limited to a few behavior patterns involving serotonergic receptors, i.e., forepaw myoclonus, shaking behavior, and thermoregulation, but are not involved in skin jerks, flat body posture, or head weaving. On the other hand, Yocca et al. (1990b) proposed that 5-HT_{1A} receptors may control the 5-HT₂ head shake response in rats, suggesting the reciprocal interaction of 5-HT_{1A} on the 5-HT₂-induced activity.

Many experimental studies have been devoted to the interaction of antidepressant drugs with 5-HT_{1A} receptors. However, at the present time, the mechanism of this interaction is not fully understood. Two interpretations of data concerning what occurs after long-term antidepressant treatment have been presented: one favors a desensitization of the postsynaptic 5-HT_{1A} receptors; the other suggests the hypersensitization of these receptors that may be related to a desensitization of the presynaptic autoreceptors in raphe. Indeed, long-term treatment with various antidepressants (IMAO, TCA, 5-HT_{1A} agonists, electroconvulsive shock) or with lithium have been shown to downregulate the postsynaptic 5-HT_{1A} receptors located in the cortex or in the hippocampus (Mizuta and Segawa, 1988, 1989; Odakaki et al., 1990; Pandey et al., 1991), or to induce a desensitization of their effect on adenylyl cyclase activity (Sleight et al., 1988; Newman and Leler, 1988a; Varrault et al., 1991;

Newman et al., 1991, 1992), or their effect on the serotonergic syndrome (Lucki and Frazer, 1982; Goodwin, 1989).

On the other hand, it was also shown that long-term treatment with antidepressants led to the hypersensitivity of the postsynaptic 5-HT_{1A} receptors or to the desensitization of the autoreceptors in the raphe, resulting in both cases in a facilitation of the serotonergic transmission. Indeed, chronic treatment with TCAs (amitriptyline, imipramine, desimipramine), or electroconvulsive shock increased the number of 5-HT_{1A} receptors in the cortex and the hippocampus and the responsiveness of hippocampal pyramidal neurons to 5-HT without modifying 5-HT_{1A} receptors in the raphe (Blier et al., 1988; Welner et al., 1989; Nowak and Dulinski, 1991). TCA, IMAO, 5-HT uptake inhibitors (zimetidine or citalopram), 5-HT_{1A} agonists (buspirone, gepirone, ipsapirone), or lithium exerted the following effects: in the raphe, they desensitized the 5-HT_{1A} receptors and reduced their number (Welner et al., 1989; Gobbi et al., 1991); they weakened their inhibitory effect on serotonergic neuronal firing (Blier and de Montigny, 1983, 1985, 1987a,b; Chaput et al., 1986, 1988; Schechter et al., 1990); they enhanced the release of 5-HT from serotonergic neurons (Sharp et al., 1991), and they attenuated their hypothermic response both in mice (Goodwin et al., 1985a; De Souza et al., 1986; Goodwin, 1989) and in humans (Lesh et al., 1990) without changing the postsynaptic activity of the 5-HT_{1A} receptors located in the hippocampus. These effects resulting in a facilitation of the serotonergic transmission are in agreement with the decrease of hippocampal 5-HT_{1A} sites observed in suicidal/depressed individuals (Cheetham et al., 1989, 1990).

In addition to these contradictory results, several experiments in which chronic treatment with different antidepressants was used failed to show any modification in [³H]8-OH-DPAT-binding sites in the hippocampus and in the raphe regions (Frazer and Hensler, 1990; Schechter et al., 1990; Newman et al., 1990, 1991; Hensler et al., 1991; Pandey et al., 1991), in the serotonergic syndrome (Lucki and Frazer, 1982), in the response of cortical or hippocampal pyramidal neurons to iontophoretically applied 5-HT (Olpe and Schellenberg, 1981; Olpe et al., 1984), or in the inhibition of the forskolin-stimulated adenylyl cyclase induced by 5-HT in hippocampus (Frazer and Hensler, 1990; Schechter et al., 1990).

It is difficult to reconcile these results and explain the discrepancies. Differences exist in the effects of different antidepressants in similar experiments, or even in the effects of a single antidepressant in the same model, depending on the investigators, not only between electrophysiological and biochemical data but also between the regulatory properties of pre- and postsynaptic 5-HT_{1A} receptors. Thus, we can make no conclusions from the adaptive changes of 5-HT_{1A} receptors in response to

the antidepressant treatments. That the heterogeneity of 5-HT_{1A} receptors is likely at various levels (genes, physicochemical properties, transduction systems, regional distribution, and regulation mechanisms) might explain some of the results reported in the literature. A better understanding of the identity and functions related to these receptors, as well as their interactions with other receptors, will certainly allow a clarification of the role of 5-HT_{1A} receptors in the mechanism of action of the antidepressants.

The 5-HT_{1A} receptors appear to be regulated by hormonal levels. The 8-OH-DPAT-induced corticosterone release was attenuated by ovariectomy in female rats, and, in contrast, this effect was enhanced by castration in male mice (Matsuda et al., 1991). These findings suggest that the adrenocortical effect of 8-OH-DPAT is altered by gonadal hormones. Indeed, estrogen treatment (3 to 5 days) in ovariectomized female rats or in castrated mice increases 5-HT_{1A}-related adenylyl cyclase activity (Clarke and Maayani, 1990), electrophysiological activity (Beck et al., 1989; Clarke and Goldfard, 1989), and corticosterone release (Matsuda et al., 1991). However, the mechanism for estrogen's enhancement of 5-HT_{1A} responsiveness is unknown.

The existence of a bidirectional intramembrane interaction between 5-HT_{1A} and galanin receptors was demonstrated by Fuxe et al. (1988; Hedlund and Fuxe, 1991). Thus, galanin (10 nM), *in vitro*, selectively reduces the affinity of the 5-HT_{1A} receptors within the rat ventral limbic cortex, without altering the binding of 5-HT_{1B} or 5-HT₂ receptors. In contrast, the 5-HT_{1A} receptor agonist, 8-OH-DPAT (10 nM), increases the affinity of the galanin receptors by approximately 55% in various telencephalic and diencephalic areas (Hedlund et al., 1991a). The increased affinity of galanin-binding sites by 8-OH-DPAT seems to reflect a G protein-independent intramembrane receptor-receptor interaction between 5-HT_{1A} and galanin receptors. This interaction, observed *in vitro*, is of biological relevance, because the receptors interact synergistically in the control of central cardiovascular regulation (Hedlund et al., 1991b).

Several opioid narcotics (sufentanil, fentanyl) at high concentrations inhibit the binding of [³H]8-OH-DPAT to rat membranes; this inhibition appears to be competitive, because these drugs increase the *K_d* without altering the number of 5-HT_{1A} sites. However, no changes in the sensitivity of 5-HT_{1A} receptors to guanine nucleotides are observed. These findings suggest that these narcotics disrupt neurotransmission at 5-HT_{1A} receptors during clinical anesthesia (Martin et al., 1991a).

It is known that alcohol consumption modifies serotonin levels (Carlsson et al., 1980; Gorelick, 1989), serotonin turnover (Morinan, 1987), and the serotonin transport system (Gross-Isseroff and Biegon, 1988) in rat and human brain. Recently, it was demonstrated that alcohol consumption coincides with the alteration of binding to

5-HT_{1A} receptors; cortical [³H]-8-OH-DPAT binding was decreased in subjects in whom alcohol was detected at autopsy (Dillon et al., 1991).

Deficits of the serotonergic system have been established in Alzheimer disease (see review by Cross, 1990) and include a reduction in the number of 5-HT_{1A}-binding sites in the frontal, temporal, and parietal cortices (Middlemiss et al., 1986). However, the modulation of [³H]-8-OH-DPAT binding by guanine nucleotides did not vary significantly in these regions in control patients and those with Alzheimer's disease, suggesting that the functional capacity of 5-HT_{1A} receptor is intact in the brain of persons with Alzheimer's disease (O'Neill et al., 1991).

8. Cellular functions. 5-HT_{1A} receptors are present in the raphe on 5-HT neurons as presynaptic autoreceptors that regulate 5-HT neuronal activity (Aghajanian, 1978; Van der Maelen et al., 1986; Sprouse and Aghajanian, 1987) and 5-HT synthesis and release (Hjorth and Magnusson, 1988; Hamon et al., 1988; Hjorth and Sharp, 1991). Meller et al. (1990) suggested the existence of a large reserve of these 5-HT_{1A} autoreceptors. Interestingly, Sinton and Fallon (1988) and Blier et al. (1990) reported that the serotonergic neurons in the dorsal raphe were less sensitive to 5-HT_{1A} agonists than were those in the median raphe, suggesting the existence of different mechanisms regulating the activity of the ascending 5-HT projections. However, Hjorth and Sharp (1991), using *in vivo* microdialysis, demonstrated that the inhibition of 5-HT release by 5-HT_{1A} was identical in several areas innervated by dorsal or median raphe, except in the globus pallidus.

5-HT_{1A} receptors are also located on nonserotonergic neurons, either as heterologous presynaptic receptors on terminals or as postsynaptic receptors on soma or dendrites. These receptors are thought to modulate the release of other neurotransmitters. For example, 8-OH-DPAT enhances ACh release in rat or guinea pig cortex (Siniscalchi et al., 1990a). 5-HT_{1A} receptors also reduce the glutamate-induced excitation of adrenergic neurons in the locus coeruleus (Charlety et al., 1991). Chaouloff and Jeanrenaud (1987) reported that 8-OH-DPAT is involved in the control of the adrenaline-induced hyperglycemia resulting from enhanced adrenaline release via 5-HT_{1A} receptor activation (Bagdy et al., 1989; Chaouloff et al., 1990). Hamon et al. (1988) reported that 5-HT_{1A} agonists (gepirone, buspirone, ipsapirone) increase the dopamine turnover, however, 8-OH-DPAT was inactive, suggesting that this effect was not mediated by 5-HT_{1A} autoreceptors. In contrast, Benloucif and Galloway (1991) using microdialysis showed that the dopamine-releasing effect occurs in the presence of TFMPP, mCPP, RU 24969, and also 8-OH-DPAT and proposed that 5-HT_{1A} receptors are involved in dopamine release.

The core of experimental results in this domain deals with the effect of 5-HT_{1A} receptors on the control of the hypothalamo-hypophysis axis. 8-OH-DPAT, buspirone,

gepirone, and ipsapirone stimulate the release of adrenocorticotropine (Gilbert et al., 1987, 1988b; Sciuillo et al., 1990), β -endorphin (Koenig et al., 1987, 1988; Sciuillo et al., 1990), corticosterone (Koenig et al., 1987, 1988; Matheson et al., 1989; Haleem et al., 1989; Matsuda et al., 1991; Owens et al., 1990), and corticotropin-releasing factor (Owens et al., 1990). These effects, presumably, are not due to an interaction with the cholinergic system (Gilbert et al., 1988a) but more likely to an action on the observed secretions via a 5-HT_{1A} postsynaptic receptor (Gilbert et al., 1988a). This mechanism might be involved in the circadian regulation of these secretions (Halasz and Banky, 1986), their negative feedback (Beaulieu et al., 1986), or their involvement in stress mechanisms (Haleem et al., 1989).

9. 5-HT_{1A} receptors and behavior. Investigations of animal behavior have been considerably facilitated by the development of selective drugs for 5-HT_{1A} receptors. These receptors appear to be involved in many kinds of behavior (cf. reviews by Lucki and Wieland, 1990; Glennon, 1990b; Murphy, 1990; Glitz and Pohl, 1991; Wilkinson and Dourish, 1991; Lucki, 1992).

The main and most obvious effect of 5-HT_{1A} agonists in vertebrates is that they provoke the serotonergic syndrome, which consists in mice and rats of an overstimulated state leading to motor modifications (hyperlocomotion, head weaving, reciprocal forepaw treading, flat body posture) (Tricklebank, 1987; Yamada et al., 1988, 1989). In humans, this syndrome results in confusion, nervousness, hyperreflexia, restlessness, hypomania, excitation, myoclony, and shivering (Sternbach, 1991). Postsynaptic 5-HT_{1A} receptors are thought to mediate these effects (Tricklebank, 1987). In the rat, the different components of the serotonergic syndrome manifest themselves at different phases during postnatal ontogenesis; hyperlocomotion and head weaving occur as early as the fifth day after birth, whereas hindlimb scratching and forepaw treading are observed only after the third postnatal week (Jackson and Kitchen, 1989).

Presynaptic autoreceptors in the median raphe are more likely to induce hypolocomotion, especially affecting the horizontal movements (Hillegaart et al., 1989; Hillegaart and Hjorth, 1989; Mittman and Geyer, 1989). Ahlenius et al. (1991) also observed an increase of the forward and peripheral locomotion under similar conditions.

Sexual behavior is controlled in part by 5-HT_{1A} activity; however, the mechanism by which it does so is not yet precisely known. In male rats, 5-HT_{1A} agonists (8-OH-DPAT, buspirone, ipsapirone, flesinoxan) facilitate sexual activity (Schnur et al., 1989; Ahlenius and Larsson, 1989; Berendsen and Broekkamp, 1990; Mos et al., 1990; Ahlenius et al., 1991), although 5-HT or its precursors have been reported to inhibit it (Ahlenius and Larsson, 1987). An explanation for this paradox might reside in the preferential presynaptic activity of 8-OH-

DPAT but also may result from a 5-HT_{1B} inhibition of male sexual behavior (Fernandez-Guasti et al., 1989).

The effect of 5-HT_{1A} on sexual behavior is sex dependent; in female rats, 8-OH-DPAT inhibits lordosis behavior (Mendelson and Gorzalka, 1986a), in contrast to its facilitatory effect in male rats. This effect is presumably due to the 5-HT_{1A} control of hypothalamic activity, because the lordosis behavior is suppressed by intrahypothalamic administration of 8-OH-DPAT (Uphouse et al., 1991a,b). The difference in the observed 5-HT_{1A} activity in the male and female might be expected, because functional and anatomical differences between males and females with respect to the 5-HT system are present (Uphouse et al., 1991b). Furthermore, this effect is species dependent. In male mice, 8-OH-DPAT inhibits sexual behavior (Svensson et al., 1987), whereas in male rats it is facilitatory. Effects are dose dependent, because whether ipsapirone and gepirone have facilitatory or inhibitory effects on lordosis behavior depends on the dose (Fernandez-Guasti et al., 1987).

5-HT_{1A} receptors participate in body temperature regulation. Agonists induce hypothermia in rats (Hjorth, 1985; Goodwin et al., 1987) and mice (Goodwin et al., 1985b). The mechanism of temperature regulation has not yet been fully clarified. In the mouse, there is good evidence that the 5-HT_{1A} receptor mediating 8-OH-DPAT-induced hypothermia are presynaptic, because 5,7-DHT administration for 2 weeks with pCPA abolished the hyperthermia induced by 8-OH-DPAT (Goodwin et al., 1985b; Bill et al., 1991). In the rat, there is some controversy as to whether thermoregulatory 5-HT_{1A} receptors are presynaptic (Goodwin et al., 1987) or postsynaptic (Hjorth, 1985; Hutson et al., 1987). The latter authors found that 5-HT depletion or 5,7-DHT lesions, rather than attenuating 8-OH-DPAT-induced hypothermia, tended to enhance the response. However, Goodwin et al. (1987) found that pCPA-induced 5-HT depletion prevented 8-OH-DPAT-induced hypothermia in the rat, concluding that the 5-HT_{1A} receptors mediating this response are presynaptic. In an attempt to resolve this controversy Bill et al. (1991), carried out two parallel series of experiments (5-HT depletion techniques by 5,7-DHT and pCPA and use of agents that facilitated the 5-HT release) in rat and mouse. They concluded that an important species difference exists, because the 8-OH-DPAT-induced hypothermia is mediated by presynaptic autoreceptors in the mouse and by postsynaptic 5-HT_{1A} receptors in the rat.

A sex difference in the hypothermic effect of 8-OH-DPAT has also been reported. This effect, more marked in female than in male rats (Carlsson and Eriksson, 1987), may reflect a differential clearance of the drug (Kato, 1974) or even a differential sensitivity of the pre- and postsynaptic 5-HT_{1A} receptors in male and female rats. Indeed, the 5-HT system is reported to be more active in females than in males; for example, females

exhibit less evidence of a behavior that requires autoreceptor activation (e.g., hyperalgesia) and greater evidence of behaviors that require activation of the postsynaptic sites (e.g., hypothermia, 5-HT syndrome) (Uphouse et al., 1991a,b).

The 5-HT_{1A} receptors play an important role in feeding behavior. 5-HT_{1A} agonists (8-OH-DPAT, ipsapirone, buspirone, gepirone, LY 165-163) increase food intake in rats (Dourish et al., 1985; Gilbert and Dourish, 1987; Hutson et al., 1988), mice (Shepherd and Rodgers, 1990), and cats (Jacobs et al., 1989). This effect is opposite to that of 5-HT itself (Blundell, 1977). This difference might be explained by the activation of the 5-HT_{1A} presynaptic autoreceptors, which decreases the serotonergic activity. This hypothesis is supported by the fact that local injection of 8-OH-DPAT in the raphe increases food intake (Hutson et al., 1986; Bendotti and Samanin, 1987) and by the absence of 8-OH-DPAT- and LY 165-163-induced hyperphagia after depletion of 5-HT by pCPA (Bendotti and Samanin, 1986; Dourish et al., 1986a,b,c). Moreover, this effect of 5-HT_{1A} agonists on feeding behavior leads to rapid tolerance development, presumably due to desensitization of the autoreceptors (Kennett et al., 1987c; Goudie et al., 1989). Hyperphagia was most evident in diestrous females and least evident in proestrous and estrous rats; these findings are interpreted as the modulation of somatodendritic 5-HT_{1A} autoreceptors by the estrous cycle (Uphouse et al., 1991b).

A major reason for the interest in 5-HT_{1A} receptors is their implication in psychiatric disorders, such as anxiety and depression. Anxiolytic properties of 5-HT_{1A} agonists in humans have been reported for buspirone (Goldberg and Finnerty, 1979; Rickels et al., 1982; Feighner et al., 1982; Goa and Ward, 1986; Robinson et al., 1989; Böhm et al., 1990; Rakel, 1990), gepirone (Csanalosi et al., 1987; Cott et al., 1988; Harto et al., 1988; Robinson et al., 1989), ipsapirone (Glaser, 1988; Beneke et al., 1988; Kuemmel et al., 1988), and tandospirone (Hirose et al., 1986; Heym et al., 1987; Shimizu et al., 1987). The anxiolytic properties of the 5-HT_{1A} agonists are clearly different from those of the benzodiazepines. A delay of several days is required before 5-HT_{1A} agonists begin to have a therapeutic effect, whereas the effect is almost immediate with benzodiazepines. Furthermore, the 5-HT_{1A} agonists do not produce sedation, ataxia, or amnesia and are not associated with withdrawal reactions following abrupt discontinuation (Goa and Ward, 1986; Harto et al., 1988; Taylor and Moon, 1991). Thus, for treating anxiety, these substances have several advantages over benzodiazepines. However, that chronic treatment with buspirone increases (41%) the number of benzodiazepine receptors in the substantia nigra suggests that the two systems interact (Gobbi et al., 1991).

Drugs related to 5-HT_{1A} have also been found to be anxiolytic in various experimental animal models (cf.

reviews by Gardner, 1986; Dourish, 1987; Traber and Glaser, 1987, Traber et al., 1988; Young and Glennon, 1988), such as electric shock- or isolation-induced aggressiveness, ultrasonic vocalization in newborn rats (Benton and Nastiti, 1988; Hard and Engel, 1988; White et al., 1991), exploration in an aversive environment (Merlo-Pich and Samanin, 1986; Costall et al., 1988b), aggressiveness in wild rats (Blanchard et al., 1988), and the drinking conflict test (Chojnacka-Wojcik and Przegalinski, 1991). On the other hand, 8-OH-DPAT increases the isolation-induced social behavioral deficit that is reversed by 5-HT_{1B} agonists, benzodiazepines, or chronic treatment with antidepressants (Francès et al., 1990). However, it should be pointed out that the role of 5-HT_{1A} agonists in animal models of anxiety is controversial, because these drugs had variable or no effects in several conflict models of anxiety in primates or rodents (Geller and Hartmann, 1982; see reviews by Traber and Glaser, 1987; Wilkinson and Dourish, 1991); 5-HT_{1A} agonists are efficient apparently only in the pigeon conflict model. Moreover, these drugs were reported to have no effect in the social interaction test in the rat (File, 1985; Pellow et al., 1987). In addition, anxiogenic-like effects were reported after systemic injections (Critchley and Handley, 1987; Moser et al., 1990).

These contradictory observations might have several explanations, one of which is that the actual efficacy of some 5-HT_{1A} agonists is lacking; more likely, the animal models used to determine the anxiolytic effects of the drugs are not adapted to the 5-HT_{1A} mechanism of action, i.e., they may measure impulsivity rather than anxiety (Soubrié, 1986). Also, the experimental conditions used to determine this activity do not consider the fact that the anxiolytic effect in humans occurs after several days of treatment.

The anxiolytic properties of 5-HT_{1A} agonists are mediated by mechanisms that have not been fully clarified. An important question resides in determining the nature of the anxiolytic compounds as full or partial 5-HT_{1A} agonists at pre- and postsynaptic receptors. The anxiolytic effects of the 5-HT_{1A} agonists in the Vogel conflict test and social interactions paradigm possibly involve presynaptic autoreceptors, because these effects are observed after local injections of 8-OH-DPAT, ipsapirone, or buspirone in the dorsal raphe area (Higgins et al., 1988, 1989). Moreover, the anxiolytic effect of an injection of buspirone in the median raphe is blocked by a 5,7-DHT lesion (Carli et al., 1989). The same authors did not observe an anxiolytic action of buspirone after injection into the dorsal raphe area in the light/dark box model.

Several experimental arguments suggest that postsynaptic 5-HT_{1A} receptors are also involved, because anxiolytic effects of buspirone are still seen when there are lesions in the raphe (Davis et al., 1988). The main hypothesis presented to explain the mechanism of anxiolytic

action of the 5-HT_{1A} agonists is based on their full activity at presynaptic autoreceptors in the raphe and their partial, or even antagonistic, properties at postsynaptic receptors. This is suggested by the numerous observations reported above on electrophysiological, biochemical, and behavioral grounds.

In summary, although the 5-HT_{1A} agonists appear to possess promising anxiolytic properties, their therapeutic effect is not yet fully characterized. It should be pointed out that anxiolytic properties of 5-HT_{1A} agonists are usually observed after a single administration in animal models, whereas this effect is obtained only after several days of treatment in humans. This raises the question of whether the therapeutic effect is observed after the increase of the serotonergic activity (acute effects) or its decrease (chronic treatment resulting in desensitization of the autoreceptors). A better understanding of the serotonergic mechanisms involved in anxiety presumably will lead to an enhancement of the therapeutic properties of these drugs.

5-HT_{1A} receptors appear to be involved also in depression and in the mechanism of action of antidepressants. The following 5-HT_{1A} agonists have been shown to have antidepressant properties in humans: buspirone (Schweizer et al., 1986; Robinson et al., 1989), gepirone (Amsterdam et al., 1987; Cott et al., 1988; Robinson et al., 1989; Jenkins et al., 1990; Raush et al., 1990), and ipsapirone (Glaser, 1988). They also exhibit these properties in the following animal models of depression (for review see Robinson et al., 1989): learned helplessness (Giral et al., 1988; Martin et al., 1990a,b, 1991c), forced swimming test (Cervo and Samanin, 1987; Cervo et al., 1988; Wieland and Lucki, 1990; Chojnacka-Wojcik et al., 1991), and open-field test (Kennett et al., 1987a).

These new antidepressants are of great therapeutic interest because they are characterized by a shorter delay of onset of action than that shown by the "classical" antidepressants and because they lack secondary effects. The mechanism of action of these drugs as antidepressants is still unknown. Cervo et al. (1988) proposed that presynaptic 5-HT_{1A} autoreceptors may play a role in the effects of antidepressants. Martin et al. (1990b, 1991c), using the learned helplessness as a paradigm, and local injections of 5-HT_{1A} agonist in the raphe or in the hippocampus, proposed that 5-HT_{1A} postsynaptic receptors were also involved.

As in the case of the anxiolytic properties, the antidepressant activity of these drugs appears to depend on pre- and postsynaptic receptors. The partial or full agonistic activity of the drugs at these receptors sites seems to play a key role in their therapeutic effect. Nevertheless, it seems that, according to the theory of a serotonergic deficit occurring in depression, the antidepressant activity of 5-HT_{1A} agonists would be related to an increase of the serotonergic activity after chronic treatment. New 5-HT_{1A} partial agonists or antagonists are

needed to determine the relative importance of the pre- and postsynaptic effects, that of particular receptor subtypes possibly located in distinct areas, and that of their regulation in the mechanism of their therapeutic activity.

The involvement of the 5-HT system in pain has been documented by several authors (LeBars, 1988). Although it does not seem that 5-HT_{1A} receptors play a major role in pain transmission, the 5-HT_{1A} agonists have been reported to have some analgesic effects (see review by Wilkinson and Dourish, 1991). The latter effect might be explained by the anxiolytic properties of the drugs. Nonetheless, Hamon's group reported the presence of 5-HT_{1A} receptors in the dorsal horns of the spinal cord and proposed that they play a role in pain transmission.

The serotonergic control of the sleep-waking cycle is well established (Jouvet, 1977; Koella, 1988). However, the 5-HT_{1A} receptors do not seem to be functionally important in this phenomenon, although 8-OH-DPAT has been reported to alter the REM sleep latency period (Kafi de St-Hilaire et al., 1987).

B. 5-HT_{1B} Receptor subtype

The 5-HT_{1B} receptor subtype was initially defined as being less sensitive to spiperone than the 5-HT_{1A} subtypes (Pedigo et al., 1981).

1. *Selective radioligands and pharmacological properties.* A marker specific for the 5-HT_{1B}-binding sites has not yet been found; however, these sites were specifically labeled by using [³H]5-HT in the presence of drugs that mask 5-HT_{1A} and 5-HT_{1C} receptors (Peroutka, 1986). They were also labeled with [¹²⁵I]iodocyanopindolol used in combination with isoprenaline, which blocks the β -adrenergic sites (Hoyer et al., 1985a; Offord et al., 1988) and [³H]dihydroergotamine in the presence of phentolamine and 8-OH-DPAT, which block the α -adrenergic and the 5-HT_{1A} sites, respectively (Hamblin et al., 1987). Recently, a small peptide, S-CM-GT NH₂, with a tyrosine residue susceptible to ¹²⁵I iodination was demonstrated to have preferential affinity for 5-HT_{1B} sites in the rat and for 5-HT_{1D} sites in the guinea pig (Boulenguez et al., 1991a,b). The radioactive iodinated form of this ligand was used to label 5-HT_{1B} receptors in the rat brain (Boulenguez et al., 1992). A novel tritiated analog of RU 24969, [³H]CP-96,501, also binds with a high affinity ($K_d = 0.21$ nM) to 5-HT_{1B} sites in rat brain (Koe et al., 1992). The latter, which has greater affinity (30-fold) for 5-HT_{1B} than 5-HT_{1D} receptors, may be a useful tool to discriminate the two receptors.

Only a few drugs are able to bind to 5-HT_{1B} receptors with a nanomolar affinity. These are 5-HT, 5-CT, cyanopindolol, dihydroergotamine, ergotamine, RU 24969, and SDZ(-)21009; yet, none of these is specific to the 5-HT_{1B} site. Other drugs bind to this site with a lower affinity (10 to 1000 nM) (mCPP, quipazine, TFMPP) (table 3) but do not discriminate between it and other receptors; for example, some of them have been reported

to recognize the 5-HT transporter with the same affinity with which they recognize 5-HT_{1B} (Wolf and Kuhn, 1991). CP-93,129, a new tetrahydropyridyl derivative structurally related to the mixed 5-HT_{1A}/5-HT_{1B} receptor ligand RU 24969, was recently reported to be a putative specific 5-HT_{1B} receptor agonist, with >150-fold higher affinity for 5-HT_{1B} versus other 5-HT₁ and 5-HT₂ sites, without interacting with the transporter (Hjorth and Tao, 1991). 5-BT was recently presented as a relatively selective 5-HT_{1B/1D} agent (Peroutka et al., 1991). The final characterization of 5-HT_{1B} receptors awaits the development of selective and potent antagonists.

2. *Regional distribution.* 5-HT_{1B} receptor subtypes occur in rat and mouse brain (Pazos and Palacios, 1985; Hoyer et al., 1985a). These receptors are not observed in guinea pig, cat, pig, calf, pigeon, frog, or humans (Heuring et al., 1987; Hoyer et al., 1986a; Pazos et al., 1988; Martial et al., 1989; Waeber et al., 1989a). Their distribution within the rodent brain is heterogeneous. The highest density of 5-HT_{1B} receptors has been found in substantia nigra, globus pallidus, dorsal subiculum, and superior colliculi.

3. *Cellular localization.* Middlemiss (1984) and Engel et al. (1986) showed that the release of 5-HT from serotonergic terminals was controlled by autoreceptors characterized as 5-HT_{1B}, strongly suggesting that these were homologous presynaptic receptors. However, their postsynaptic location as heterologous presynaptic receptors is also documented by the lack of effect of serotonergic neuron lesions on the number of 5-HT_{1B} receptors (Vergé et al., 1986) and by the existence of a serotonergic inhibition of the ACh release in synaptosomes (Harel-Dupas et al., 1991b).

4. *Molecular structure.* The cloning of the 5-HT_{1B} gene was recently reported in rat by Voigt et al. (1991). The receptor consists of a monomeric chain (386 amino acids) having seven spanning domains and homology with the canine RDC4 receptor (5-HT_{1D}) (63%) and with the 5-HT_{1A} receptor (43%). A new 5-HT_{1D} subtype (5-HT_{1D β}) has been cloned in humans by Weinshank et al. (1992), and its sequence was reported by Adham et al. (1992). The sequence homology of this subtype is 93% with that of the 5-HT_{1B} gene in the rat, suggesting that these two genes are species homologues (Hartig et al., 1992). The pharmacological properties of 5-HT_{1B} and 5-HT_{1D β} receptors expressed in COS-7 cells are quite different (namely, for iodocyanopindolol), although the sequences of the genes are very close (the differences being from 0 to 2 amino acids in each transmembrane region). This suggests that a mutation affecting only a few nucleotides (i.e., Asp-385) plays a very important role in the binding of pindolol derivatives.

In situ hybridization experiments revealed the expression of a high level of 5-HT_{1B} mRNA in raphe, in agreement with the nature of presynaptic autoreceptors. A high density was also observed in areas rich in 5-HT

TABLE 3
Affinity values of drugs for 5-HT_{1B} recognition sites in mammalian brain membranes

Drug	Affinity	Reference	Drug	Affinity	Reference
Very high affinity ($K_1 < 10$ nM)			GR 43175	398	Hoyer and Schoeffter, 1991
(±)ICYP	0.32	Hoyer et al., 1985a	5-MeO-T	398	Schlicker et al., 1989
SDZ(-)21009	0.42	Hoyer et al., 1985a	Methysergide	520	Peroutka, 1986
DHE	0.79	Hoyer and Schoeffter, 1991		1,584	Hoyer and Schoeffter, 1991
RU 24969	0.38	Peroutka, 1986	NAN-190	616	Van Wijngaarden et al., 1990
	3.8	Hoyer et al., 1985a			
	5.8	Van Wijngaarden et al., 1990	MDL 72832	630	Hoyer and Schoeffter, 1991; Van Wijngaarden et al., 1990
	6.8	Hamblin et al., 1987			
(-)CYP	1.8	Engel et al., 1986	5-NH ₂ -T	676	Hoyer et al., 1985a; Engel et al., 1986
SDZ(±)21009	2.95	Engel et al., 1986			
(±)CYP	5	Hoyer and Schoeffter, 1991	2-Me-5HT	724	Van Wijngaarden et al., 1990
	5.4	Schlicker et al., 1989			
5-CT	5.12	Hoyer and Schoeffter, 1991	Flesinoxan	812	Van Wijngaarden et al., 1990
5-CONH ₂ -T	5.13	Engel et al., 1986			
5-HT	2.45	Van Wijngaarden et al., 1990	Cyproheptadine	840	Peroutka, 1986
			Amitriptyline	840	Peroutka, 1986
	7.6	Peroutka, 1986	Very low affinity ($K_1 > 1,000$ nM)		
	11	Hamblin et al., 1987	(±)ICI 118-551	1,230	Hoyer et al., 1985a
	16.5	Schoeffter and Hoyer, 1989b	(±)ICI 89406	2,454	Hoyer et al., 1985a
	24.5	Hoyer and Schoeffter, 1991	Mesulergine	1,148	Van Wijngaarden et al., 1990
High affinity ($K_1 = 10-100$ nM)					
Ergotamine	10.9	Hoyer and Schoeffter, 1991		12,589	Hoyer and Schoeffter, 1991
5-MY	15	Peroutka, 1986		13,200	Hoyer et al., 1985a
(-)Pindolol	15.8	Hoyer and Schoeffter, 1991	Ritanserin	1,737	Van Wijngaarden et al., 1990
	77.6	Hoyer et al., 1985a			
S-CM-GTNH ₂	22.4	Boulenguez et al., 1991a	Ketanserin	1,910	Hoyer and Schoeffter, 1991
Metergoline	25.7	Hoyer et al., 1985a	WB 4101	1,995	Hoyer and Schoeffter, 1991
	39.8	Hoyer and Schoeffter, 1991		4,400	Peroutka, 1986
	40.7	Engel et al., 1986	DOI	2,041	Van Wijngaarden et al., 1990
	52	Peroutka, 1986			
Eltoprazine	25-38	Sijbesma et al., 1990	LY 53857	3,162	Hoyer and Schoeffter, 1991
SDZ(+)-21009	31.6	Hoyer and Schoeffter, 1991	Yohimbine	3,162	Hoyer and Schoeffter, 1991
5-BT	40	Peroutka, 1991a	Phentolamine	4,270	Hoyer et al., 1985a
W-N-CH ₃ -5HT	44.6	Hoyer et al., 1985a	Mianserin	4,680	Hoyer et al., 1985a
(-)Propranolol	46.8	Hoyer et al., 1985a		6,390	Hoyer and Schoeffter, 1991
	50.1	Hoyer and Schoeffter, 1991		10,000	Peroutka, 1986
TFMPP	45	Peroutka, 1986	Spiperone	4,790	Hoyer et al., 1985a
	48	Van Wijngaarden et al., 1990		5,011	Hoyer and Schoeffter, 1991
				5,370	Engel et al., 1986
	131	Schoeffter and Hoyer, 1989	(+)Pindolol	5,128	Engel et al., 1986
	398	Hoyer and Schoeffter, 1991	Cinanserin	6,200	Peroutka, 1986
Methiothepin	50	Hoyer and Schoeffter, 1991		6,309	Hoyer and Schoeffter, 1991
Bufotenine	52	Peroutka, 1986		10,000	Hoyer et al., 1985a
(+)CYP	56	Engel et al., 1986	5,6-DHT	6,165	Engel et al., 1986
(+)Propranolol	85.1	Engel et al., 1986	MK 212	9,332	Engel et al., 1986
Low affinity ($K_1 = 100-1,000$ nM)			DP-5-CT	12,581	Hoyer and Schoeffter, 1991
Isamoltane	112	Schoeffter and Hoyer, 1989b	8-OH-DPAT	1,778	Van Wijngaarden et al., 1990
CGS 12066B	114	Schoeffter and Hoyer, 1989b			
Sumatriptan	154	Van Wijngaarden et al., 1990		24,000	Peroutka, 1986
				60,100	Hoyer and Schoeffter, 1991
d-LSD	151	Engel et al., 1986	Ipsapirone	3,548	Van Wijngaarden et al., 1990
	170	Peroutka, 1986			
Methergine	199	Hoyer and Schoeffter, 1991		52,000	Peroutka, 1986
Lisuride	199	Hoyer and Schoeffter, 1991		125,892	Hoyer and Schoeffter, 1991
mCPP	251	Hoyer and Schoeffter, 1991	Buspirone	3,019	Van Wijngaarden et al., 1990
	338	Schoeffter and Hoyer, 1989b			
Quipazine	260	Peroutka, 1986		68,000	Peroutka, 1986
	316	Hoyer and Schoeffter, 1991		125,890	Hoyer and Schoeffter, 1991
	309	Schoeffter and Hoyer, 1989	Spiroxatrine	125,892	Hoyer and Schoeffter, 1991
5-MeO-DMT	390	Peroutka, 1986	Gepirone	125,893	Hoyer and Schoeffter, 1991

nerve terminals, such as hippocampus (CA1), cerebellum, caudate putamen, and layer IV of the cortex, in agreement with the nature of heterologous presynaptic receptors. However, this mRNA was not found in globus pallidus and substantia nigra, areas that contain the greatest density of 5-HT_{1B} receptors. This might be the result of the expression of the genes in the cell body of the neuron outside the globus pallidus and the migration of the receptor protein toward the terminals inside this region.

5. Transduction system. The 5-HT_{1B} receptors are negatively coupled to adenylyl cyclase. In rat substantia nigra, 5-HT_{1B} agonists inhibit forskolin-stimulated adenylyl cyclase activity (Bouhelal et al., 1988; Schoeffter and Hoyer, 1989a). In hamster cultured fibroblasts, such agonists inhibit the enzyme activity stimulated by cholera toxin (Seuwen et al., 1988). However, surprisingly, the autoreceptor present on serotonergic terminals in the hippocampus is not coupled to G_i, G_o, or G_o proteins as shown by the lack of interaction with pertussis toxin, cholera toxin, and *N*-ethyl-maleimide (Blier, 1991). These results suggest that 5-HT_{1B} receptor-coupling mechanisms are heterogeneous.

6. Regulation. As in the case of 5-HT_{1A} receptors, the 5-HT_{1B} sites are modulated by GTP and its nonhydrolysable analogs. In rat brain (Hamblin et al., 1987; Stratford et al., 1988; Ariani et al., 1989) and in mouse liver (Ciaranello et al., 1990), the presence of guanine nucleotides shifts the equilibrium to the nonbinding state of the site (see section III.A.7).

To our knowledge, the pre- or postnatal development of 5-HT_{1B} receptors has not been extensively studied. Voigt et al. (1991) examined the developmental expression of 5-HT_{1B} mRNA. They showed that no quantitative changes were observed during development in most brain areas; however, in caudate-putamen, thalamus, and cerebellum, the high density of the mRNA observed at P0, P6, and P12 strongly decreased at the adult age. A moderate decrease in 5-HT_{1B} receptor number has been observed in the caudate nucleus (−27%) and in the substantia nigra (−20%) between 3 and 22 months in rats (Gozlan et al., 1990).

The regulation that occurs following serotonergic neuronal degeneration induced by 5,7-DHT is not yet well understood; indeed, the number of 5-HT_{1B} sites is not modified in rat brain except in the pars reticulata in the substantia nigra, where it increases (Weissmann et al., 1986) or decreases (Vergé et al., 1986). When nigrostriatal fibers degenerate after intrastratial injection of kainic acid, 5-HT_{1B} sites decrease in the substantia nigra; this result indicates that 5-HT_{1B} sites are located on the terminals of nonserotonergic fibers originating from the striatum (Palacios and Dietl, 1988; Hamon et al., 1990b). Accordingly, the related adenylyl cyclase inhibitory activity is also reduced (Hamon et al., 1990b).

Antidepressants interact with the 5-HT_{1B} receptor

function. In the rat, lithium interferes with regulation of 5-HT release via the inactivation of the hippocampal 5-HT_{1B} receptors (Hide and Yamawaki, 1989). In rat hippocampal slices or synaptosomes, the inhibition of the K⁺-evoked ACh release is mediated by 5-HT_{1B} receptors; antidepressants at low concentrations, in vitro, partially reverse this inhibitory effect (Bolanos and Fillion, 1989; Harel-Dupas et al., 1991a). Additionally, the effect of TFMPP on 5-HT_{1B} sites, which reduces the "isolation-induced social behavior deficit," is antagonized by chronic treatment with antidepressant (fluoxetine or phenazine), suggesting the existence of a link between these antidepressants and the 5-HT_{1B} receptor function (Francès and Khidichian, 1990). Using an animal model of depression (learned helplessness), Edwards et al. (1991c) demonstrated that the number of 5-HT_{1B} receptors increased in the cortex, the hippocampus, and the septum but decreased in the hypothalamus. However, Martin and Puech (1991), using the same experimental model, did not obtain similar results. Furthermore, Montero et al. (1991) did not find any modification in the number of 5-HT_{1B}-binding sites after acute, chronic, or even prenatal treatment with antidepressants (chlorimipramine, tianeptine, iprindole).

These results do not clearly demonstrate the involvement of 5-HT_{1B} receptors in depression or in the mechanism of action of antidepressants. However, in the experimental model of depression, the number of 5-HT_{1B} sites is increased in serotonergic areas, in agreement with the observation that, in vitro, antidepressants antagonize the cellular functional effect of the receptors. Whether these receptors, or their equivalent in humans, are significantly involved in depression remains to be established.

7. Cellular functions. A major function of the 5-HT_{1B} receptors is the control of 5-HT release from the serotonergic neuron terminals (presynaptic homologous autoreceptors), as demonstrated by Middlemiss (1984), Engel et al. (1986), Göthert et al. (1987), and Chaput and de Montigny (1988b). These receptors are also located on nonserotonergic terminals (presynaptic heterologous receptors) where they inhibit the release of other neurotransmitters, i.e., ACh in rat hippocampus (Maura and Raiteri, 1986; Maura et al., 1989; Harel-Dupas et al., 1991b), noradrenaline in peripheral tissues (Molderings et al., 1987, 1990; Molderings and Göthert, 1990), and possibly dopamine, as shown in vivo by microdialysis (Benloucif and Galloway, 1991). They also stimulate the release of prolactin in the rat (Van de Kar et al., 1989a,b). Thus, it appears that the 5-HT_{1B} receptors may play an important role in the control of CNS function by modulating the release of other neurotransmitters at the level of the nerve terminal.

Serotonin stimulates the proliferation of vascular smooth muscle cells in synergy with platelet-derived growth factor and insulin (Nemecek et al., 1986). Its role

as a regulator of neuronal growth in fetal tissue has been proposed on the basis of several experimental studies (Lauder and Krebs, 1978; Haydon et al., 1984; Chubakov et al., 1986; Whitaker-Azmitia et al., 1987; see review of CNS by Hamon et al., 1989). Seuwen et al. (1988) showed that 5-HT_{1B} receptors were directly involved in the mitogenic effect that 5-HT has on hamster cultured fibroblasts (CCL39). This effect is sensitive to pertussis toxin and, therefore, depends on coupling with a G_i protein; however, it does not seem that the production of cAMP or IP₃ is modified (Seuwen and Pouyssegur, 1990). Thus, the 5-HT_{1B} receptors that are involved in mitogenesis may have a transduction mechanism different from those involved in cAMP or IP₃ production. Although it is plausible that 5-HT_{1B} receptors play a role in cell proliferation, in the CNS or during synaptogenesis this remains to be demonstrated.

8. 5-HT_{1B} receptors and behavior. The absence of 5-HT_{1B} receptors in humans and many other species limits behavioral studies to mice and rats. Moreover, these investigations are hampered by the absence of selective ligands. Despite these restrictions, a number of results have been reported. TFMPP, mCPP, and RU 24969 decrease body temperature (Tricklebank et al., 1986) and food intake (Bendotti and Samanin, 1987; Kennett et al., 1987b). These effects probably involve receptors located on nonserotonergic neurons because the activity is still observed after pCPA administration which depletes the cells of 5-HT. It has also been proposed that fenfluramine, sertraline, and fluoxetine interact with feeding behavior through 5-HT_{1B} receptors, although this hypothesis is far from being proven (Wilkinson and Dourish, 1991).

The role of 5-HT_{1B} receptors in rat sexual behavior has been studied with various agonistic drugs which, unfortunately, lack specificity; an inhibitory effect is generally observed and may be exerted through the 5-HT_{1B} receptors; this effect is opposite to that of the 5-HT_{1A} presynaptic activity (Berendsen and Broekkamp, 1987; Fernandez-Guasti et al., 1989). How 5-HT_{1B} receptors participate in rat sexual behavior is not yet totally understood.

TFMPP, RU 24969, CGS 12066B, and mCPP reduce stress in the isolation-induced social deficit model (Francès, 1988a,b; Francès and Monier, 1991) and aggressiveness in a model of social interaction (Olivier et al., 1989). Therefore, these results suggest that 5-HT_{1B} receptors play a role in the behavior studied in these experimental paradigms. Moreover, in an animal model of depression (learned helplessness), these agonistic drugs inhibit the effect of serotonin reuptake inhibitors (Martin and Puech, 1991); in the forced swimming test, they reduce the duration of immobility and antagonize the effects of antidepressants (Cervo et al., 1989). In light of the recent finding that 5-HT_{1B} agonists interact at nanomolar concentrations with the 5-HT transport sys-

tem, the observed effects would appear to be independent of the 5-HT_{1B} receptor function. More specific 5-HT_{1B} receptor agonists or antagonists are needed to further elaborate the participation of 5-HT_{1B} receptors in these experimental models.

C. 5-HT_{1D} Receptor Subtype

The existence of the 5-HT_{1D} receptor subtype was proposed by Heuring and Peroutka (1987) on the basis of binding studies performed with bovine brain using [³H]5-HT labeling in the presence of drugs masking 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} subtypes; the remaining binding was called 5-HT_{1D}.

1. Selective radioligands and pharmacological properties. The [³H]5-HT binding observed under the experimental conditions generally used strictly represents 5-HT_{1nonA, nonB, nonC} binding, because it is observed with drugs that selectively mask 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} sites. This binding is limited mainly to 5-HT_{1D} sites but also may include other sites having the same high affinity for 5-HT and a different affinity for particular ligands. Until recently, no ligand specific for 5-HT_{1D} was available; Bruinvels et al. (1991) reported that serotonin-5-O-carboxymethylglycyl-¹²⁵I-tyrosinamide labeled a homogeneous population of 5-HT_{1D} sites in human substantia nigra. This radiolabeled-ligand has been used for a direct visualization of 5-HT_{1D} receptors in human brain (Palacios et al., 1992).

The 5-HT_{1D} subtype is characterized by its high affinity (<10 nM) for 5-HT, 5-CT, 5-MeO-T, metergoline, and ergometrine. It has some affinity (10 to 100 nM) for 5-BT, *d*-LSD, methiothepin, ergotamine, RU 24969, yohimbine, and methysergide (table 4). However, none of these drugs is specific for this receptor. GR 43175 (sumatriptan) has been presented as one of the best selective agonists for 5-HT_{1D} sites ($K_d = 17$ nM) (Waeber et al., 1989d; Schoeffter and Hoyer, 1989b; Peroutka and McCarthy, 1989; Peroutka, 1991a); this drug, however, recognizes 5-HT_{1B} sites with a similar affinity ($K_d = 27$ nM) and 5-HT_{1A} sites with a slightly lower affinity ($K_i = 100$ nM) (Peroutka and McCarthy, 1989). Other authors have reported lower affinities of 5-HT_{1D} for this drug: $K_d = 68$ nM (Van Wijngaarden et al., 1990) and 251 nM (Sumner and Humphrey, 1989).

Is the 5-HT_{1D} receptor the sole 5-HT_{1nonA, nonB, nonC} subtype? The experimental conditions used to observe the so-called 5-HT_{1D}-binding site, in fact, revealed all binding sites other than 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C}, because [³H]5-HT binding was measured in the presence of drugs masking 5-HT_{1A}, 5-HT_{1B}, and 5-HT_{1C} receptors. Therefore, the remaining binding, whether homogeneous or heterogeneous, may represent not only 5-HT_{1D} receptors but also other binding subtypes that are not discriminated by 5-HT itself.

The first evidence for a 5-HT_{1D} heterogeneity was given by Waeber et al. (1988c), who examined [³H]5-HT

TABLE 4
Affinity values (K_i) of drugs for different 5-HT_{1nonA,nonB,nonC} recognition sites in mammalian brain membranes

Drugs	5-HT _{1D} nonmurides species				"5-HT _{1D} " rat cortex striatum	5-HT _{1R} human cortex	5-HT _{1nonA,nonB,nonC} rat cortex	5-HT _{1R} rabbit caudate
	Heuring et al. (1987)	Hoyer and Schoeffter (1991)	Bruinvels et al. (1991)	Herrick-Davis (1989)	Herrick-Davis and Titeler (1988b)	Leonhardt et al. (1989)	Fayolle et al. (1988)	Xiong and Nelson (1989)
<10 nM								
5-HT	3.2	3.98	6.16	1.9–2.6	2.1–1.8	3.5	2.5	10.96
5-CT	0.75	2.5	0.6	0.8–2.9	48–33	910	122.5	
5-MeO-T	2.5	7.07		4.1–11	70–72		138	
Metergoline	4.2	0.79	7.76					57.54
Ergometrine		0.319		0.9				
Sumatriptan			4.67					
10–100 nM								
Ergotamine		15.8	7.41	0.9		155		
DHE		19.9	10.96				202.5	
d-LSD	24	6.3		0.7–3.3	10–7.7		44.5	
RU 24969	25	50					800	69
Methiothepine	36	50.1	70.8				2,150	169.8
Tryptamine	36	39.8						
Yohimbine	39	79	15.48					
DP-5-CT		63						
GR 43175		31						
Lisuride		36.6						
100–1,000 nM								
Cyanopindolol		125						
Methysergide	120	3.9		8.8–15	41–100	59	10,000	120
N-N-DMT	190							
TFMPP	280	630		250–450	520–220	570	2,680	69
Mianserine	480	398	338				1,260	
WB 4101	590	316						
8-OH-DPAT	700	1,258	28	200–520	820–870	826	1,010	562
Quipazine	800	1,258						
MDL 72832		398						
SDZ(-)21009		398						
CGS 12066 B		446	8.7					
<1,000 nM								
Cyproheptine	1,200							
mCPP	1,400	1,584						
Mesulergine	6,500	6,309		1,200–1,800	3,200–2,800		5,000	7,244
(-)pindolol	7,000	6,309	954		>10,000			
(-)propranolol	7,200	3,162						
Haloperidol	8,200							
Metoclopramide	9,000							
Spiroxitrine		7,943						
Ipsapirone	16,000	12,589	1,905	5,200–9,200	1,100–1,500		4,950	
Buspirone		31,622	19,054					
Ketanserine	53,000	1,000		3,200–6,200	>10,000	>10,000	8,500	
Ritanserine		1,584					727	
Siperone	>100,000	5,011						2,754
MDL-72222	>100,000			>10.00	>10,000		>10,000	
(+)DOB				970–1,300	3,400–1,800	556		
CP 93129			954					
Isamoltane			1,174					

binding in the presence of 8-OH-DPAT (100 nM) and mesulergine (100 nM); the displacing curves presented for 5-CT in bovine, porcine, and human caudate membranes were clearly biphasic, which would correspond to two populations of receptors representing about 50% each. Sumner and Humphrey (1989), using newborn pig caudate, confirmed that 5-CT found two subpopulations of the 5-HT_{1nonA nonB nonC} binding at about the same ratio and demonstrated that the high-affinity component for

5-CT was selectively displaced by sumatriptan ($K_i = 251$ nM). This particular property led Peroutka (1991a), using calf, guinea pig, pig, human cortex and caudate, to define the 5-HT_{1D} sites as those binding [³H]5-HT in the presence of 8-OH-DPAT (100 nM) and mesulergine (100 nM) and displaced by 10⁻⁵ M sumatriptan. Moreover, Mahle et al. (1991), using [³H]5-CT binding in the presence of 100 nM 8-OH-DPAT and mesulergine, demonstrated in the guinea pig brain that the high-affinity binding for 5-

CT was heterogeneous, indicating the existence of two compartments having high ($K_i = 27$ nM) and low affinity ($K_i = 400$ nM) for sumatriptan in the cortex; this heterogeneity was not observed in the striatum.

The heterogeneity of the 5-HT_{1nonA nonB nonC} binding was also studied by Leonhardt et al. (1989) using pindolol (1000 nM) and mesulergine (100 nM); these authors confirmed and extended the findings previously obtained with human cortex, showing high- and low-affinity subpopulations for 5-HT ($K_i = 4$ and 155 nM, respectively). The 5-HT low-affinity component (50% of the binding) was called 5-HT_{1E}, and its pharmacological properties were examined; no drug was found to have a high affinity for this site, except 5-HT itself ($K_d = 3.5$ nM) and methysergide ($K_i = 59$ nM). Interestingly, this binding was GTP dependent, suggesting that it might be related to a G protein. The regional distribution of 5-HT_{1E} is heterogeneous within the brain and initially reported to parallel that of 5-HT_{1D} (Palacios et al., 1991). However, Lowther et al. (1991) showed, in postmortem human brain samples, that the 5-HT_{1D}/5-HT_{1E} ratio differed depending on the brain region; the proportion of 5-HT_{1D} was high in globus pallidus and that of 5-HT_{1E} was high in caudate-putamen. Moreover, different regional distributions were also demonstrated for the 5-HT_{1E} binding, indicating a high proportion of these sites in cortex versus striatum (Mahle et al., 1991). Whether the 5-HT_{1E} site is a true functional receptor remains to be established.

The existence of another receptor, 5-HT_{1R}, different from 5-HT_{1D} in the strictest sense, which is not 5-HT_{1A}, 5-HT_{1B} or 5-HT_{1C}, was reported by Xiong and Nelson (1989), who observed [³H]5-HT binding in the presence of 8-OH-DPAT (100 nM) and mesulergine (100 nM) in the rabbit caudate nucleus. The affinities of 5-HT_{1R} for methiothepin, methysergide, and spiperone differ from those of 5-HT_{1D}. However, 5-HT and sumatriptan were not examined in these studies, so the differences or similarities between 5-HT_{1R} and 5-HT_{1E} are still not known.

On the other hand, in rat brain cortex, Fayolle et al. (1988), using spiperone (1000 nM) and propranolol (3000 nM), reported the existence of a 5-HT_{1nonA nonB nonC} binding characterized by pharmacological properties different from those of 5-HT_{1D}. In particular, the site they found has a low affinity for 5-HT ($IC_{50} = 245$ nM). The functional nature of this site was indicated by the pharmacological correlation of its binding with a positive coupling to adenylyl cyclase activity. Moreover, the regulation of this receptor was studied during postnatal development in the rat brain cortex (Zifa et al., 1988). This receptor develops mainly during synaptogenesis (the second and third week after birth), and a significant activity is already present at birth. This receptor may be involved in the mechanism that functionally links the CNS and the immune system; rabies virus infection in

rats induces a rapid and marked decrease in the density of the 5-HT_{1nonA nonB nonC} receptors in rat brain cortex (Fillion et al., 1990).

A further indication of the heterogeneity of 5-HT_{1D} receptors, depending on the region or the species considered, is the variation in the abilities of different drugs to compete for 5-HT_{1D} binding in calf, pig, guinea pig, and human cortex and caudate (Peroutka, 1991a).

In conclusion, the 5-HT_{1nonA nonB nonC} binding corresponds not only to 5-HT_{1D}, in the strictest sense, but also to other sites. Some of these sites may represent functional receptors belonging to a 5-HT_{1D} family.

2. Regional distribution. 5-HT_{1D} sites have been found in the brain of the following species (for reviews, see Waeber et al., 1990a): pigeon (Waeber et al., 1989a), hamster (Beer et al., 1992), guinea pig (Waeber et al., 1989a; Limberger et al., 1991), rabbit (Limberger et al., 1991), dog (Beer et al., 1992), pig (Waeber et al., 1988c; Sumner and Humphrey, 1989), calf (Waeber et al., 1988c; Heuring and Peroutka, 1987), monkey (Waeber et al., 1989c), and humans (Waeber et al., 1988a,c; Hoyer et al., 1988; Herrick-Davis and Titeler, 1988b; Palacios et al., 1992). In these species, 5-HT_{1B} sites are absent. It has been proposed that 5-HT_{1D} plays the same role in these species as does 5-HT_{1B} in rat, mouse, and opossum. This proposition is based not only on the structural homology between these two subtypes but also on the similarities between their pharmacological properties, regional distribution, transduction systems, and cellular functions as homologous or heterologous presynaptic receptors (Hoyer and Middlemiss, 1989). However, it is likely that 5-HT_{1D} sites also exist in the rat brain (Herrick-Davis and Titeler, 1988b; Limberger et al., 1991) or the rat periphery (Berendsen and Broekkamp, 1991a,b). Further studies are needed to clarify the situation.

The regional distribution of 5-HT_{1D} sites in the mammalian brain is heterogeneous. In the substantia nigra, basal ganglia, and striatonigral pathway (Waeber et al., 1990b), these sites are dense, accounting for 90% of the 5-HT₁ binding (Waeber and Palacios, 1990). A lower density is observed in the hippocampus, cortex, and raphe. 5-HT_{1D} receptors also have been identified in porcine coronary artery (Schoeffter and Hoyer, 1990) and in canine basilar artery (Connor et al., 1989). The presence of these receptors in the vascular bed may be responsible for some pharmacological properties of 5-HT_{1D} agonists (see section III.C.8).

3. Cellular localization. The localization of 5-HT_{1D} receptors has been shown to be mostly postsynaptic on nonserotonergic terminals. Indeed, quinolinic acid lesion of striatal cellular bodies induces a decrease of 5-HT_{1D} receptors in substantia nigra and in globus pallidus, indicating their localization on nonserotonergic terminals (presynaptic heterologous). However, the fact that their numbers decrease also in striatum under these experimental conditions suggests their possible location

either on cell bodies, or on dendrites, or on terminals of interneurons (Waeber et al., 1990b). Moreover, Waeber and Palacios (1990) demonstrated in guinea pig the presynaptic localization of 5-HT_{1D} receptors on nonserotonergic terminals in superior colliculi using unilateral enucleation. In humans, binding experiments on samples from brain tissue from patients with Huntington's chorea, where the striatal intrinsic neurons degenerate, demonstrated a decrease in 5-HT_{1D} receptor number in this area, whereas the [³H]paroxetine binding, which labels the serotonergic terminals, was not affected (Waeber and Palacios, 1989; Gonzalez-Heydrich and Peroutka, 1991). The location of 5-HT_{1D}-binding sites on serotonergic terminals is likely, although it has not yet been demonstrated on the basis of lesion experiments; however, functional evidence indicates that some of these receptors are homologous presynaptic receptors.

4. Molecular structure. Hamblin and Metcalf (1991) recently cloned the human 5-HT_{1D} receptor (MA6A clone) by using a probe consisting of canine thyroid cDNA (RDC4), obtained by polymerase chain reaction (Libert et al., 1989). This clone carries an intronless gene encoding 377 amino acids; the seven hydrophobic domains of the protein are characteristic of G protein-linked receptors. The amino-terminal region contains three glycosylation sites, the second extracellular loop contains one, and the third intracellular loop (83 amino acids) possesses two cAMP-dependent protein kinase sites and other potential phosphorylation sites, such as serine and threonine residues. The COOH-terminal region is very short (19 residues) and lacks the cysteine residue present in many members of this family. The amino acid sequences for MA6A and RDC4 have the same length and are 88% identical. The most probable localization for the RDC4 gene is at the 1p34.3-1p36.3 region of the human genome (Libert et al., 1991).

Among the sequences that have been reported for members of the G protein-linked receptor family, the one most similar to that of the 5-HT_{1D} receptor is that of the 5-HT_{1A} receptor (43% deduced amino acid identity); the similarity between the human 5-HT_{1D} receptor and other serotonin receptors is less marked, i.e., 37% for the *Drosophila* 5-HT_{dro} receptor, 33% for the rat 5-HT_{1C}, and 31% for the rat 5-HT₂.

Expressed in COS-7 cells, the 5-HT_{1D} human gene induces the production of a protein that binds serotonergic drugs with the following order of potency: 5-CT > 5-MeOT = 5-HT > RU 24969 > yohimbine > TFMPP > 8-OH-DPAT > mesulergine = spiperone = ICS 205-930. That the affinities of dihydroergotamine and metergoline are higher for this cloned receptor than for those in human membranes, and the existence of other minor pharmacological differences, raises various questions, e.g., whether different 5-HT_{1D} subtypes exist (Hamblin and Metcalf, 1991). This hypothesis was confirmed because a second gene also expressing a 5-HT_{1D} receptor

has been cloned (5-HT_{1Dβ}) (Weinshank et al., 1992). Its sequence, presented by Adham et al. (1992), is distinct from the 5-HT_{1D} gene presented by Hamblin and Metcalf (1991), termed 5-HT_{1Dα}, the sequence homology being only 77% in the membrane-spanning domains; surprisingly, the pharmacological properties are identical. The 5-HT_{1Dβ} human gene closely resembles the rat 5-HT_{1B} gene (Adham et al., 1992), although they code for receptors having very different pharmacological properties (i.e., the pindolol derivatives have a high efficacy for the sole 5-HT_{1B} receptors).

Therefore, it appears, on the one hand, that two different genes code for receptors having very similar pharmacological properties (5-HT_{1Dα} and 5-HT_{1Dβ}) and represent intraspecies differences and, on the other hand, that two very similar genes code for receptors having different pharmacological properties (5-HT_{1Dβ} and 5-HT_{1B}) and represent interspecies variations. Thus, the 5-HT_{1D} receptors appear as a multigenic family with different members in the same species, i.e., 5-HT_{1Dα} and 5-HT_{1Dβ} in humans and 5-HT_{1Dα} (Hartig et al., 1992) and 5-HT_{1B} in rodents. Two partial serotonin 5-HT_{1D} receptor sequences have been cloned in the mouse, suggesting the existence of two additional receptors of the 5-HT_{1D} type, i.e., different from those already discussed (Weydert et al., 1992). Because the 5-HT_{1B} receptor in rodents and the 5-HT_{1D} receptor in the other species have the same functional profile (Hoyer and Middlemiss, 1989), their respective roles have to be defined more closely if they actually are present in the same species.

The existence of this large family of receptor subtypes present in a single species may correspond to specific function, particular regional or cellular distribution, and/or particular mechanisms of regulation of a given receptor. The fact that different receptors present similar or different pharmacological characteristics raises the problem of the validity of the pharmacological classification and that of the pharmacological screening models. One of the final goals of this research will be to determine the functional role of each of these subtypes.

5. Transduction system. The 5-HT_{1D} receptor appears to be negatively coupled to adenylyl cyclase activity in calf substantia nigra, as demonstrated by the inhibition of the forskolin-stimulated enzyme activity (Hoyer and Schoeffter, 1988; Schoeffter et al., 1988). In other brain areas, such as the striatum, negative coupling was not found (Waeber and Palacios, 1990). These authors explained the lack of activity as being due to either a "dilution" of the effect by the high basal adenylyl cyclase activity in this area or the coupling of the 5-HT_{1D} receptor to G proteins different from the G_i. It is also possible that 5-HT_{1D} receptors are heterogeneous in the brain.

The transduction system was also examined in transfected cells (Chinese hamster ovary-K1); it consisted of a pertussis toxin-dependent negative coupling with adenylyl cyclase (Hamblin and Metcalf, 1991).

6. *Regulation.* Like all receptors linked to G proteins, 5-HT_{1D} receptors are modulated by GTP in two binding states which are presumably related to their functional activity (Herrick-Davis and Titeler, 1989; Harrington and Peroutka, 1990b).

The density of 5-HT_{1D}-binding sites was studied by quantitative autoradiography in brain slices collected postmortem from patients with Parkinson disease and Huntington's chorea (Waeber and Palacios, 1989; Gonzalez-Heydrich and Peroutka, 1991); no change in the density of 5-HT_{1D} receptors was observed in brains from patients with Parkinson's disease, whereas a significant decrease occurred in the basal ganglia and substantia nigra of brains from patients with Huntington's chorea.

7. *Cellular functions.* The role of the 5-HT_{1D} receptor as an autoreceptor that is situated on 5-HT terminals and that inhibits the release of serotonin is well documented for guinea pig (Middlemiss et al., 1988; Schipper and Tulp, 1988), pig (Schlicker et al., 1989), and humans (Galzin et al., 1989). This receptor is also present on nonserotonergic terminals where it regulates the K⁺-evoked release of ACh in the guinea pig hippocampus (Harel-Dupas et al., 1991a) and noradrenaline in the human saphenous vein (Molderings et al., 1990).

8. *5-HT_{1D} receptors and behavior.* It is difficult to determine the role of 5-HT_{1D} receptors in behavior as long as there is no specific agonist or antagonist. Recently, a novel serotonergic agent, sumatriptan (formerly called GR 43175), was found to be a selective 5-HT_{1D} receptor agonist. This drug was reported to be extremely effective in the acute treatment of migraine and to have minimal side effects (Doenicke et al., 1988; Perrin et al., 1989; Tfelt-Hansen et al., 1989; Ferrari et al., 1989; Byer et al., 1989; Ferrari, 1991). 5-HT has an important role in the pathophysiology of the migraine believed to result from excessive dilation of cerebral blood vessels (Anthony, 1986). Sumatriptan, which does not penetrate the blood-brain barrier (Sleight et al., 1990; Deliganis and Peroutka, 1991), blocks this phenomenon through 5-HT₁-like receptors mediating the constriction of arteriovenous anastomosis (Humphrey et al., 1990; Peroutka, 1991b,c).

The 5-HT₁-like receptors are identified as 5-HT_{1D} receptors in vascular models, such as the relaxation of pig coronary arteries (Schoeffter and Hoyer, 1990) or the contraction of human saphenous vein (Bax et al., 1992). The 5-HT₁-like receptors controlling the porcine carotid circulation, however, does not seem to be identical with the 5-HT_{1D} subtype (Den Boer et al., 1992). If the 5-HT₁-like receptor is in fact the 5-HT_{1D} receptor, then this receptor may prove to be an important target in the acute treatment of the migraine.

The contralateral rotation induced by an intracerebral, unilateral local infusion of 5-HT into the guinea pig substantia nigra constitutes an experimental animal model that is claimed to selectively involve 5-HT_{1D} re-

ceptors (Higgins et al., 1991b). The locomotor changes observed in this model may be analogous to those observed in patients with Huntington's chorea, who exhibit a selective decrease in 5-HT_{1D} receptors in the caudate. The definitive classification of this behavioral response will require more specific agonists and antagonists to become available.

The "hindlimb scratching" induced by systemic injection of serotonergic compounds in rats is thought to involve peripheral 5-HT_{1D}-like receptors (Berendsen and Broekkamp, 1991a). This behavior decreases with age (Berendsen and Broekkamp, 1991b). Intracerebroventricular administration of sumatriptan induces hypothermia and hyperlocomotion in mice (Bill et al., 1989). Because sumatriptan also has a high affinity for 5-HT_{1B}, and 5-HT_{1A} receptors, further pharmacological characterization is needed to attribute these effects solely to the 5-HT_{1D} receptors. The role of 5-HT_{1D}-like receptors in rodents remains to be established.

9. *Conclusion.* The existence of 5-HT_{1D} receptors is now well established. The use of sumatriptan has allowed this receptor subtype to be more precisely characterized. The existence of other sites or receptors, resembling 5-HT_{1D} insofar as they are 5-HT_{1nonA}, _{nonB}, _{nonC}-binding sites, has been established. Specific ligands are needed to characterize the molecular, cellular, and functional properties of these receptors.

IV. 5-HT₂ Receptors

The 5-HT₂ receptor was initially identified as a site that could bind [³H]spiroperidol and that had specific serotonergic pharmacological properties (Leysen et al., 1978). Two subtypes have been reported for the 5-HT₂ family. Recent pharmacological and molecular biological studies strongly suggest that the 5-HT_{1C} receptor subtype (originally classified as a 5-HT₁-like site) should be included in the 5-HT₂ receptor family. The members of the 5-HT₂ receptor family are characterized by their high affinity for serotonergic antagonists, such as cyproheptadine, mesulergine, methysergide, mianserin, metergoline, and pizotifen.

A. "Classical" 5-HT₂ Receptor

The functional correlates of 5-HT₂ binding were recognized early because of the availability of high-affinity antagonists. Numerous experimental works concerning this subject have been reviewed (Lyon and Titeler, 1988; Göthert, 1990; Leysen, 1990). Therefore, the present review will be limited to recent studies.

1. *Selective radioligands and pharmacological properties.* In initial studies, [³H]spiperone was used to label the 5-HT₂-binding site (Leysen et al., 1978; List and Seeman, 1981). Other radioligands were demonstrated thereafter to recognize this site: [³H]mianserin (Peroutka and Snyder, 1981), [³H]metergoline (Hamon et al., 1981), [³H]ketanserin (Leysen et al., 1982b), [³H]mesulergine (Closse, 1983), ¹²⁵I-LSD (Hartig et al., 1983; Engel et al.,

1984; Nakada et al., 1984), ^{125}I -N¹-methyl-2-*l*-LSD (Hoffman et al., 1985, 1987), [^3H]N-methyl-spiperone (Lyon et al., 1986), [^3H]7-amino-ketanserin (Wouters et al., 1986a) and [^{125}I]7-amino-8-iodo-ketanserin (Wouters et al., 1986b; Schotte and Leysen, 1989). These radioligands were antagonists; only recently have agonistic ligands been developed: [^3H]DOB (Titeler et al., 1985, 1987), [^{77}Br]DOB (Wang et al., 1988; Peroutka et al., 1988) and [^{125}I]DOI (McKenna and Peroutka, 1989).

There is controversy regarding the nature of agonist binding to 5-HT₂ receptor. Various radioligand-binding studies were interpreted as demonstrating the existence of two distinct 5-HT₂ sites, a 5-HT_{2A} receptor possessing a high affinity for agonists and antagonists and a 5-HT_{2B} receptor possessing a low affinity for agonists and a high one for antagonists (Pierce and Peroutka, 1989); other investigators proposed the existence of two affinity states for the same 5-HT₂ receptor, corresponding to high and low affinities for the agonists (Lyon et al., 1987; Leonhardt and Titeler, 1989). For example, the antagonistic drugs recognize [^3H]ketanserin and [^{77}Br]DOB sites with the same affinity, whereas the agonists recognize [^3H]DOB sites with an affinity 200 times higher than that for [^3H]ketanserin sites (McKenna and Peroutka, 1989). Moreover, it was shown that only part of the [^3H]ketanserin-binding sites was labeled by 5-HT₂ agonists (19% in rat cortex) (Lyon et al., 1987; Peroutka et al., 1988) and that 5-HT displaced [^3H]ketanserin with a shallow curve (slope of the curve corresponding to a Hill coefficient = 0.67) in rat and human cortical membranes (Pierce and Peroutka, 1989; McKenna and Peroutka, 1989). These data suggest that [^3H]ketanserin labels more than one population of 5-HT₂ receptors. By contrast, 5-HT competition curves in bovine cortex present a Hill coefficient of 0.98, suggesting that [^3H]ketanserin labels a single site; this result is in agreement with the finding that [^3H]DOB cannot be detected in bovine membranes. However, to define distinct receptor subtypes on the basis of agonist-binding data in different species can be dangerous (Leysen, 1990; Strange and Peroutka, 1990).

To provide more evidence in favor of a two-state hypothesis or a two-receptor hypothesis, Hartig et al. (1990a) and Teitler et al. (1990) used the cloned human or rat 5-HT₂ receptor. They demonstrated, by transfecting mammalian fibroblasts (NIH 3T3) and green monkey kidney cells (COS-7), that a single 5-HT₂ gene expressed a protein that bound both agonists and antagonists. Interestingly, [^3H]DOB sites represented only approximately 30% of the density of [^3H]ketanserin labeled binding sites. These data suggest that the 5-HT₂ receptor gene product contains both the agonist- and the antagonist-binding sites, supporting the hypothesis of the existence of two distinct, but noninterconverting, states which differ in their binding characteristics. However, posttranslational receptor changes resulting in two

slightly different forms of the 5-HT₂ receptors cannot be ruled out.

In addition to the high affinity for a series of serotonergic antagonists shared by the members of the 5-HT₂ family, the classical 5-HT₂ receptors (labeled by antagonistic radioligands) show a high affinity for chlorpromazine, cinanserin, ketanserin, ritanserin, and spiperone; moreover, they have a low affinity ($K_d = 100$ to 1000 nM) for 5-HT and its tryptamine derivatives (5-MeOT, 5-OH-DMT, 5-MeO-DMT) and no affinity for ligands selective for 5-HT₁ (8-OH-DPAT, RU 24969) or 5-HT₃ (2-methyl-5-HT, phenylbiguanide) (table 5).

2. Regional distribution in the brain. The distribution of 5-HT₂ receptors (see reviews by Palacios and Dietl, 1988; Pazos et al., 1988) has been studied in rat (Pazos et al., 1985a) and in human brain (Hoyer et al., 1986b; Biegon et al., 1986; Pazos et al., 1987b). It is heterogeneous in the brain, and some species differences have been reported. In the cortex (frontal, entorhinal, cingulate), these sites occur at high density, whereas in the rat brain, the layer IV is labeled more strongly than it is in the human brain, where layers III and V are the most heavily labeled. In the hippocampus, the subiculum is weakly labeled, whereas the CA1 and CA3 layers show a higher density of sites. In the basal ganglia, the caudate, putamen, and nucleus accumbens are labeled with an intermediate density, whereas the globus pallidus has a low density. Interestingly, the unspecific binding of [^3H]ketanserin in these areas is unusually high.

The nucleus basalis of Meynert is characterized by a low density of 5-HT₂ receptors. The thalamus shows a heterogeneous labeling (from intermediate to low density), whereas the hypothalamus, the corpus mamillare, and the cortex are the most enriched areas. The midbrain area ranges from low to very low density, and in the cerebellum these receptors are present at very low density.

3. Cellular localization. Lesion of the serotonergic system provoked by 5,7-DHT do not reduce the number of 5-HT₂ sites in rats (Fischette et al., 1987); therefore, the 5-HT₂ receptors are located on nonserotonergic neurons. The fact that cortical [^3H]ketanserin-binding sites decreased in Alzheimer dementia, paralleling the loss of somatostatin immunoreactivity and the decrease of GABA concentration in this region, suggests that 5-HT₂ receptors could be located on intrinsic GABA- or somatostatin-containing neurons in the cortex. In situ, hybridization studies have confirmed this assumption and demonstrated that cells intrinsic to the neocortex express mRNA coding for 5-HT₂ receptors (Mengod et al., 1990). Blue et al. (1988) observed a close spatial relationship between 5-HT₂ receptors and fine 5-HT axons. Because the cortex contains a high density of fine 5-HT axon terminals arising from the dorsal raphe nucleus, it appears likely that 5-HT₂ receptors may be selectively linked to this particular type of 5-HT axon terminal,

TABLE 5
Affinity values of drugs for ^3H -antagonists or ^3H -agonists of 5-HT₂ recognition sites in mammalian brain*

Drugs	^3H -antagonists K_i (nM)	^3H -agonists K_i (nM)
Agonists		
(-)-DOB	24 ^c	0.39 ^e , 0.62 ^d
(±)DOB	41 ^c	0.79 ^f
(±)DOI		1.3 ^c
(+)-DOB	146 ^c	2.3 ^c , 18 ^d
TFMPP	161 ^c , 251 ^a	16 ^c
mCPP	199 ^a	22 ^d
Quipazine	228 ^c	17 ^c , 31 ^d
5-OH-DMT	297 ^c	6 ^c
5-MeO-T	305 ^c	5 ^c
5-MeO-DMT	616 ^c	15 ^c
Spiroxastrine	630 ^a	
Buspirone	794 ^a	
RU 24969	777 ^c , 1,000 ^a	42 ^c , 91 ^d
5-HT	928 ^c , 3,162 ^a	7.8 ^c , 3.5 ^d
DMT	1,183 ^c	64 ^c
DP-5-CT	5,011 ^c	48 ^c
Tryptamine	2,005 ^c	320 ^d
8-OH-DPAT	5,350 ^c , 10,000 ^a	>10,000 ^b
5-CT	19,952 ^a	633 ^c , 2,700 ^d
WB 4101	1,258 ^a , >10,000 ^a	
GR 43175	195,526 ^a	
Antagonists		
Spiroxastrine	0.4 ^c , 0.5–1.2 ^a , 1.6 ^c , 2 ^b	0.8 ^c , 3.8 ^d
Ketanserin	0.39–2.1 ^a , 1.2 ^c , 3.1 ^b	1.3 ^c , 4.8 ^d
Ritanserin	1.48 ^a	
(+)Butaclamol	2.4 ^c	13 ^c , 29 ^d
Chlorpromazine	2.8 ^c , 3.3–20 ^a	7 ^c , 70 ^d
Cinanserin	2–41 ^a , 4.8 ^c , 19 ^c , 21 ^b	3.8 ^c , 7.2 ^d
Metergoline	0.28–0.8 ^a , 1 ^c	2.7 ^d
Mesulergine	3.98 ^a	4.5 ^d
Methiothepin	0.4–1.9 ^a , 1.58 ^c	6.5 ^d
Pitozifen	0.28–6.5 ^a , 4.4 ^b	
d-LSD	2.5–8.2 ^a , 2.5 ^c	4.8 ^d
DHE	2.5 ^c	
Cyproheptadine	3.16 ^c , 3.46 ^a	
Mianserin	1.4–13 ^a , 5 ^b , 7.9 ^c	
Methysergide	0.94–12 ^a , 2.5 ^c	
Clozapine	2–16 ^a , 34 ^b	
Pireperone	1.58 ^a	
Amitriptyline	4.2–21 ^a	
Lisuride	5.01 ^c	
Chlorimipramine	9.8–82 ^a	
Haloperidol	22–48 ^a	
Trazodone	24 ^b	
Imipramine	37–260 ^a	
LY 53857	50 ^a	
MDL 72832	158 ^a	
NAN-190	218 ^f	
BMY 7378	263 ^f	
Nomifensine	372 ^a	
(±)Propranolol	590–758 ^a	
Yohimbine	660 ^a	
(+)Propranolol	2,490 ^c	1,301 ^c
MDL 73005	3,162 ^f	
(-)Butaclamol		10,000 ^d
ICS 205-930		17,000 ^d
Prazosin	>10,000 ^a	>10,000 ^d
Cyanopindolol	31,622 ^a	
SDZ(-)21009	10,000 ^a	
(-)Pindolol	29,512 ^a –39,800 ^a	
(±)CYP	29,512 ^a	
(+)CYP	25,118 ^a	
(-)CYP	26,302 ^a	
(+)Pindolol	38,018 ^a	

* References: ^a Leysen et al., 1982a; ^b Conn and Sanders-Bush, 1986a; ^c Lyon and Titeler, 1988; ^d Peroutka et al., 1988; ^e Hoyer and Schoeffter, 1991; ^f Van Wijngaarden et al., 1990; ^g Engel et al., 1986.

mediating the effects of dorsal, but not median raphe, projections.

4. *Molecular structure.* In 1988, Pritchett et al., by screening a rat brain library, isolated a cDNA clone carrying cDNA homologous to the 5-HT_{1C} receptor. After transfection in embryonic human renal cells, this clone produced receptor sites having the characteristics of 5-HT₂ receptors, although 5-HT itself had a higher affinity for the cloned receptor than it did for receptors occurring naturally in membranes.

Julius et al. (1990) also cloned this gene from rat brain and corrected the published sequence of Pritchett et al. for one nucleotide. The sequence of the human 5-HT₂ receptor, cloned by Kao et al. (1989), corresponds closely to that corrected sequence and shows a high degree of identity with the rat sequence (90%). The entire receptor protein consists of 471 amino acids. The slight difference of three amino acids in the membrane-spanning domain and of two amino acids in the third intracellular loop might explain the difference in binding exhibited for mesulergine ($K_i = 4$ nM in rat and 150 nM in humans; Hoyer et al., 1986b). The receptor gene (HTR2) has been localized on human chromosome 13 (q14-q21) (Sparkes et al., 1991) and on mouse chromosome 14 (closely linked with the marker ES 10) (Liu et al., 1991).

The 5-HT₂ receptor is homologous to the 5-HT_{1C} receptor (78% identity in transmembrane domains: 141 amino acids of 180 amino acids are identical), whereas it is only 41% identical for the 5-HT_{1A} receptor (73 of 177 amino acids). This receptor is characterized by the presence of a leucine zipper in the first membrane-spanning domain in rat and human sequences (such a zipper is also present in the 5-HT_{1A} receptor but not in the 5-HT_{1C} receptor). This feature may favor dimerization of the receptor or its interaction with other membranal proteins.

Buck et al. (1991) studied the functional importance of NH₂ and COOH terminals using deletion mutants expressed in *Xenopus* oocytes. These authors observed an insignificant loss of electrophysiological activity when extracellular NH₂-terminal and intracellular COOH-terminal residues from 398 to 471 amino acids were deleted. However, when cysteine-397 was replaced by serine, the activity was totally lost. That deletion of amino acids 398 to 471 does not markedly reduce activity suggests that this part of the receptor does not play a major role in the regulatory processes.

Weinstein and Osman (1990), studying a theoretical model of interaction of 5-HT with this receptor, proposed that 5-HT was recognized by two aromatic residues separated by a hydrophilic residue and that these recognitions led to receptor activation through a structural rearrangement. This model, which is based on neutral interaction, differs from the one proposed by the same authors for the 5-HT_{1A} receptor, which involved a proton transfer.

5. Transduction systems. The 5-HT₂ receptors appear to be directly coupled to the phosphoinositide turnover; this coupling increases IP₃ production in brain cortical tissue in vitro (Conn and Sanders-Bush, 1984, 1986a) and in vivo (Hide et al., 1989). However, the stimulating effect of serotonergic agonists has not been reported to be mediated by 5-HT₂ receptors in other areas of the brain (Janowsky et al., 1984; Conn and Sanders-Bush, 1985). 5-HT₂ receptors are also directly involved in phosphatidyl inositide metabolism in human (De Chaffoy de Courcelles et al., 1985) and rabbit blood platelets (Schachter et al., 1985), in aortic myocytes (Roth et al., 1986; Cory et al., 1986), in A₇T₅ cultured smooth muscle cells (Coughlin et al., 1984; Doyle et al., 1986), in WRK₁ cells originating from a rat mammary tumor (Cory et al., 1987), and in a pituitary cell line P11 cells (Ivins and Molinoff, 1990, 1991). In C6 glioma cells, Ananth et al. (1987) presented experimental evidence for the coupling of 5-HT₂ receptors with the IP₃ production; however, the pharmacological properties of this effect did not entirely correlate with that of 5-HT₂ receptors (Sanders-Bush, 1988b). In American Tissue Culture Collection human embryonic kidney 293 cells transfected with the 5-HT₂ receptor gene, 5-HT also induces phospholipid breakdown and increases of intracellular Ca⁺⁺ (Pritchett et al., 1988).

It was also shown that serotonin, via a 5-HT₂ receptor, stimulates phospholipase A₂ and thus induces the release of arachidonic acid in hippocampal and cortical neurons (but not in glial cells); this mechanism is independent of inositol phospholipid hydrolysis (Felder et al., 1990).

5-HT₂ receptors play an electrophysiological role in facial motor neurons, initially examined by McCall and Aghajanian (1980), Van der Maelen and Aghajanian (1980), and, more recently, others (see review by Andrade and Chaput, 1991a), in nucleus accumbens (North and Uchimura, 1989) and brain cortical neurons (Davies et al., 1987; Araneda and Andrade, 1988). Pharmacological analysis revealed that the 5-HT-induced depolarization of the neuronal cells was 5-HT₂ dependent. Two distinct mechanisms were involved: a slow depolarization phase resulting from the closing of K⁺ channels, which reduced K⁺ conductance, and a posthyperpolarization phase controlled by a Ca⁺⁺-regulated potassium channel. These slow responses seem to involve the breakdown of membrane phosphatidylinositides; however, this mechanism has not yet been totally elucidated.

Pritchett et al. (1988) found that in *Xenopus laevis* oocytes transfected with the 5-HT₂ receptor gene, 5-HT was able to induce a rapid Cl⁻ influx through a Ca⁺⁺-dependent chloride channel. The physiological role of this mechanism remains to be demonstrated.

6. Regulation. The 5-HT₂ receptors were reported to be modulated by guanyl nucleotides, because these compounds significantly reduced the number of 5-HT₂ receptors (Kendall and Nahorski, 1983; Battaglia et al., 1984).

However, this effect was not confirmed (Peroutka et al., 1979; Engel et al., 1984; Shearman and Strange, 1988). The 5-HT₂ receptors labeled with antagonist radioligands were not affected by GTP and its analogs, whereas the affinity state (or the site) labeled by the 5-HT₂ agonists was guanyl nucleotide sensitive (Lyon et al., 1987; Teitler et al., 1990).

The regulation of the 5-HT₂ receptors during ontogenesis has been studied in rat brain (Roth et al., 1991). Roth et al. showed that mRNA-expressing 5-HT₂ receptors appeared in parallel to the binding sites, i.e., both increased markedly (eight times for the sites, 13 for the mRNAs) from embryonic day 17 to postnatal days 10 to 13 and then were reduced by 50% after days 25 to 27. These receptors appeared to be functional in young immature rats (day 8) as Claustre et al. (1988b) demonstrated, by examining the 5-HT₂-induced phosphatidyl inositide breakdown; the stimulating effect of 5-HT was six times greater in immature than in adult rat cortex.

In aged rats, 5-HT₂ receptor sites continue to diminish in number (-20%), as reported by Battaglia et al. (1987) and Gozlan et al. (1990). A similar age-dependent reduction in 5-HT₂ receptors has been reported to occur in the human brain (Wong et al., 1984; Marcusson et al., 1984; Reynolds et al., 1984; Gross-Isseroff et al., 1990).

Serotonergic denervation by 5,7-DHT generally does not modify the binding properties of the 5-HT₂ receptors (Blackshear et al., 1981; Quik and Azmitia 1983; Leysen et al., 1983; Barbaccia et al., 1983b; Hall et al., 1984; Stockmeier and Kellar, 1986; Conn and Sanders-Bush, 1986a; Fischette et al., 1987; Butler et al., 1990), nor does it modify IP₃ production (Conn and Sanders-Bush, 1986a; Godfrey et al., 1988b). These results indicate, first, that 5-HT₂ receptors are postsynaptically located and, second, that they are regulated by a mechanism different from that controlling other postsynaptic neurotransmitter receptors; the latter receptors are usually "upregulated" after denervation, resulting in the induction of a hypersensitivity. However, the following variations have been reported after serotonergic denervation: a decrease (Leysen et al., 1982a) or an increase in the number of 5-HT₂ receptors (Barbaccia et al., 1983b; Heal et al., 1985), an enhancement of IP₃ production (Butler et al., 1990), and a facilitation of 5-HT-induced head twitches, reflecting 5-HT₂ activation (Heal et al., 1985; Godfrey et al., 1988b).

Given that agonist exposure elicits a desensitization of 5-HT₂ receptors, the generally observed lack of denervation-hypersensitivity for these receptors suggested that the receptors are not under the tonic control of the serotonergic system (for further discussion, see Sanders-Bush, 1990) or that 5-HT itself is not the natural full agonist for this receptor (Roth et al., 1984, 1985). These authors also proposed the existence of a corresponding peptidic endogenous ligand.

A nonclassical regulation of the 5-HT₂ receptors is

observed after antagonist administration. Rather than eliciting the classical supersensitivity and upregulation, chronic antagonist administration elicits a decrease in 5-HT₂ receptor density (Blackshear and Sanders-Bush, 1982; Blackshear et al., 1983; Gandolfi et al., 1985; May et al., 1986; Leysen et al., 1986; Roth et al., 1989; Roth and Ciaranello, 1991; Pranzatelli, 1991) and a subsensitive phosphoinositide response (Conn and Sanders-Bush, 1986a). Interestingly, Roth and Ciaranello (1991) showed that the decrease of 5-HT₂ binding observed after mianserin treatment is not paralleled by a decrease in mRNA production, suggesting a posttranscriptional regulation mechanism.

Similarly, chronic treatment with a partial agonist (LSD) or with full agonists (DOI, DOB, and DOM) also decreases the B_{max} for [³H]ketanserin (Buckholtz et al., 1988; Leysen et al., 1989; McKenna et al., 1989; Pranzatelli, 1991), and [³H]DOB (Pranzatelli, 1991) causes desensitization of the head shake behavior in rats (Leysen et al., 1989).

Thus, the action of 5-HT₂ agonists and antagonists on the regulation of the 5-HT₂ receptors is not totally elucidated. A possible explanation is that 5-HT₂ receptors receive a very low stimulation under physiological conditions. Because of the low stimulation, the receptors could exist in a supersensitive state, and then, their chronic blockade would not produce further supersensitivity. The significant desensitization and downregulation of the receptor could be a defense mechanism against excessive stimulation (Sanders-Bush, 1990). The elucidation of the mechanism of 5-HT₂ receptor downregulation by antagonists, and the question of whether it differs from the agonists-induced downregulation, requires additional studies. Isolated cells in culture represent a promising model for these studies (Ivins and Molinoff, 1991).

The regulation of 5-HT₂ receptors observed after long-term antidepressant treatment stimulated much interest in how 5-HT₂ receptors participate in depression and in the mechanism of action of antidepressant drugs. Chronic treatment with antidepressants usually causes a downregulation of the 5-HT₂-binding sites (Peroutka and Snyder, 1980; Kellar et al., 1981; Blackshear and Sanders-Bush, 1982; Barbaccia et al., 1983b; Goodwin et al., 1984; Kendall and Nahorski, 1985; Metz and Heal, 1986; Mizuta and Segawa, 1989). This decrease is also observed after treatment with lithium (Treiser and Kellar, 1980; Wajda et al., 1986; Mizuta and Segawa, 1989; Newman et al., 1990) and some antipsychotics (Wilmot and Szezpanik, 1989; Matsubara and Meltzer, 1989; O'Dell et al., 1990). This downregulation of the sites is also accompanied by a decrease of the 5-HT₂ receptor activity as evidenced by a decrease in IP₃ production (Kendall and Nahorski, 1984, 1985; Conn and Sanders-Bush, 1986a; Godfrey et al., 1988b; Newman and Lerer, 1988b; Newman et al., 1990) and head twitch response (Goodwin

et al., 1984, 1986; Metz et al., 1985; Godfrey et al., 1988b). Roth and Ciaranello (1991) demonstrated that chronic treatment with mianserin fails to affect 5-HT₂ mRNA levels in rat brain, although it reduces (44 to 59%) the number of 5-HT₂ receptors.

Accordingly, the number of 5-HT₂ receptors is increased in depressed suicidal patients studied postmortem (Stanley and Mann, 1983; Mann et al., 1986; McKeith et al., 1987; Arora and Meltzer, 1989; Arango et al., 1990; Yates et al., 1990) and in blood platelets of depressed patients (Biegon et al., 1987, 1990). Yates et al. (1990) further showed that in still depressed patients treated with antidepressants, the 5-HT₂ binding is similar to the control; in depressed patients after euthymic recovery, however, the 5-HT₂ sites were markedly decreased. Other authors observed no modification in the brains of depressed patients examined postmortem (Crow et al., 1984; Cheetham et al., 1988). In an animal model of depression (learned helplessness), the number of 5-HT₂ receptors was either unchanged (Martin et al., 1990a) or increased (Barone et al., 1990).

On the other hand, electroconvulsive shock increases the number of 5-HT₂ receptors (Kellar et al., 1981; Vetulani et al., 1981; Green et al., 1983; Newman et al., 1987; Biegon and Israeli, 1987; Godfrey et al., 1988b); this effect may be sex dependent (Biegon and Israeli, 1987). The mechanisms responsible for 5-HT₂ receptor regulation by different antidepressive treatments remain to be defined.

5-HT₂ receptors may be involved in other psychiatric diseases because their number is reduced in persons suffering from schizophrenia (Mita et al., 1986), Parkinson's disease (Reisine et al., 1877; Maloteaux et al., 1988; Cheng et al., 1991), Alzheimer's disease (Cross et al., 1988; Cheng et al., 1991), and senile dementia of Lewy body type (Cheng et al., 1991).

The functional properties of 5-HT₂ receptors are modified by the GABA_B agonist baclofen in vivo (Metz et al., 1985) and in vitro (Godfrey et al., 1988a), by noradrenergic denervation (Eison et al., 1988), and by the administration of β -adrenergic agonists (Handley and Singh, 1986). These results suggest that GABAergic and adrenergic systems participate in the control of 5-HT₂ receptor activity.

Interestingly, 5-HT_{1B} receptors can modulate 5-HT₂-mediated behaviors, such as the DOI-induced ear scratch stereotypy in mice. Simultaneous costimulation of 5-HT_{1B} receptors by both TFMPP and RU 24969 potently inhibits this behavior (Darmani et al., 1990a). 5-HT_{1A} receptor activation also seems to inhibit 5-HT₂ receptor function. Indeed, the 5-HT₂ receptor-mediated head shakes, evoked in rats by DOI, could be attenuated by 8-OH-DPAT (Berendsen and Broekkamp, 1990). In addition, the 5-HT₂ receptor-mediated quipazine-induced head shake response in rats could be blocked by the 5-HT_{1A} agonists 8-OH-DPAT, buspirone, gepirone, and

ipsapirone (Yocca et al., 1990a). Similarly, the 5-HT₂ receptor-mediated head twitches, evoked in mice and rats by DOI, was inhibited by 8-OH-DPAT (Arnt and Hyttel, 1989; Darmani et al., 1990b). Moreover, chronic buspirone administration also produces downregulation of 5-HT₂ receptors in rats (Taylor and Hyslop, 1991).

These results, together with the behavioral and biochemical data demonstrating a negative control of 5-HT₂ receptors on the 5-HT_{1A} receptor function (section III.A.7), strongly demonstrate an interaction between the two receptor systems. Although the mechanism of this interaction is unknown, it is possible that it involves a second-messenger production, because the dibutyryl-cAMP can enhance the density of the 5-HT₂-binding sites that regulate the expression of the 5-HT₂ receptor gene (Shigeri et al., 1992) and because the PKC activation can desensitize 5-HT_{1A} receptor function (Raymond, 1991; Liu and Albert, 1991). The 5-HT_{1A}-5-HT₂ interaction may be involved in the reciprocal modulations between 5-HT₁ and 5-HT₂ receptors proposed in the "5-HT receptor imbalance" hypothesis (Deakin, 1988, 1989). This hypothesis explains how a 5-HT_{1A} deficit may be involved in a depressive syndrome and how 5-HT₂/5-HT_{1C} excessive neurotransmission may be involved in anxiety.

The existence of an endogenous ligand interacting directly with the 5-HT₂ receptors has been proposed by Costa's group. A cerebral peptidic fraction (6000 daltons) was isolated that displaced [³H]ketanserin in rat brain cortex (Roth et al., 1984, 1985; Barbaccia and Costa, 1986; Apud and Ito, 1991). Its function is not known.

7. Cellular functions. 5-HT₂ receptors are involved in the control of neurotransmitters other than 5-HT. K⁺-evoked ACh release is selectively inhibited by 5-HT₂ agonists in the hippocampus (Muramatsu et al., 1988a) and rat brain cortex (Muramatsu et al., 1990), whereas the activation of 5-HT₂ receptors increases the basal release of ACh in the striatum (Bianchi et al., 1989). These effects are reduced by antidepressant drug treatment (Siniscalchi et al., 1990b; Muramatsu et al., 1990). 5-HT₂ receptors, and possibly 5-HT_{1C} receptors, have potent and direct excitatory actions on the GABAergic neurons of the thalamic reticular nucleus; this postsynaptic response appears to result from the suppression of a potassium current (McCormick and Wang, 1991). Experiments in which serotonin agonists and antagonists were applied intraventricularly suggest that the serotonergic negative control of noradrenergic release in the locus coeruleus is mediated through 5-HT₂ receptors (Gorea and Adrien, 1988). 5-HT₂ receptors also inhibit the K⁺-evoked release of glutamate and aspartate in the cerebellum (Maura et al., 1988, 1991) and the release of dopamine in the striatum (Muramatsu et al., 1988b). In the spinal cord, their activation increases the K⁺-evoked release of substance P (Iverfeldt et al., 1986).

DOI increases the secretion of renin by activating both

peripheral and brain 5-HT₂ receptors (Rittenhouse et al., 1991). 5-HT₂ receptors also regulate the secretion of vasopressin (Rittenhouse et al., 1990) and play a secondary role in regulating the activity of the hypothalamo-hypophysis axis (Koenig et al., 1987).

8. 5-HT₂ receptors and behavior. 5-HT₂ receptors are involved in motor behavior in rats and mice. The head twitch is induced in rodents by serotonergic agonists and antagonized by the following selective antagonists: metergoline, cinanserin, cyproheptadine, methysergide, 2-bromo-LSD, piramperone, and ketanserin (Malick et al., 1977; Friedman and Dallob, 1979; Matthews and Smith, 1980; Peroutka et al., 1981; Colpaert and Janssen, 1983a,b; Lucki et al., 1984; Heal et al., 1985; Ögren and Fuxe, 1989). The "wet dog shake" is also inhibited by 5-HT₂ antagonists (Bedard and Pycocock, 1977; Matthews and Smith, 1980; Vetulani et al., 1980; Yap and Taylor, 1983; Fone et al., 1989). 5-HT₂ receptors may mediate the contraction of back muscles, but 5-HT_{1C} activation may also be required for this activity (Fone et al., 1989).

Gudelsky et al. (1986, 1987) reported that 5-HT₂ receptors are involved in temperature regulation. These authors showed that hyperthermia induced by MK 212 is antagonized by ketanserin; however, the involvement of 5-HT_{1C} and possibly 5-HT_{1A} receptors in this mechanism was not excluded. Similarly, the hypothermia induced by 5-MeO-DMT might be controlled via 5-HT₂ receptors and also 5-HT_{1C} and 5-HT_{1A} subtypes (Gudelsky et al., 1986).

It is not likely that 5-HT₂ receptors are directly involved in regulating food intake. However, Hewson et al. (1988), who studied the effects of quipazine, and Schechter and Simansky (1988), who studied the effects of DOI, have proposed that these receptors play a role in reducing food intake. Wilkinson and Dourish (1991) concluded that the anorectic effects of 5-HT₂ agonists, particularly DOI, might be mediated by 5-HT_{1C} receptors. Peripheral 5-HT₂ activity may also be responsible for these effects (Massi and Marini, 1987).

It is likely that 5-HT₂ receptors play a role in sexual behavior in female rats because selective 5-HT₂ antagonists inhibit lordosis behavior (Mendelson and Gorzalka, 1986b; Hunter et al., 1985) and 5-HT₂ agonists facilitate it (James et al., 1989). The dual effect of LSD suggests the involvement of several types of receptors (Sietnieks and Mayerson, 1983). Although 5-HT₂ antagonists reduce male sexual activity (Mendelson and Gorzalka, 1986b) and the 5-HT₂ agonist DOI stimulate global sexual activity in male rats (Foreman et al., 1989), the actual role of 5-HT₂ receptors is still unclear, because LSD and DOM do not alter male sexual behavior (Ahlenius and Larsson, 1987) and DOI has also been reported to inhibit the copulatory behavior in male rats; the latter effect is reversed by 5-HT₂ antagonists (Watson and Gorzalka, 1991).

In various experimental behavioral tests (Berge, 1982;

Post et al. 1986), analgesic effects of 5-HT₂ agonists were blocked by specific 5-HT₂ antagonists. These effects seem to involve other neurotransmission systems because the 5-HT₂ receptors exert negative effects on the firing of the dopaminergic and noradrenergic neurons located supraspinally which control nociception (Alhaider, 1991). Furthermore, it was recently reported that the nociceptive actions of intravenously administered 5-HT require dual activation of 5-HT₂ and 5-HT₃ receptors subtypes (Meller et al., 1991).

5-HT₂ receptors play an important role in regulating sleep. In rats, specific 5-HT₂ agonists (RP 62203, DOM, and DOI), (Dugovic and Wauquier, 1987; Dugovic et al., 1989; Stutzmann et al., 1990) decrease the light and REM sleep and increase the duration of slow-wave sleep. Certain 5-HT₂ antagonists (ritanserin) block the increase in slow-wave sleep but do not affect REM sleep, suggesting that 5-HT₂ receptors may not be involved to the same extent in the two cases. Other 5-HT₂ antagonists (2-(2-dimethylamino-2-methylpropylthio)3-phenyl quinoline and ICI 169-369), however, alter REM sleep (Tortella et al., 1989). In humans, 5-HT₂ antagonists increase slow-wave sleep (Idzikowski et al., 1986; Sharpley et al., 1990b); interestingly, this effect did not occur in depressed patients (Sharpley et al., 1990a). 5-HT₂ receptors are regulated during circadian rhythm (Wesemann and Weiner, 1990), and they might also regulate the sleep-waking cycle. For example, ritanserin is able to counteract jet lag (Hoppenbrouwers et al., 1988).

Although anxiolytic properties of 5-HT₂ have been described in various experimental models, these properties are not yet fully established. 5-HT₂ antagonists (cyproheptadine, methysergide, metergoline, pizotifen, ritanserin) exhibit anxiolytic properties; these effects are limited and not consistently dose related, as shown in the conflict test (Meert and Colpaert, 1986) and the elevated X-maze test (Critchley and Handley, 1987). However, in the Vogel conflict test, as well as in the open-field test, ritanserin appears to act in a dose-dependent manner (Colpaert et al., 1985). RP 62203, a new 5-HT₂ antagonist, is anxiolytic in the elevated X-maze test, whereas ritanserin is not (Stutzmann et al., 1991).

5-HT₂ antagonists are not yet recognized as being anxiolytic in humans, although from preliminary clinical observations it would seem that ritanserin (Ceulemans et al., 1985; Reyntjens et al., 1986; Pangalia-Ratu Langi and Janssen, 1988; Meert and Janssen, 1989) and trazodone (Al-Yassiri et al., 1981) play an anxiolytic role. These effects might be due to the presence of 5-HT₂ receptors in the amygdala, an area that has been proposed to play an important role in this disorder (Deakin, 1989).

The number of 5-HT₂ receptors has been shown to decrease in schizophrenic patients (Mita et al., 1986). 5-HT₂ antagonists are being developed as neuroleptics on the basis of experimental results (Ortmann et al., 1982;

Gelders et al., 1986; Wander et al., 1987; Lowe et al., 1988; Van der Heyden, 1989). However, the neuroleptic activity of the 5-HT₂ antagonists has not yet been clearly demonstrated.

5-HT₂ receptors may play an important role in the mechanism of action of hallucinogenic drugs (Glennon, 1985; Glennon et al., 1985; Sadzot et al., 1989). However, this mechanism is far from well understood. Initially, the 5-HT₂ antagonistic properties of *d*-LSD were thought to be responsible for its hallucinogenic properties (Pierce and Peroutka, 1988), but some authors think that, on the contrary, these properties are due to its agonistic activities (Glennon, 1990a). It has also been proposed that 5-HT_{1C} receptors are involved in these effects (Glennon, 1990a).

Serotonergic agonists favor electroconvulsions and pentylenetetrazol-induced seizures in mice. That this effect is antagonized by ketanserin suggests the participation of 5-HT₂ receptors in epilepsy (Löscher and Czuczwar, 1985).

The role of 5-HT₂ receptors in the periphery is important, particularly in cardiovascular physiology, but is beyond the scope of this review (Hollenberg, 1988; Saxena et al., 1989; Wiernsperger, 1990).

It is surprising that 5-HT₂ receptors, which are strongly regulated in many pathological circumstances, apparently do not play an important physiological role. It was proposed that they receive little stimulation under normal physiological conditions and function only in "emergency" or pathological conditions (Leysen, 1990).

B. 5-HT_{1C} Receptors

The 5-HT_{1C} receptors, considered initially as 5-HT₁-like receptors on the basis of their relatively high affinity for [³H]5-HT, are present at high density in the choroid plexus (Pazos et al., 1985b). Their pharmacological characteristics are similar to those of the 5-HT₂ receptors. It was, therefore, suggested that they might be related to these receptors (Hoyer, 1988). This has now been definitely established; comparison of their nucleotide sequences showed 5-HT_{1C} and 5-HT₂ receptors to be homologous (see review by Hartig et al., 1990a,b).

1. *Selective radioligands and pharmacological properties.* The 5-HT_{1C} sites were initially identified by using [³H]5-HT, [³H]mesulergine, and [³H]LSD (Pazos et al., 1985b) and later by using ¹²⁵I-LSD (Yagaloff and Hartig, 1985), ¹²⁵I-1-methyl-LSD (Yagaloff and Hartig, 1986), [³H]mianserin (Sanders-Bush and Breeding, 1988), [³H]SCH 23390 (Nicklaus et al., 1988), and [³H]SCH 23982 (Hoyer and Karpf, 1988).

At present, no ligand has been found that specifically recognizes the 5-HT_{1C} receptor. This receptor has a medium affinity for 5-HT ($K_d = 10$ to 30 nM), whereas other 5-HT₁ subtypes exhibit a high affinity for 5-HT ($K_d = 1$ to 5 nM); 5-HT₂ and 5-HT₃ receptors exhibit a low affinity ($K_d = 900$ to 3000 and 600 to 900 nM,

respectively). The 5-HT_{1C} site also shows a low affinity for 5-CT ($K_d = 630$ nM). The serotonergic drugs that are antagonistic at the 5-HT₂ receptors generally are also recognized by 5-HT_{1C} sites with a high affinity (<10 nM). However, the 5-HT_{1C} site recognizes ketanserin and cinanserin with 10 to 30 times lower affinity than do 5-HT₂ sites; spiperone has a 1000-fold lower affinity. The latter drug, therefore, allows 5-HT₂ and 5-HT_{1C} receptor subtypes to be distinguished. The phenylalkylamine derivatives (DOM, DOB, DOI), considered to bind selectively at 5-HT₂ receptors, also recognize 5-HT_{1C} receptors with the same affinity (Glennon et al., 1992).

The 5-HT₂ receptor antagonists are potent and competitive displacers at the 5-HT_{1C} sites in pig choroid plexus (Sahin-Erdemli et al., 1991). Thus, the 5-HT_{1C} receptors apparently do not undergo allosteric modulation as described previously for 5-HT₂ receptors (Kauermann and Frenken, 1985). Interestingly, the 5-HT_{1A} agonist 8-MeO-2'-chloro-8-methoxy-2[N-propyl-N-propylamino]tetraline irreversibly blocks not only the 5-HT_{1A} receptors but also the 5-HT_{1C} receptors in the rat choroid plexus (Radja et al., 1989).

2. *Regional distribution within the brain.* Several laboratories have demonstrated the high concentration of 5-HT_{1C} receptors in the choroid plexus of different species: rat (Yagaloff and Hartig, 1985; Peroutka, 1986), pig (Pazos et al., 1985b; Yagaloff and Hartig, 1985), mouse (Mengod et al., 1990), and humans (Hoyer et al., 1986b). Transgenic mice spontaneously developing a choroid plexus tumor exhibit in this area the highest density of 5-HT_{1C} receptor found in any tissue (>6.000 fmol/mg protein) (Yagaloff et al., 1986).

Autoradiographic studies demonstrated that 5-HT_{1C} receptors are present at low levels in many other areas of the brain, including the substantia nigra, globus pallidus, cerebral cortex, and olfactory tubercles (Pazos et al., 1985b, 1988). Studies of the distribution of the mRNA encoding for 5-HT_{1C} receptors confirmed and extended these results (Molineaux et al., 1989; Hoffman and Mezey, 1989; Mengod et al., 1990) and suggested that the function of the 5-HT_{1C} receptors is not limited to that performed in the choroid plexus.

3. *Cellular localization.* The 5-HT_{1C} receptors have been studied more extensively in choroid plexus than in any other tissue. They appear to be located on epithelial cells where they regulate the production and composition of cerebrospinal fluid (Pazos and Palacios, 1985). Dopaminergic lesions do not affect 5-HT_{1C} binding in this area (Palacios and Dietl, 1988), suggesting that 5-HT_{1C} receptors are not located on dopaminergic neurons. Provoking specific lesions in the serotonergic neurons by using 5,7-DHT-hypersensitized 5-HT_{1C} receptors, as measured by the increase of IP₃ production (Conn et al., 1987), strongly suggested that these receptors are located on nonserotonergic neurons.

4. *Molecular structure.* Lübbert et al. (1987a,b) cloned

the 5-HT_{1C} receptor by using a cloning strategy based on electrophysiological assays of *X. laevis* oocytes. These cells were injected with mRNA from a mouse choroid plexus tumor previously fractionated by gel electrophoresis. The 5-HT_{1C} mRNA expressed in the oocytes was detected by voltage-clamp measurements of currents induced by 5-HT. Then, a cDNA library was constructed using this mRNA, and a clone carrying a 5-HT_{1C} gene fragment of 1.9 kb was identified. Julius et al. (1988) used an mRNA fraction from the choroid plexus to construct a cDNA library transcribed in vitro with T7 RNA polymerase. The cDNA transcripts injected into oocytes or into NIH 3T3 fibroblasts were screened to select those producing a functional 5-HT_{1C} receptor. One cDNA clone containing the entire 5-HT_{1C}-coding region was isolated and characterized.

The 5-HT_{1C} receptor, cloned from rat brain, is a monomeric protein belonging to the large multigenic family characterized by seven membrane-spanning domains and linked to a G protein. It consists of 460 amino acids with a molecular weight of 51,899 daltons; the amino terminal (19 amino acids) possesses a glycosylation site. An aspartate residue located in the second transmembrane domain may play an important role in agonist recognition, as occurs in the β -adrenoceptor (Strader et al., 1989). The third intracellular loop (77 amino acids) and the carboxy terminal (29 amino acids) contain potential phosphorylation sites, i.e., four of the last 12 amino acids of the COOH terminal are serine. This receptor does not possess a leucine zipper. Its sequence is homologous to that of 5-HT₂ (78% identity in the transmembranal domains) but not to that of the 5-HT_{1A} receptor (40% identity in transmembranal domain).

Recently, Yu et al. (1991) cloned the mouse 5-HT_{1C} gene and proposed, on the basis of hydrophobicity analysis, that the corresponding encoded protein contains eight hydrophobic domains capable of forming membrane-spanning α -helices; this observation contrasts with the usual "seven-helix" paradigm described for other GTP-related receptors. The existence of this hydrophobic region, designated domain E, near the amino terminal of the receptor (Yu et al., 1991), raises the question of its functional significance. The 5-HT_{1C} receptor gene, designated *Htr1c*, has been assigned to the X chromosome, and thus, its chromosomal location differs from that of 5-HT_{1A} and 5-HT₂ genes mapped to autosomes.

5. *Transduction system.* 5-HT_{1C} receptors are coupled to the breakdown of phosphatidyl inositides as shown in rat (Conn and Sanders-Bush, 1986b) and pig (Hoyer et al., 1989); these receptors are not linked to adenylyl cyclase activity (Palacios et al., 1986). IP₃ production controls a Ca²⁺-dependent chloride channel (Lübbert et al., 1987a,b) via a calcium-calmodulin dependant protein kinase (Tohda et al., 1991a,b) and a Ca²⁺-independent potassium channel (Panicker et al., 1991) in *Xenopus* oocytes transfected with 5-HT_{1C} mRNA. However, the

control of these channels has not yet been confirmed in brain tissue.

Recently, Hartig et al. (1990b) reported that 5-HT_{1C} activity may be directly linked to the stimulation of cyclic GMP production.

6. *Regulation.* The postnatal development of 5-HT_{1C} receptors has been studied in rat brain choroid plexus. The number of binding sites increases markedly between days 3 and 20 after birth (Zilles et al., 1986), and the 5-HT_{1C} receptor develops earlier than do other 5-HT₁ subtypes. In parallel to the 2-fold increase in 5-HT_{1C} sites in rat brain, production of the corresponding mRNA increases 5-fold between days 17 and 27 (Roth et al., 1991). Roth et al. (1990) reported that in the raphe a "burst" expression for the receptor gene occurs at postnatal day 7; these authors suggested that a transcriptional process was involved.

The mechanisms responsible for regulating 5-HT_{1C} receptors by exogenous or endogenous compounds are not currently clear (see review by Roth et al., 1990). The modifications of mRNA production following treatment with various drugs (antidepressants, lithium) generally have not been studied in parallel with alterations of the binding sites. To our knowledge, only one study has been performed in which chronic treatment with mianserin was used. In this study, the production of mRNA increased, whereas the number of 5-HT_{1C}-binding sites decreased (Roth et al., 1989). These results suggest that mianserin regulates 5-HT_{1C} receptors through a post-transcriptional mechanism.

The alteration of 5-HT_{1C} binding by antidepressants in the choroid plexus was studied Mizuta and Segawa (1989). 5-HT_{1C} binding partially disappeared after chronic treatment with imipramine and lithium. In vitro studies with *Xenopus* oocytes showed that imipramine and desipramine, at nanomolar concentrations, block the chloride current induced by 5-HT_{1C} activation (Tohda et al., 1989; Tohda and Nomura, 1991). Thus, the mechanism of action of some antidepressants may involve 5-HT_{1C} receptors.

Recently, Walter et al. (1991) cloned the gene of a protein that controls the sensitivity of both the 5-HT_{1C} and the M₁ muscarinic receptors; the regulatory mechanism has not yet been explored, but it may represent an important phenomenon in serotonergic physiopathology.

7. *Cellular functions.* The main function attributed to the 5-HT_{1C} receptor is the control of the exchanges between the CNS and cerebrospinal fluid (Hartig, 1989b). In particular, Tsutsumi et al. (1989) and Tsutsumi and Sanders-Bush (1990) demonstrated that the production of transferrin by choroid plexus epithelial cells is regulated through 5-HT_{1C} receptors.

A potentially important role of the 5-HT_{1C} and 5-HT₂ receptor gene is that of protooncogenes (Julius et al., 1989, also see review by Julius, 1991). When the receptor gene is expressed in transfected fibroblasts NIH 3T3,

and these cells are then exposed at length to 5-HT, foci form that can induce malignant tumors in athymic or nude mice. Nevertheless, 5-HT_{1C} activation in neuronal cells does not lead to cell proliferation under normal conditions. Therefore, whether the 5-HT_{1C} gene acts as a protooncogene apparently depends closely on its cellular environment.

8. *5-HT_{1C} receptors and behavior.* A clear role of 5-HT_{1C} in behavior has not yet been established because of the lack of specific ligands known for this receptor. However, motor effects have been produced especially by piperazine agonists (TFMPP and mCPP) and unselective ligands, such as MK 212, which induce hyperlocomotion (Sills et al., 1984; Kennett and Curzon, 1988; Lucki et al., 1989; Klodzinska et al., 1989). The latter effect is blocked by metergoline, methysergide, and mianserin but not by ketanserin or α_1 -, α_2 -, or β -adrenergic antagonists, suggesting the specific involvement of 5-HT_{1C} receptors.

Dourish et al. (1989) proposed that postsynaptic 5-HT_{1C} receptors located in the ventromedial hypothalamus play an important role in regulating feeding, because mCPP and TFMPP also decrease food intake in rats (Kennett and Curzon, 1988, 1991; Curzon and Kennett, 1990).

mCPP and TFMPP, which induce dysphoria (Murphy et al., 1989) and panic in humans (Charney et al., 1987), are also anxiogenic in rats (Kennett et al., 1989; Whitton and Curzon, 1990). Their effect might be related to 5-HT_{1C} receptors, because these behaviors are blocked by mianserin, cyproheptadine, metergoline but not ketanserin and cyanopindolol. However, the observation that ritanserin, which has a high affinity for 5-HT_{1C} ($K_d = 1.9$ nM), is not active does not favor this hypothesis.

Finally, 5-HT_{1C} receptors may play a role in initiating migraine attacks (Fozard and Gray, 1989). Recent evidence favoring this proposal is that ergotamine and dihydroergotamine, effective antimigraine agents, are potent 5-HT_{1C} receptor antagonists in piglet choroid plexus (Brown et al., 1991).

V. 5-HT₃ Receptors

The first indications of the existence of another serotonergic receptor, called "M receptor" by Gaddum and Picarelli (1957), were obtained from pharmacological studies carried out on isolated guinea pig ileum. These receptors control muscle contraction via ACh release, and their effect is antagonized by morphine. Later, Fozard et al. (1979) found that cocaine inhibits the sympathomimetic-like effects induced by these receptors in isolated rabbit heart. This observation led to the development and use of several cocaine derivatives (MDL 72222, ICS 205-930, and BRL 24924) (see review by Richardson and Buccheit, 1988).

Richardson and Engel (1986) proposed that the 5-HT₃ peripheral receptors are heterogeneous and include the

5-HT_{3A} subtype present in the vagus nerve, superior cervical ganglion, and sensory nerves, the 5-HT_{3B} subtype present in sympathetic and parasympathetic nerves, and the 5-HT_{3C} subtype present in enteric nerves. However, because the experimental models used to define these hypothetical subtypes make use of different tissues and species, doubt remains concerning their existence (Hoyer, 1990). Rapid developments in molecular biology will soon make it possible to characterize potential subtypes.

Various reviews deal with 5-HT₃ receptors, including peripheral types (Fozard, 1984; Richardson and Engel, 1986; Richardson and Buccheit, 1988). The present review will be limited to those receptors found in the CNS.

A. Selective Radioligands and Pharmacological Properties

Specific 5-HT₃-binding sites in the CNS were initially demonstrated by Kilpatrick et al. (1987) using [³H]GR 65630. Other specific radioligands have now been discovered and used to characterize these receptors, including [³H]zacopride (Barnes et al., 1988a), [³H]quipazine (Peroutka and Hamick, 1988, Milburn and Peroutka, 1989), [³H]ICS 205-930 (Hoyer and Neijt, 1988), [³H]BRL 43694 (granisetron) (Nelson and Thomas, 1989), [³H]QICS 205-930 (Watling et al., 1988), [³H]GR 67330 (Kilpatrick et al., 1990b), [³H]LY 278-584 (Wong et al., 1989; Gehlert et al., 1991), and ¹²⁵I-zacopride (Hamon et al., 1990a).

To further characterize the 5-HT₃-binding sites, Michel et al. (1991) pretreated membranes with various reagents known to chemically modify specific amino acid residues. Among all the reagents tested, N-bromosuccinimide, a specific tryptophan reagent, modified the binding characteristics of [³H]zacopride (60% reduction of the B_{max}), suggesting that intact tryptophan residue(s) are required for the specific binding of [³H]zacopride. Further studies are needed for a direct demonstration of this involvement.

Many serotonergic agonistic drugs are not highly selective and generally have only a moderate affinity for the receptor: 5-HT (100 to 800 nM), α -methyl-5-HT (200 nM), bufotenine (210 nM), phenylbiguanide (300 to 1000 nM), 2-methyl-5-HT (approximately 1000 nM) (table 6).

Recently, Kilpatrick et al. (1990b) discovered a high-affinity agonist, 1-(*m*-chlorophenyl)-biguanide ($K_i = 0.3$ nM). Interestingly, the same authors observed that [³H]GR 67330 could label a heterogeneous population of binding sites in the entorhinal cortex, which might correspond either to different 5-HT₃ subtypes or to different conformations of the same receptor, in a manner similar to those described for the nicotinic receptor (Changeux, 1981). In fact, agonistic drugs displace 5-HT₃ radioligands with a Hill coefficient >1, suggesting the occurrence of complex interactions, possibly related to confor-

mational changes induced by these agonists. These phenomena are not observed with antagonists.

Many antagonists are specific and have a very high affinity, e.g., quipazine, SDZ 206-830, LY 278-584, zacopride, GR 38032 (ondansetron), GR 67330, 1H-indole-3-carboxylic acid-*trans*-octahydro-3-oxo-2,6-methano-2H-quinolizin-8-yl-ester methane sulfonate, ICS 205-930 (tropisetron), BRL 43694 (granisetron), MDL 72222, BRL 24924, and BRL 47204. The 5-HT₁ and 5-HT₂ antagonists do not interact with a high affinity with 5-HT₃ receptors (table 7).

Computerized modeling has been used to study the pharmacophore structure and, in particular, to predict the affinities with which new drugs bind to 5-HT₃ receptors (Schmidt and Peroutka, 1989, 1990). The advantage of this approach is that agents active at the receptor can be analyzed and identified with minimal dependence on the use of animals and radioactive compounds.

B. Regional Distribution within the Brain

The distribution of the 5-HT₃ receptors in the peripheral tissues is mainly, if not totally, neuronal (enteric nerve system, parasympathetic, sympathetic, sensorial system) (Fozard, 1984; Richardson and Buccheit, 1988).

The regional distribution of these receptors in the CNS has been studied in various species: rat (Kilpatrick et al., 1987; Barnes et al., 1990a), mouse (Waeber et al., 1988b; Kilpatrick et al., 1989), ferret (Kilpatrick et al., 1989; Barnes et al., 1988b), rabbit (Kilpatrick et al., 1989), cynomolgus monkey (Kilpatrick et al., 1990a), and human brain (Waeber et al., 1989b; Barnes et al., 1989a). These studies demonstrated that the highest density of 5-HT₃-binding sites occurs in the area postrema; other areas of the brain also contain 5-HT₃ sites: entorhinal cortex > retrosplenic cortex > frontal cortex > amygdala > hippocampus > accumbens > septum > thalamus > hypothalamus; the cerebellum is almost totally devoid of specific sites. When compared with other serotonergic receptors, the 5-HT₃ receptors exhibit a particularly low density in the brain (approximately 10 times less in the richest areas).

The presence of the 5-HT₃ receptors in the area postrema is of particular interest because this area is involved in the emesis mechanism, and anti-5-HT₃ compounds are antiemetics when administered systemically or injected locally into this area. The 5-HT₃ receptors involved in this phenomenon have not yet been precisely localized, but it has been shown that 5-HT₃ receptors are present on dorso-vagal complex afferent fibers in the tractus solitarius, a region close to the area postrema (Reynolds et al., 1989a,b; Pratt and Bowery, 1989; Pratt et al., 1990).

Neuronal cell lines may provide a good source of 5-HT₃ because the murine neuroblastoma cell line NIE 115 (Hoyer and Neijt, 1988) and the neuroblastoma-glioma hybrid NG 108-15 (Hoyer and Neijt, 1987; Neijt

TABLE 6
Affinity values of drugs for 5-HT_{1C} recognition sites in mammalian brain membranes

Drug	Affinity	Reference	Drug	Affinity	Reference
Very high affinity ($K_i < 10$ nM)					
Metergoline	0.51	Hoyer et al., 1985b	Ketanserin	28	Yagaloff and Hartig, 1985
	0.64	Engel et al., 1986; Hoyer and Schoeffter, 1991		30	Sanders-Bush, 1988b
Ritanserin	0.2	Sanders-Bush, 1988b		50.1	Pazos et al., 1985b
	0.54	Van Wijngaarden et al., 1990		95	Van Wijngaarden et al., 1990
	1.25	Hoyer and Schoeffter, 1991	97.7	Hoyer and Schoeffter, 1991	
Mesulergine	1.0	Sanders-Bush, 1988b	Pirempereone	12.5–25	Pazos et al., 1985b
	1.6	Hoyer et al., 1985b; Hoyer and Schoeffter, 1991		50	Hoyer and Schoeffter, 1991
Methysergide	1.2–3.9	Pazos et al., 1985b	Ergometrine	61,000	Peroutka, 1986
	1.9–3.6	Pazos et al., 1985b		31.6	Hoyer and Schoeffter, 1991
	2.5	Hoyer and Schoeffter, 1991	Tryptamine	46.7	Engel et al., 1986
Mianserin	62	Peroutka, 1986	Ergotamine	50	Hoyer and Schoeffter, 1991
	1.9	Yagaloff and Hartig, 1985	TFMPP	12	Van Wijngaarden et al., 1990
	2.0	Sanders-Bush, 1988b		50	Schoeffter and Hoyer, 1989a
	6.3–12	Pazos et al., 1985b	63	Hoyer and Schoeffter, 1991	
Methiothepin	10	Hoyer and Schoeffter, 1991	5-MeO-DMT	1,000	Peroutka, 1986
	3.16–15	Pazos et al., 1985b		57	Peroutka, 1986
	27	Schlicker et al., 1989	87	Hoyer, 1988	
	25	Hoyer and Schoeffter, 1991	5-NH ₂ -T	70	Engel et al., 1986
<i>d</i> -LSD	4.4	Yagaloff and Hartig, 1985	Imipramine	100	Sanders-Bush, 1988b
	5.24	Schoeffter and Hoyer, 1989a	α -Dihydroergocriptine	100	Hoyer and Schoeffter, 1991
1-NP	5.24	Schoeffter and Hoyer, 1989a	Low affinity ($K_i = 100$–$1,000$ nM)		
SCH 23390	5.49	Hoyer, 1988	Cinanserin	74	Yagaloff and Hartig, 1985
LSD	5–7.9	Pazos et al., 1985b		199	Hoyer and Schoeffter, 1991
	11.7	Engel et al., 1986	(+)Butaclamol RU 24969	199–1,584	Pazos et al., 1985b
12.5	Hoyer and Schoeffter, 1991	158		Peroutka, 1986	
DOI	6.45	Van Wijngaarden et al., 1990	47.8	Pazos et al., 1985b	
	18.6	Hoyer, 1988		Van Wijngaarden et al., 1990	
Pitozifen	7.94	Hoyer and Schoeffter, 1991	150	Peroutka, 1986	
	7.94	Hoyer and Schoeffter, 1991	158–199	Pazos et al., 1985b	
LY 53857	7.94	Hoyer and Schoeffter, 1991	316	Hoyer and Schoeffter, 1991	
ICI 169-369	9.33	Sahin-Erdemli et al., 1991	Bufotenine	70	Hoyer, 1988
				110	Peroutka, 1986
5-HT	11	Peroutka, 1986	(-)Propranolol	158	Hoyer et al., 1985b
	30.4	Yagaloff and Hartig, 1985		407	Hoyer and Schoeffter, 1991
	31.6	Hoyer and Schoeffter, 1991	Quipazine	186	Schoeffter and Hoyer, 1989
		9,900		Peroutka, 1986	
Cyproheptadine	8.3	Hoyer, 1988	Bromocriptine	199	Hoyer and Schoeffter, 1991
	12.58	Hoyer and Schoeffter, 1991		398	Hoyer and Schoeffter, 1991
mCPP	13.3	Engel et al., 1986	WB 4101	1,100	Peroutka, 1986
	19.9	Hoyer and Schoeffter, 1991	5-BT	370	Peroutka, 1991b
Lisuride	19.9	Hoyer and Schoeffter, 1991	5,6-DHT	410	Peroutka, 1986
			2-Me-5HT	707	Engel et al., 1986
5-MeO-T	23.4	Schlicker et al., 1989		489	Van Wijngaarden et al., 1990
	25	Peroutka, 1986	5-CO-NH ₂ -T MDL 72832	1,445	Engel et al., 1986
24.5	Hoyer and Schoeffter, 1991	588		Engel et al., 1986	
Methergine	24.5	Hoyer and Schoeffter, 1991	501	Van Wijngaarden et al., 1990	

TABLE 6—Continued

Drug	Affinity	Reference	Drug	Affinity	Reference
MK 212	691	Engel et al., 1986	Amitriptyline	5,800	Peroutka, 1986
(±)Propranolol	588	Schlicker et al., 1989	Spiroxitrine	7,943	Hoyer and Schoeffter, 1991
	736	Yagaloff and Hartig, 1985	Buspirone	2,940	Sanders-Bush, 1988b
	1,200	Sanders-Bush, 1988b		7,940	Hoyer and Schoeffter, 1991
NAN-190	602	Van Wijngaarden et al., 1990		110,000	Peroutka, 1986
PAPP	602	Schoeffter and Hoyer, 1989b	Sumatriptan	8,128	Van Wijngaarden et al., 1990
5-CT	630	Hoyer and Schoeffter, 1991	(±)ICYP	9,770	Hoyer et al., 1985b
Phentolamine	891	Hoyer and Schoeffter, 1991	Ipsapirone	9,000	Peroutka, 1986
	1,250	Yagaloff and Hartig, 1985		31,622	Hoyer and Schoeffter, 1991
	Very low affinity ($K_1 > 1,000$ nM)		SDZ(+)-21009	8,912	Engel et al., 1986
Sipiperone	1,000–1,995	Pazos et al., 1985b		10,000	Hoyer and Schoeffter, 1991
	1,150–1,258	Hoyer et al., 1985b; Hoyer and Schoeffter, 1991	CGS 12066B	12,882	Schlicker et al., 1989
	1,148	Engel et al., 1986	Gepirone	25,118	Hoyer and Schoeffter, 1991
	4,800	Peroutka, 1986		26,200	Sanders-Bush, 1988b
SDZ(-)-21009	1,995	Pazos et al., 1985b	(+)Pindolol	39,810	Hoyer et al., 1985b
	5,011	Hoyer and Schoeffter, 1991		65,600	Hoyer and Schoeffter, 1991
Haloperidol	4,600	Sanders-Bush, 1988b	Yohimbine	39,810	Hoyer and Schoeffter, 1991
8-OH-DPAT	5,011	Pazos et al., 1985b		42,657	Schlicker, 1989
	7,200	Peroutka, 1986, Hoyer and Schoeffter, 1991	(±)CYP	>10,000	Schlicker et al., 1989
	7,700	Yagaloff and Hartig, 1985	<i>l</i> -LSD	>5,000	Yagaloff and Hartig, 1985
	16,500	Sanders-Bush, 1988b		>10,000	Peroutka, 1986
			Urapidil	15,848	Hoyer and Schoeffter, 1991

et al., 1988a; Bolanos et al., 1990) possess these receptors at high density, i.e., about 50 times that found in brain tissues.

C. Molecular Structure

The 5-HT₃ receptor is a ligand-gated ion channel, which, when activated, causes fast, depolarizing responses, similar to those seen following nicotinic ACh receptor activation (Changeux et al., 1987). Recently, Maricq et al. (1991), using expression cloning, isolated a cDNA clone encoding a 5-HT₃ receptor from NCB-20 cells. They injected size-fractionated polyadenylated mRNA from NCB-20 cells into *Xenopus* oocytes and tested for the presence of serotonin-gated currents that could be blocked by selective 5-HT₃ antagonists. They then constructed a cDNA library from the positive mRNA fraction; the RNA transcripts were injected separately into *Xenopus* oocytes to select those producing the 5-HT₃ functional receptor.

The sequence analysis revealed that this receptor is a new member of the ligand-gated ion channel superfamily. The predicted protein corresponds to 487 amino acids and contains four hydrophobic transmembrane regions

(M1 to M4), a large cytoplasmic loop connecting M3 to M4, and a large amino-terminal domain containing the Cys-Cys loop, a signature of this family, thought to participate in the formation of a disulfide bond.

The pharmacological and electrophysiological characteristics of the cloned receptor are largely consistent with the properties of native 5-HT₃ receptors. The mRNA encoding this receptor is identified in brain, spinal cord, and heart but not in cerebellum, liver, spleen, or intestine; the absence of signal in the intestine suggests that the intestinal 5-HT₃ receptor is a distinct subtype encoded by a separate gene.

The receptor protein was previously solubilized by Miquel et al. (1990) and McKernan et al. (1990).

D. Transduction System

The 5-HT₃ receptor appears to be directly linked to the opening and closing of an ionic channel that is not selective for monovalent cations (K⁺ and Na⁺) and which is independent of a G protein. Its activation induces a rapid depolarization. Derkach et al. (1989), using a preparation of peripheral nerves (submucous plexus), were the first to demonstrate this electrophysiological func-

TABLE 7
Affinity values of drugs for 5-HT₁ recognition sites in mammalian brain membrane or cellular cultures

Drug	Affinity	Reference	Drug	Affinity	Reference
Very high affinity ($K_i < 10$ nM)			Low affinity ($K_i = 100-1,000$ nM)		
GR 67330	0.038	Kilpatrick et al., 1989	Mianserin	5.8	Nelson and Thomas, 1989
SDZ 206-830	0.158	Hoyer and Neijt, 1988		64.5	Hoyer and Neijt, 1988
	0.93	Kilpatrick et al., 1990b		223	Bolanos et al., 1990
QICS 205-930	0.45	Watling et al., 1988	SDZ 210-205	33.1	Hoyer and Neijt, 1988
BRL 24682	0.331	Nelson and Thomas, 1989	Bufotenine	33.8	Milburn and Peroutka, 1989
LY 278-584	0.575	Milburn and Peroutka, 1989	Pitozifen	42	Schmidt and Peroutka, 1990
	0.71	Schmidt and Peroutka, 1990	Clozapine	49	Schmidt and Peroutka, 1990
LY 258-458	0.75	Milburn and Peroutka, 1989		204	Bolanos et al., 1990
	0.94	Schmidt and Peroutka, 1990	Low affinity ($K_i = 100-1,000$ nM)		
Zacopride	0.1	Nelson and Thomas, 1989	Metoclopramide	120	Milburn and Peroutka, 1989
	0.27	Kilpatrick et al., 1989		160	Nelson and Thomas, 1989
	0.32	Hoyer and Neijt, 1988		223	Hoyer and Thomas, 1988
	0.42	Milburn and Peroutka, 1989		323	Barnes et al., 1989b
	0.99	Kilpatrick et al., 1990b		338	Bolanos et al., 1990
	1.9	Barnes et al., 1989b		363	Kilpatrick et al., 1987
S(-)Zacopride	0.325	Bolanos et al., 1990		1,400	Kilpatrick et al., 1990b
Tropisetron	0.38	Milburn and Peroutka, 1989	Indalpine	120	Schmidt and Peroutka, 1990
(ICS 205-930)	0.40	Nelson and Thomas, 1989		1,202	Bolanos et al., 1990
	0.812	Hoyer and Neijt, 1988	Cizapride	134	Nelson and Thomas, 1989
	0.89	Kilpatrick et al., 1989		512	Bolanos et al., 1990
	1.9	Barnes et al., 1988a	Cyproheptadine	140	Schmidt and Peroutka, 1990
	3.09	Kilpatrick et al., 1987		263	Hoyer and Neijt, 1988
	3.23	Kilpatrick et al., 1990b	2-Methyl-5HT	87	Milburn and Peroutka, 1989
Ondansetron	0.87	Nelson and Thomas, 1989		210	Kilpatrick et al., 1990b
(GR 38032)	1.62	Kilpatrick et al., 1989		426	Bolanos et al., 1990
	2.88	Kilpatrick et al., 1987		620	Nelson and Thomas, 1989
	4.78	Barnes et al., 1988a		954	Hoyer and Neijt, 1987
	6.02	Bolanos et al., 1990		977	Neijt et al., 1988a
	13.48	Hoyer and Neijt, 1988		1,122	Barnes et al., 1988a
RP 62203	0.26	Doble, 1990		1,445	Van Wijngaarden et al., 1990
SDZ 206-792	0.724	Hoyer and Neijt, 1988	Amitriptyline	260	Schmidt and Peroutka, 1990
Granisetron	0.26	Nelson and Thomas, 1989	LY 278-989	330	Schmidt and Peroutka, 1990
(BRL 43694)	0.38	Milburn and Peroutka, 1989	Imipramine	570	Schmidt and Peroutka, 1990
	0.588	Kilpatrick et al., 1987	ICI 169-369	680	Schmidt and Peroutka, 1990
	1.17	Kilpatrick et al., 1989		990	Nelson and Thomas, 1989
	1.41	Hoyer and Neijt, 1988	5-HT	130	Kilpatrick et al., 1990b
	2.69	Barnes et al., 1988a		160	Nelson and Thomas, 1989
	4.17	Kilpatrick et al., 1990b		363	Van Wijngaarden et al., 1990
Quipazine	0.22	Kilpatrick et al., 1989		380	Bolanos et al., 1990
	0.501	Milburn and Peroutka, 1989		645	Barnes et al., 1988a
	1.23	Nelson and Thomas, 1989		891	Hoyer and Neijt, 1987
	2.04	Hoyer and Neijt, 1988	Phenylbiguanide	34	Milburn and Peroutka, 1989
GR 65630	1.34	Barnes et al., 1989b		130	Kilpatrick et al., 1990b
	1.58	Kilpatrick et al., 1987		446	Bolanos et al., 1990
BRL 47204	1.07	Nelson and Thomas, 1989		1,348	Hoyer and Neijt, 1987
R(+)-Zacopride	2.29	Bolanos et al., 1990		1,819	Neijt et al., 1988a
BRL 24924	3.16	Hoyer and Neijt, 1988	TFMPP	160	Schmidt and Peroutka, 1990
	23.9	Kilpatrick et al., 1990b		2,089	Van Wijngaarden et al., 1990
SDZ 210-204	5.01	Hoyer and Neijt, 1988	5,7-DHT	190	Schmidt and Peroutka, 1990
	7.8	Neijt et al., 1988a	Very low affinity ($K_i > 1,000$ nM)		
MDL 72222	5.3	Nelson and Thomas, 1989	NAN-190	1,202	Van Wijngaarden et al., 1990
	6.3	Hoyer and Neijt, 1988; Milburn, 1989	Cocaine	1,513	Milburn and Peroutka, 1989
	37.15	Bolanos et al., 1990		3,311	Barnes et al., 1989b
	42.1	Kilpatrick et al., 1990b		3,630	Kilpatrick et al., 1987
	54.98	Kilpatrick et al., 1987		12,400	Kilpatrick et al., 1990b
LY 211-000	1.9	Schmidt and Peroutka, 1990	α -Methyl-5-HT	1,600	Kilpatrick et al., 1990b
	4.2	Milburn and Peroutka, 1989	5-MeO-T	1,900	Schmidt and Peroutka, 1990
High affinity ($K_i = 10-100$ nM)			Mepyramine	2,400	Nelson and Thomas, 1989
mCPP	10	Milburn and Peroutka, 1989	Methiothepin	3,000	Nelson and Thomas, 1989
	40.2	Kilpatrick et al., 1990b		9,700	Kilpatrick et al., 1990b
	48.9	Bolanos et al., 1990	RU 24969	3,800	Van Wijngaarden et al., 1990
	54.9	Kilpatrick et al., 1987	Ritanserin	5,300	Nelson and Thomas, 1989
	102	Hoyer and Neijt, 1988		7,244	Van Wijngaarden et al., 1990
MK 212	12	Schmidt and Peroutka, 1990	Metergoline	7,400	Nelson and Thomas, 1989
			8-OH-DPAT	12,589	Bolanos et al., 1990
				17,200	Kilpatrick et al., 1990b
			Methysergide	19,500	Kilpatrick et al., 1990b

tion. Numerous studies have since been performed on peripheral tissue preparations (see review by Andrade and Chaput, 1991a; Goldfarb, 1990) and neurons or cell cultures (Yakel and Jackson, 1988; Lambert et al., 1989; Neijt et al., 1988b; Peters and Lambert, 1989). The observed electrophysiological responses consist of a very rapid (100 to 300 ms) opening of the channel, leading to an increase in cationic conductance. This response, in contrast to 5-HT₁ or 5-HT₂ G protein-related responses, is rapidly desensitized; it is not sensitive to GTP or to pertussis toxin.

Interestingly, 5-HT₃ receptors are present on lymphocytes, where they regulate a voltage-gated conductance, accelerating the inactivation process of the cell (Choquet and Korn, 1988).

Recent experimental results indicate that a 5-HT₃-like receptor may be coupled to a G protein. In cingulate and entorhinal cortex, brain areas rich in 5-HT₃ receptors, the IP₃ production is stimulated by 5-HT₃ agonists and inhibited by 5-HT₃ antagonists (Ashby et al., 1990; Edwards et al., 1991a,b). Phenylbiguanide and 2-methyl-5-HT (Ashby et al., 1991) induce a current-dependent suppression of the firing rate in medial prefrontal cortical cells; this phenomenon may be related to this G protein-linked molecular mechanism. Additionally, cyclic GMP production in 108 CC15 cells is stimulated by 5-HT₃ agonists (Reiser and Hamprecht, 1989). These findings suggest that subtypes other than the 5-HT₃ receptor described before also may exist but are structurally different from it, although they possess pharmacological similarities.

E. Regulation

The mechanism controlling the function of the 5-HT₃ receptor is probably one of conformational changes leading to a rapid desensitization (Derkach et al., 1989; Yakel and Jackson, 1988), as is usually observed for receptors linked directly to an ionic channel. Because these receptors were discovered only recently, there have been few studies of the mechanisms by which they are regulated (Kilpatrick et al., 1987). Chronic treatment of rats with lithium led to a decrease of the binding affinity in the hippocampus but not of the maximal number of 5-HT₃ sites; imipramine treatment, in contrast, did not alter these receptors (Mizuta and Segawa, 1988).

F. Cellular Functions

5-HT₃ receptors have been demonstrated to facilitate the electrically evoked release of [³H]5-HT from guinea pig frontal cortex and rat hypothalamic slices (Galzin and Langer, 1991); this effect was mimicked by 2-CH₃-5-HT and antagonized by ondansetron (100 nM) and ICS 205-930 (10 nM). Thus, these results strongly suggest the existence of presynaptic facilitatory autoreceptors of a 5-HT₃ type.

However, the main function of the 5-HT₃ receptors is the modulation of the neuronal release of other neuro-

transmitters. Initially, Feuerstein and Hertting (1986) showed that 5-HT stimulated the release of noradrenaline from hippocampal slices; this effect was antagonized by ICS 205-930 and MDL 72222, but only at high concentrations (10 μM and 1 μM respectively). Therefore, the actual involvement of 5-HT₃ receptors in the release of noradrenaline is not firmly established. Blandina et al. (1991) obtained contradictory results. Using rat hypothalamic slices, they demonstrated that 5-HT₃ receptors inhibit noradrenaline release.

5-HT₃ receptors control dopamine release. In accumbens and amygdala, 5-HT₃ agonists stimulate the release of dopamine as shown in vivo (Costall et al., 1987b,c,d; Hagan et al., 1990) or by microdialysis (Imperato and Angelucci, 1989; Carboni et al., 1989a). A similar effect occurs in the striatum, as shown in experiments on superfused tissue slices (Blandina et al., 1988, 1989). Yi et al. (1991) proposed that the stimulatory effect of 5-HT on dopamine release in rat striatum synaptosomes does not result from the activation of 5-HT₃ but rather occurs via the transport of 5-HT into the dopaminergic terminals. In any case, the 5-HT₃ control of dopaminergic activity has been proposed to explain the antipsychotic activities of 5-HT₃ antagonists (see review by Costall et al., 1990), because an increase in dopaminergic activity is thought to favor psychosis (Costall et al., 1987d; Tricklebank, 1989; Hagan et al., 1990; Abbott, 1990).

Barnes et al. (1989b) showed that 5-HT₃ receptors also mediate inhibition of K⁺-evoked ACh release from rat entorhinal cortex.

Finally, Alhaider et al. (1991) showed that the serotonin 5-HT₃ receptors appear to be involved in controlling the GABAergic system, i.e., they evoke the release of GABA in spinal cord. Furthermore, activation of the 5-HT₃ receptors increases the K⁺-evoked GABA release in rat caudate-putamen; ICS 205-930 and MDL 72222 antagonize this effect (Meyer et al., 1991). Moreover, 5-HT₃ receptors mediate the serotonergic activation of the GABAergic interneurons inhibiting the hippocampal CA1 pyramidal cells (Ropert and Guy, 1991). However, in guinea pig hippocampal synaptosomes, 5-HT₃ agonists inhibit K⁺-evoked GABA release, and this inhibition is blocked by 5-HT₃ antagonists (Cloez et al., 1990). These findings suggest that 5-HT₃ receptors can control GABA release through several different mechanisms.

G. 5-HT₃ Receptors and Behavior

Under normal conditions, 5-HT₃ antagonists appear to have very few effects; this would favor their therapeutic use. The main therapeutic interest of 5-HT₃ antagonists resides in their activity as antiemetics, an action especially useful in conjunction with cancer therapies involving cytotoxic drug treatments (cisplatin, doxorubicin, or cyclophosphamide) or radiation. The antiemetic effect has been demonstrated in several experimental

models: ferret (Miner and Sanger, 1986; Miner et al., 1986, 1987; Costall et al., 1986, 1987a; Gylys et al., 1988; Fukuda et al., 1991) and cat and dogs (Gylys et al., 1988; Smith et al., 1988). Clinical studies have also clearly established that 5-HT₃ antagonists are antiemetic in humans (Leibundgut and Lancranjan, 1987; Cunningham et al., 1987, 1989; Carmichael et al., 1988; Marty et al., 1989; Miln and Heel, 1991; Jones et al., 1991; Smith et al., 1991).

Recently, ondansetron also has been proposed as a safe and effective antiemetic for the treatment of postoperative nausea and vomiting after general anesthesia (Larjani et al., 1991). The 5-HT₃ antagonists (ICS 205-930, MDL 72222, zacopride, and ondansetron) are not active in apomorphine-induced emesis (Miner et al., 1987) and are inefficient in "motion-sickness" in cats (Lucot, 1989) or in humans (Stott et al., 1989); motion-sickness is prevented by 5-HT_{1A} agonists in the cat (Lucot and Crampton, 1989). These results suggest that emesis may be provoked by several distinct mechanisms.

The 5-HT₃-related emesis presumably originates from a central effect involving the area postrema (Higgins et al., 1989), but it may also partly involve a peripheral effect. Indeed, both cytotoxic drugs and radiation disrupt the gastrointestinal mucosa, inducing the release of 5-HT from the enterochromaffin cells; this 5-HT stimulates the 5-HT₃ receptors located on vagal fibers which, in turn, may initiate emesis through a mechanism involving the area postrema and the nucleus of the solitarius tract (see review by Andrews et al. 1988; Aapro, 1991).

The first indication that the 5-HT₃ receptors are involved in pain was proposed in a study of inflammatory-related nociception (Eschaliere et al., 1989). Antinociceptive effects of intrathecal and intracerebroventricular administration of 5-HT or 2-methyl-5-HT, and antagonized by MDL 72222 or ICS 205-930, were reported for the rat (Glaum et al., 1988; Giordano, 1991; Alhaider et al., 1991). Similarly, 4-amino-5-chloro-N-[2-pyrrolidylmethyl]-2,3-dihydrobenzo[b]furan-7-carboxamide, a novel 5-HT₃ antagonist, produced significant analgesia in the formalin-induced inflammatory pain test in rats (Sufka and Giordano, 1991). These findings support the hypothesis that 5-HT₃ has a role in the spinal transmission of nociceptive impulses, probably by controlling GABA release (Alhaider et al., 1991).

5-HT₃ antagonists may also be useful in the treatment of migraine (Loisy et al., 1985), but this application has still to be established.

5-HT₃ receptors do not play an important role in feeding behavior as do 5-HT_{1A} (Hutson et al., 1988) and 5-HT_{1C} receptors (Kennett and Curzon, 1988). Nevertheless, ICS 205-930 has been shown to increase gastric emptying in rats and guinea pigs (Buchheit et al., 1985), and BRL 24924 blocks the inhibitory effect of 5-HT on the gastric secretion of peptides (Johansen and Bech,

1991). In the dog, on the other hand, 5-HT₃ antagonists do not display these properties (Gullikson et al., 1991).

A great deal of interest is developing in the potential activities of the 5-HT₃ receptors in psychiatry. The 5-HT₃ receptors may play a role in tolerance-dependence mechanisms. 5-HT₃ antagonists decrease or inhibit the effects of withdrawal following the chronic use of nicotine, cocaine, ethanol, and benzodiazepines (Costall et al., 1988a, 1989a,b,c; Carboni et al., 1989b; Goudie and Leathley, 1990) and decrease ethanol consumption (Sellers et al., 1988; Oakley et al., 1988). However, effects on withdrawal symptoms have not been found consistently (Mos et al., 1989; Carboni et al., 1989a). Therefore, although the decrease in dopaminergic activity mediated by 5-HT₃ antagonists favors their use as antipsychotic drugs, additional studies are needed to assess whether they can be developed as therapeutic tools in that domain.

5-HT₃ receptors may play an interesting role in the neurochemical processes involved in anxiety. Indeed, low doses of ondansetron, granisetron, tropisetron, MDL 72222, and zacopride increase social interaction in rats under aversive conditions (Tyers et al., 1987; Tyers, 1989; Jones et al., 1988; Piper et al., 1988; Costall et al., 1988a, 1989b; Young and Johnson, 1991). In primates, these drugs reduce aggressiveness directed against threatening stimuli (Tyers et al., 1987; Jones et al., 1988; Piper et al., 1988; Costall et al., 1988a). However, these anxiolytic properties are not evidenced in all experimental tests, e.g., Vogel conflict test (Jones et al., 1988; Tyers et al., 1987), conditioned suppression in pigeon (Ahlers and Barrett, 1988), punished exploratory behavior (Mos et al., 1989), and elevated X-maze (Johnsson and File, 1988). Moreover, ondansetron and granisetron at high doses exhibit anxiogenic properties (Costall et al., 1988a).

These results and the fact that 5-HT₃ antagonists have a delay of onset of action longer than that of the benzodiazepines indicate that these drugs operate by different modes of action. In an attempt to determine the neuroanatomical site of action for the disinhibitory, perhaps anxiolytic, properties of 5-HT₃ receptor antagonists, Higgins et al. (1991a) examined the effects of such compounds in the social interaction test following injection into amygdala or dorsal raphe nucleus of the rat. Their results suggest that the amygdala, rather than the dorsal raphe nucleus, is involved in the effects of the 5-HT₃ antagonists. The anxiolytic therapeutic properties of 5-HT₃ antagonists have still to be confirmed.

Finally, interesting properties have been revealed in experimental models of cognition. For example, ondansetron and granisetron, unlike the benzodiazepines, increase "learning" in rodents and primates (Barnes et al., 1990b; Chugh et al., 1991). These properties might be exploited for a particular therapeutic application, such as in the treatment of Alzheimer's disease.

VI. Other 5-HT Receptors

Several recent pharmacological studies have led to the proposal that, in addition to 5-HT₁, 5-HT₂, and 5-HT₃ receptors, a 5-HT₄ receptor may also exist. Although it was present in several tissues where it had different functions, the absence of labeling of its binding sites did not allow a final classification as a fully recognized receptor (see review by Bockaert et al., 1992).

Dumuis et al. (1988b) obtained results with neuronal primary cell cultures of mouse embryo colliculi suggesting the existence of a new type of receptor that is positively coupled to adenylyl cyclase activity (table 8). They showed that 5-HT ($K_m = 109$ nM) and 5-MeO-T increase cAMP production, whereas tryptamine, bufotenin, and 2-methyl-5-HT are partial agonists and 8-OH-DPAT and RU 24969 are inefficient. Among the serotonergic antagonists, only ICS 205-930 (tropisetron) competitively inhibits 5-HT-induced cAMP production. This effect was observed at high concentrations of ICS 205-930 ($K_i = 710$ nM; 100 times greater than that required to inhibit 5-HT₃ receptor activity), suggesting that the observed phenomenon is not related to 5-HT₃ receptor function (Dumuis et al., 1988a). Moreover, MDL 72222 is a very weak antagonist of this effect ($K_i > 5000$ nM), whereas it is a potent 5-HT₃ antagonist. In addition, 5-HT₁ and 5-HT₂ antagonists are inefficient in inhibiting cAMP production. Dumuis et al. (1988b) proposed that the receptor exerting this effect was the same as that previously identified in guinea pig hippocampus (R_L) that

has a low affinity for 5-CT and that increases cAMP production (Shenker et al., 1987). They suggested calling the receptor 5-HT₄.

More recent findings established three families of compounds as 5-HT₄ agonists: indolamine derivatives (5-HT, 5-CT, 5-MeOT, bufotenin, 5-MeO-N-N-DMT), benzamides (metoclopramide, BRL 24924, BRL 20967, cisapride, and zacopride) (Dumuis et al., 1989a,b; Bockaert et al., 1990), and azabicycloalkyl benzimidazolone (BIMU 1, BIMU 8) (Dumuis et al., 1991). Recently, DAU 6285 (Dumuis et al., 1992) and SDZ 205-557 (Buchheit et al., 1991, 1992) were proposed as selective antagonists for 5-HT₄ receptors. These compounds, having a lower affinity for 5-HT₃ receptors, may represent a new interesting tool for investigating the pharmacological and physiological properties of 5-HT₄ receptors. So far, the recognition sites for these receptors have not been identified.

5-HT₄ receptors appear to mediate slow excitatory responses to 5-HT in the brain, because they elicit a slow membrane depolarization and a decrease in the calcium-activated afterhyperpolarization in rat hippocampal pyramidal neurons (Andrade and Chaput, 1991b). Inhibition of K⁺ channels also was mediated by 5-HT₄ receptors; it was demonstrated using individual collicular patch-clamped neurons (see Bockaert et al., 1992). The mechanism by which these receptors inhibit K⁺ channels seems to involve cAMP production and consequent activation of cAMP-dependent protein kinase A.

TABLE 8

Affinity of agonists at 5-HT₄ receptors positively coupled to adenylyl cyclase in mouse embryo colliculi neurons in primary culture and in adult guinea pig hippocampal membranes

Agonistic drugs	Colliculi neuron EC ₅₀ (nM)		Hippocampal membrane EC ₅₀ (nM)	
Cisapride	72.44	(Dumuis et al., 1989b)	210	(Bockaert et al., 1990)
BIMU 8	72	(Dumuis et al., 1991)		
5-HT	92	(Dumuis et al., 1989a)		
	106	(Dumuis et al., 1991)	263	(Dumuis et al., 1988b)
	109	(Dumuis et al., 1988b)		
5-MeO-T	100	(Dumuis et al., 1988b)		
BRL 24924	116	(Dumuis et al., 1989a)	33	(Bockaert et al., 1990)
	125	(Dumuis et al., 1989b)		
BIMU 1	360	(Dumuis et al., 1991)		
RU 28253	560	(Dumuis et al., 1988b)	3,500	(Dumuis et al., 1988b)
Zacopride	1,122	(Dumuis et al., 1989b)		
Bufotenine	1,580	(Dumuis et al., 1988b)	29,500	(Dumuis et al., 1988b)
5-CT	3,160	(Dumuis et al., 1988b)		
BRL 20627	3,235	(Dumuis et al., 1989b)		
Metoclopramide	4,570	(Dumuis et al., 1989a)		
5-MeO-N,N-DMT	17,780	(Dumuis et al., 1988b)		
Tryptamine	>10,000	(Dumuis et al., 1988b)		
d-LSD	>10,000	(Dumuis et al., 1988b)		
2-CH ₃ -5-HT	>10,000	(Dumuis et al., 1988b)	Inactive	(Dumuis et al., 1988b)
8-OH-DPAT	Inactive	(Dumuis et al., 1988b)	Inactive	(Dumuis et al., 1988b)
Ipsapirone	Inactive	(Dumuis et al., 1988b)		
Buspirone	Inactive	(Dumuis et al., 1988b)		
Metergoline	Inactive	(Dumuis et al., 1988b)		
Methysergide	Inactive	(Dumuis et al., 1988b)		
l-Phenylbiguanide	Inactive	(Dumuis et al., 1988b)		
RU 24969	Inactive	(Dumuis et al., 1988b)		

These receptors are also present at the periphery where they control the contraction of guinea pig ileum (Craig and Clarke, 1990) and ascending colon (Elswood et al., 1991) and relaxation of esophageal tunica muscularis mucosae in rat (Baxter et al., 1991) and humans (Kaumann, 1990; Kaumann et al., 1991). It is generally believed that the stimulation of the 5-HT₄ receptor in the ileum causes the release of ACh, because atropine attenuates the contraction elicited by 5-MeO-T (Fozard, 1990) and by low concentrations of 5-HT (Eglen et al., 1990). Stimulation of 5-HT₄ receptors by 5-MeO-T also causes an increase in the electrically evoked twitch contraction of the guinea pig ileum due to an increase in the evoked ACh release (Craig and Clarke, 1990; Kilbinger and Wolf, 1992). Moreover, this receptor is involved in the serotonin-induced cardiac effects in both porcine and human heart (Villalon et al., 1991, Ouadid et al., 1991).

5-HT₄ receptors might represent a functionally important family of serotonergic receptors; however, further experimentation is needed to determine whether this is so. Furthermore, additional work should be done to elucidate the biochemical, molecular, and cellular characteristics of this receptor.

VII. Conclusion

The explosive development of research in the serotonergic system led to the discovery that different 5-HT receptors are involved in numerous physiopathological functions mediated by serotonergic neurotransmission. Initially, the receptors were often suspected of being artifactual because the existence of multiple receptors for a single neurotransmitter was considered unrealistic. The results of recent studies using molecular biological techniques clearly confirm the reality of the existence of several types of serotonergic receptors. The characterization of their molecular structure has confirmed the initial classification of Bradley et al. (1986) and has demonstrated subclasses of receptors that are coupled to G proteins as well as those that function as an ionic channel. These observations now constitute a general framework in which new receptor subtypes can be inserted.

That multiple receptors exist for a single neurotransmitter constitutes an important adaptive advantage. Indeed, the multiplicity of the 5-HT receptors allows differential interactions of 5-HT with biological tissues. The variety of the effects produced depends on the presence and the number of a given receptor subtype in the considered tissue, their cellular localization, i.e., pre- or postsynaptic, their pharmacological specificity, the nature of their transduction system, and the expression of the gene coding for the particular receptor subtype.

The serotonergic cerebral system is both highly localized, the neuronal cell bodies being all contained in the raphe nuclei, and ubiquitous because the serotonergic fibers innervate all brain areas. These observations support the hypothesis that the serotonergic system may

control aspects of CNS function through the modulation of other neurotransmitter activities. The multiplicity of the serotonergic receptors also constitutes an important factor in the selectivity of the serotonergic interactions with other neurotransmitter systems.

The mode of action of the serotonergic system generally involves two mechanisms: one corresponding to a synaptic transmission; the other corresponding to a non-synaptic one, analogous to an endocrine-like neuromodulation. These two mechanisms have been described as "wiring transmission" and "volume transmission," respectively (Fuxe et al., 1989). They require the existence of receptor types having different affinities for the neurotransmitter, i.e., the nonsynaptic transmission can occur only via receptors having a high-affinity constant similar to that exhibited by 5-HT₁ receptors; however, the latter receptors can also be involved in the synaptic form of transmission.

Because multiple 5-HT receptors control various molecular and cellular activities, it is not surprising that the serotonergic system participates in a variety of quite different physiological events such as temperature regulation, food intake, and sexual behavior. Because dysfunctions in these activities will have important pathological consequences, it is important to know the fundamental mechanisms involved in the function of the serotonergic system and to define the pharmacological agents that interact with the activities linked to the stimulation or activation of the 5-HT receptors.

In the near future, it is expected that each of the serotonergic receptors will be defined on the basis of their molecular characteristics, thus providing access to a precise identification of the binding epitopes for both serotonin agonists and antagonists as well as to the molecular mechanisms involved in the activation of each receptor. A precise characterization of receptor properties will allow the development of potent, selective, and stereospecific ligands. Such studies should lead to the characterization of the molecular, cellular, and functional features of the receptors as well as the regulatory mechanisms that particularly affect their gene expression. It is hoped that new and more efficient treatments may result from such basic studies.

Addendum. During the editing of this manuscript, significant findings were reported; among the most important ones was the demonstration of the existence and cloning of 5-HT_{1E} receptors (McAllister et al., 1992) and of a new receptor subtype mediating inhibition of adenylyl cyclase (S31) (Levy et al., 1992).

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